

Apolipoprotein B-containing lipoproteins in atherogenesis

Jan Borén¹✉, Chris J. Packard² & Christoph J. Binder³

Abstract

Apolipoprotein B (apoB) is the main structural protein of LDLs, triglyceride-rich lipoproteins and lipoprotein(a), and is crucial for their formation, metabolism and atherogenic properties. In this Review, we present insights into the role of apoB-containing lipoproteins in atherogenesis, with an emphasis on the mechanisms leading to plaque initiation and growth. LDL, the most abundant cholesterol-rich lipoprotein in plasma, is causally linked to atherosclerosis. LDL enters the artery wall by transcytosis and, in vulnerable regions, is retained in the subendothelial space by binding to proteoglycans via specific sites on apoB. A maladaptive response ensues. This response involves modification of LDL particles, which promotes LDL retention and the release of bioactive lipid products that trigger inflammatory responses in vascular cells, as well as adaptive immune responses. Resident and recruited macrophages take up modified LDL, leading to foam cell formation and ultimately cell death due to inadequate cellular lipid handling. Accumulation of dead cells and cholesterol crystallization are hallmarks of the necrotic core of atherosclerotic plaques. Other apoB-containing lipoproteins, although less abundant, have substantially greater atherogenicity per particle than LDL. These lipoproteins probably contribute to atherogenesis in a similar way to LDL but might also induce additional pathogenic mechanisms. Several targets for intervention to reduce the rate of atherosclerotic lesion initiation and progression have now been identified, including lowering plasma lipoprotein levels and modulating the maladaptive responses in the artery wall.

Sections

Introduction

Initiation of atherosclerosis: role of LDL retention

Early stages of atherosclerosis: modification of retained LDL and cellular responses

Progression of atherosclerosis: cholesterol crystal formation, DAMPs and inflammatory response

Beyond LDL: relative atherogenicity of apoB-containing lipoproteins

Conclusions

¹Department of Molecular and Clinical Medicine, University of Gothenburg, Gothenburg, Sweden. ²Institute of Cardiovascular and Medical Sciences, University of Glasgow, Glasgow, UK. ³Department of Laboratory Medicine, Medical University of Vienna, Vienna, Austria. ✉e-mail: jan.boren@wlab.gu.se

Key points

- LDL is the main carrier of cholesterol in the blood and of circulating cholesterol into the artery wall.
- LDL is a proven causative factor for atherosclerotic cardiovascular disease, and reducing the plasma levels of LDL substantially reduces cardiovascular risk.
- Subendothelial retention of LDL and other apolipoprotein B-containing lipoproteins is the primary trigger for the development of atherosclerosis.
- Retained LDL becomes modified in the artery wall, and the focal accumulation of modified lipoproteins triggers the recruitment of monocytes and macrophages.
- Damage-associated molecular patterns, formed when retained LDL is modified, induce a maladaptive immune response.
- Different species of apolipoprotein B-containing lipoproteins are not equally atherogenic; triglyceride-rich lipoproteins and their remnants and lipoprotein(a) are markedly more atherogenic than LDL.

Introduction

Cardiovascular disease is the leading cause of death worldwide, causing more than 19 million deaths annually¹. Atherosclerosis is the major disease process underlying approximately 85% of these deaths¹. Elevated plasma concentrations of LDL are causally associated with atherosclerotic cardiovascular disease (ASCVD), and lowering LDL levels reduces ASCVD events in humans^{2,3}. Although the role of LDL as the prime causative agent of atherosclerosis has been questioned³, and competing hypotheses have been proposed to explain the early stages of atherogenesis, the accumulation of LDL and other apolipoprotein B (apoB)-containing lipoproteins in the intima of the artery wall is now established as the primary trigger for the development of atherosclerosis^{3–8}. These trapped lipoproteins and their by-products induce maladaptive local responses that lead to atherosclerotic plaque initiation, progression and maturation.

ApoB, the major structural apolipoprotein of LDL and other atherogenic lipoproteins, is required for their formation and metabolism. ApoB exists in two isoforms: apoB100, consisting of 4,536 amino acids, and apoB48, a truncated version containing the first 2,152 amino acids³. Both proteins are encoded by the *APOB* gene, from which a single mRNA is transcribed. ApoB48 is generated by RNA editing, which introduces a stop codon at residue 2,153. In humans, apoB100 is expressed in the liver and secreted on VLDL, whereas apoB48 is expressed in the intestine and found on chylomicrons³. As chylomicrons and VLDL circulate through the bloodstream, their triglycerides are hydrolysed by lipoprotein lipase located on the endothelial surface of blood vessels. Remnants are formed that are either cleared from the circulation by the liver or, in the case of VLDL, converted to LDL, the main carrier of cholesterol.

ApoB differs from other apolipoproteins in that it does not transfer between different lipoproteins³ owing to its size and the presence of antiparallel β -sheets that form robust lipid-binding structures^{9,10}. Given that all apoB-containing lipoproteins contain a single apoB100

protein, plasma apoB concentration is a direct measure of the number of circulating atherogenic lipoproteins^{2,3,11}. Although all apoB-containing lipoproteins are thought to contribute to the formation and progression of atherosclerotic plaques, the main focus to date has been on LDL because it is by far the most abundant atherogenic lipoprotein species^{2,3}. In this Review, we present new insights into the role of apoB-containing lipoproteins in atherogenesis, with an emphasis on the mechanisms that lead to atherosclerotic plaque initiation and growth. First, we describe in detail the causative role that LDL has in the early stages of atherogenesis, and then we widen the discussion to explore emerging concepts of the atherogenicity of other apoB-containing lipoproteins. A large amount of the evidence presented comes from animal models and in vitro studies, and care has to be exercised in translating the findings to human atherosclerosis. When possible, key experimental findings are linked to clinical observations.

Initiation of atherosclerosis: role of LDL retention Influx of LDL into the artery wall

To enter the artery wall, lipoproteins must cross the vascular endothelium. Genetic evidence has disproved the idea that LDL diffuses passively across the endothelium and instead shows that active transcytosis occurs (Fig. 1), mediated by a regulated vesicular pathway involving caveolae^{12–14}, scavenger receptor class B member 1 (SR-B1)¹⁵ and activin receptor-like kinase 1 (ref. 16). These results explain the studies showing that the greatest lipid accumulation and intimal thickening in rabbit aortas occurred in areas where the endothelium had regenerated after de-endothelialization, rather than in adjacent denuded areas (that is, areas of the intima without an endothelial lining)^{17,18}. SR-B1 seems to have a key role in LDL transcytosis by interacting with the guanine nucleotide exchange factor dedicator of cytokinesis protein 4 (DOCK4)^{19,20}. Interestingly, expression of SR-B1 and DOCK4 is higher in human atherosclerotic arteries than in unaffected arteries²⁰. Oestrogens inhibit LDL transcytosis by reducing endothelial SR-B1 levels through the G protein-coupled oestrogen receptor^{21,22}. This observation might help to explain the lower risk of ASCVD in premenopausal women²³. Conversely, factors such as activation of the NACHT, LRR and PYD domain-containing protein 3 (NLRP3) inflammasome or hyperglycaemia increase LDL transcytosis^{24,25}, which might contribute, at least in part, to the accelerated atherosclerosis observed in people with type 2 diabetes mellitus²⁶. Studies have shown that rapid correction of hypercholesterolaemia in mice improves endothelial barrier function, limiting LDL ingress²⁷. However, the underlying mechanisms are unclear and need to be validated in humans.

Endothelial glycocalyx. The luminal surface of all vascular endothelial cells is covered by the endothelial glycocalyx, which comprises membrane-bound, negatively charged proteoglycans, glycoproteins, glycolipids and glycosaminoglycans²⁸. The endothelial glycocalyx has been proposed to function as a regulator of LDL transendothelial transport; when through the glycocalyx, LDL can cross the endothelium via transcytosis. Studies on the coronary arteries of cholesterol-fed pigeons, and subsequent studies on the carotid arteries of hypercholesterolaemic mice, have shown that the thickness of the glycocalyx is reduced in the regions containing atherosclerotic plaque^{29–32}. However, experimental evidence is still lacking as to the wider pathogenic relevance of this phenomenon, and the exact nature of the interaction between LDL and the glycocalyx remains to be elucidated.

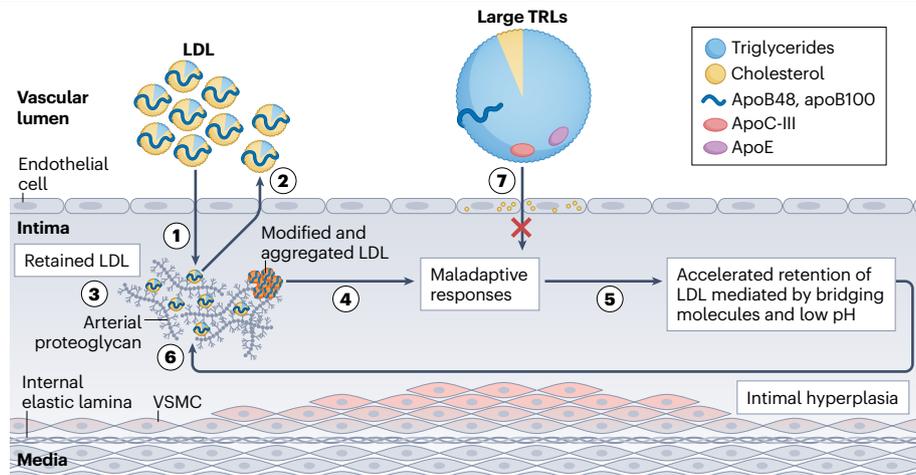


Fig. 1 | Early stages of atherosclerosis. (1) To enter the arterial wall, lipoproteins must cross the endothelium by transcytosis. The transport vesicles are approximately 100 nm in diameter, so transcytosis is restricted to lipoproteins <70 nm in diameter (LDL and smaller triglyceride-rich lipoprotein (TRL) remnants). (2) Most lipoproteins that enter the arterial wall flow back into the circulation. (3) However, a fraction of LDL is trapped in the arterial wall by binding to the extracellular matrix. This binding is mediated by ionic interactions between positively charged amino acid residues in apolipoprotein B-100 (apoB100) and negatively charged sulfate or glycosaminoglycan chains of artery wall proteoglycans. (4) Modification of the retained LDL by secretory sphingomyelinase (produced by the endothelium and atherosclerotic plaque macrophages) and phospholipases leads to loss of LDL stability and aggregation. (5) The modified LDL triggers maladaptive responses that accelerate

atherosclerotic lesion development. For example, activated macrophages release bridging molecules, including lipoprotein lipase, which facilitate accelerated binding of LDL to the artery extracellular matrix. The inflammatory process also leads to local acidification of extracellular fluids, which protonate histidine residues, increasing the net positive charge of the LDL particle. (6) Activated macrophages also stimulate transformation of vascular smooth muscle cells (VSMCs) from a contractile to a proliferative state, resulting in increased synthesis of LDL-binding proteoglycans. (7) Although large TRLs (chylomicrons and VLDL₁) are too large to enter the artery wall by transcytosis, a novel mechanistic link has been revealed between large TRLs and vascular disease, mediated by the induction of lipid droplets in the endothelium^{191,193}. The mechanism seems to involve activation of nuclear factor- κ B and upregulation of vascular cell adhesion molecule, which promotes atherosclerosis and plaque inflammation^{191–193}.

Electrostatic interactions between apoB100 and artery wall proteoglycans

The interaction between LDL in the subendothelial space and artery wall components is mediated by electrostatic attraction between negatively charged sulfate or carbohydrate groups of artery wall proteoglycans and positively charged amino acid residues in apoB. In 1949, Faber reported on an association between intimal mucopolysaccharides and cholesterol deposition in the artery wall³³. This seminal work inspired further investigation, including the early studies of Camejo and colleagues^{8,34} that led to the identification of eight clusters of positively charged amino acids in apoB100 that bind to proteoglycans in human isolated artery^{35–38}.

To determine which of these clusters of positively charged amino acids are functional in intact lipoproteins, we generated transgenic mice expressing mutant forms of human apoB100 (ref. 39). We identified residues 3359 to 3369 (termed site B) in apoB100 as the primary proteoglycan-binding site and, in particular, the essential role of positively charged arginine and lysine residues in this site for the interaction between LDL and artery wall proteoglycans^{39,40}. Subsequent experiments in mice with mutations in site B confirmed the crucial link between LDL atherogenicity and the proteoglycan-binding activity of apoB100 (ref. 4), thereby demonstrating that vascular retention of apoB100-containing lipoproteins is an initiating event in atherosclerosis. Site B is located in the carboxy-terminal half of apoB100 and coincides with the LDL receptor-binding site⁴¹. Site B is, therefore, not present in apoB48. However, we also identified a proteoglycan-binding site in the amino terminus (residues 84 to 94) of apoB48 (ref. 42).

In apoB100, this site is masked by the C-terminal portion of apoB100 and is, therefore, not functional.

Subsequent studies confirmed the existence of a third functional binding site in apoB100, termed site A (residues 3148 to 3158)⁴³, which was originally proposed by Camejo and colleagues⁴⁴. In vitro, site A becomes functional when LDL is modified by secretory group IIA phospholipase A₂ (sPLA₂), which catalyses the hydrolysis of surface phospholipids on LDL, resulting in the formation of smaller and denser particles that have increased affinity for proteoglycans⁴⁵. Elevated plasma levels of sPLA₂ and another phospholipase, lipoprotein-associated phospholipase A₂ (Lp-PLA₂), have been identified as important risk factors for ASCVD^{46,47}. One might hypothesize that an association between elevated levels of small, dense LDL and an increased risk of ASCVD might be, at least partly, due to the exposure of site A on apoB100, secondary to the action of these enzymes^{48,49}. However, in clinical trials involving individuals at high risk of a cardiovascular event, treatment with varespladib (an inhibitor of sPLA₂) or darapladib (an inhibitor of Lp-PLA₂) was not associated with a reduced risk of ASCVD events compared with placebo^{50,51}. Similarly, in Mendelian randomization studies, gene variants that lowered Lp-PLA₂ activity were not associated with a decreased in ASCVD events⁵¹. These results indicate that sPLA₂ and Lp-PLA₂ are unlikely to be causal risk factors for ASCVD.

What is more important: the rate of LDL influx into the artery wall or subendothelial retention of LDL?

Early research by Schwenke and Carew in cholesterol-fed rabbits showed that whereas the rate of LDL ingress did not differ significantly

between areas of normal artery that were deemed either 'susceptible' or 'resistant' to subsequent atherosclerotic plaque development, the susceptible regions had greater LDL accumulation⁵². These findings suggest that mechanisms that facilitate selective retention of LDL determine the degree of local accumulation of the lipoprotein. Tran-Lundmark and colleagues confirmed this notion in molecular studies that involved crossing mice expressing heparan sulfate (HS)-deficient perlecan (also known as HSPG2) with hypercholesterolaemic *ApoE*^{-/-} mice⁵³. Compared with *ApoE*^{-/-} control mice, the influx of LDL into the aorta wall was increased, whereas subendothelial retention of LDL was significantly reduced in *ApoE*^{-/-} mice with HS-deficient HSPG2 (ref. 53). Importantly, atherosclerotic lesions were also markedly reduced in these *ApoE*^{-/-}, HS-deficient HSPG2 mice⁵³, underscoring the importance of LDL retention over the degree of endothelial permeability to LDL as the main pathogenic mechanism. Of note, only a small amount of LDL is retained in the artery wall by proteoglycan binding; most particles that enter subsequently leave without contributing to atherosclerotic lesion growth⁵³.

Determinants of LDL retention by artery wall proteoglycans

LDL composition. The protein composition of apoB-containing lipoproteins affects their interaction with artery wall proteoglycans. For example, apoB48-containing lipoproteins typically contain numerous apoE molecules, which have a proteoglycan-binding domain that is almost identical to that found in apoB100 (ref. 54). In addition, apoC-III, a small exchangeable apolipoprotein, increases the binding of LDL to proteoglycans while decreasing the hepatic uptake of apoB-containing lipoproteins, leading to increased lipoprotein accumulation in the vessel wall^{55,56}. The mechanisms underlying this observation remain unclear because apoC-III lacks the positively charged domains that are necessary for binding to artery wall proteoglycans^{55,56}. However, in vitro studies of LDL from individuals with type 2 diabetes have shown that an increased apoC-III to apoB molar ratio in LDL is associated with an altered lipid composition in LDL and increased proteoglycan binding⁵⁵. This study suggested that a more fluid monolayer on the LDL particle surface facilitates the acquisition of additional apoC-III molecules, and consequent conformational changes in apoB100 led to an increase in its affinity for proteoglycans⁵⁵.

Other changes in LDL quality might also affect its binding to artery wall proteoglycans. Changes in core lipid composition, such as increased cholesterol or triglyceride content⁵⁷, might induce conformational changes in apoB that affect its interaction with proteoglycans⁴³. Cholesterol enrichment of LDL correlates with increased affinity for proteoglycans⁴³. This phenomenon is likely to be due to a conformational change in site B in apoB100. Likewise, triglyceride-rich LDL is less atherogenic than cholesteryl ester-enriched LDL in *ApoE*^{-/-} mice⁵⁸. This result is in accordance with the findings of an earlier study showing a reciprocal relationship between the triglyceride content of LDL and the number of exposed lysine amino groups in apoB100 (ref. 59). In addition, LDL isolated from patients with type 1 diabetes was retained in the artery wall of diabetic mice to a greater extent than LDL from healthy control individuals⁶⁰. Although glycation of LDL is one potential modification that might increase retention in the artery wall, other as-yet-unidentified mechanisms might also be involved⁶⁰.

Artery wall constituents. Although most proteoglycans show affinity for LDL, proteoglycans with elongated glycosaminoglycan chains seem to have a crucial role in binding to LDL. Histology studies of human arteries have shown strong co-localization between LDL and

the proteoglycans biglycan, decorin and versican core protein^{61,62}. The composition of proteoglycans in arteries varies between animal species, and HSPG2 seems to be the primary proteoglycan that binds to lipoproteins in the mouse aorta⁵³.

Other components of the artery wall can also influence the subendothelial retention of LDL. For example, the expression of chondroitin sulfate proteoglycan 4 (also known as NG2) is increased in early atherosclerotic lesions in *ApoE*^{-/-} mice, and NG2-positive synthetic vascular smooth muscle cells (VSMCs) increase LDL binding and uptake by macrophages in the artery wall⁶³. Interestingly, NG2 glycosaminoglycan chains are not essential for LDL binding to NG2 (ref. 63), highlighting the unique ability of NG2 among LDL-binding proteoglycans to bind LDL through hydrophobic interactions.

We have shown that the cytokine tumour necrosis factor ligand superfamily member 13 (also known as APRIL) confers atheroprotection by binding to HS chains on HSPG2 and thereby limits LDL retention, macrophage accumulation and necrotic core formation⁶⁴. In light of these findings, interventions that increase the interaction between APRIL and HSPG2 could protect against atherosclerosis. Indeed, atherosclerosis was reduced in *ApoE*^{-/-} mice treated with an anti-APRIL antibody that increased the APRIL-HSPG2 interaction⁶⁴. Moreover, blocking sulfated glycosaminoglycans in the artery wall in *ApoE*^{-/-} mice with monoclonal antibodies inhibits the interaction between LDL and proteoglycans⁶⁵.

Regional intimal hyperplasia. The focal pattern of atherogenesis is striking, with lesions developing in the vicinity of artery branches and along the inner curvature of arteries, while other segments of the artery network remain relatively unaffected. The reason for the selective accumulation of LDL in these vulnerable areas is unclear. However, sites prone to develop atherosclerosis often have intimal hyperplasia^{62,66-70}, a thickening of the innermost layer of the vessel wall due to the accumulation of VSMCs and artery wall proteoglycans^{61,62} (Fig. 1). Interestingly, some vascular interventions (such as percutaneous coronary intervention and coronary artery bypass graft surgery) promote intimal hyperplasia and the accelerated retention of apoB-containing lipoproteins^{71,72}.

Findings by Nakashima and colleagues support the role of intimal hyperplasia in the pathogenesis of atherosclerosis⁶². The researchers examined lipid deposition and macrophage infiltration in the coronary arteries of people aged <40 years who had died from non-cardiovascular causes, and found that extracellular lipids accumulate in areas of diffuse intimal thickening⁶². The investigators also showed that the lipid accumulation precedes the infiltration of macrophages⁶². Further elegant studies by Bentzon and colleagues confirmed that LDL retention is the rate-limiting factor for LDL accumulation in the mouse aortic arch, and showed that LDL-retaining regions can be subdivided according to their capacity for continued LDL accumulation under hypercholesterolaemic conditions^{73,74}.

Lesion microenvironment. Atherosclerotic lesions are often hypoxic environments with elevated lactate levels and local acidification of extracellular fluids^{75,76}. We demonstrated a profound influence of local pH on the binding of LDL to artery wall proteoglycans, highlighting the crucial role of positively charged histidine residues in increasing LDL binding to human coronary arteries⁷⁷. Histidine residues remain uncharged at pH 7.4, but become protonated and positively charged at lower pH; the proportion of protonated histidine residues increases from approximately 4% at pH 7.4 to 25% at

pH 6.5 (ref. 77). Our results indicate that LDL binding can be inhibited at lower pH by blocking either site B or protonated histidine residues, suggesting that both components are required simultaneously for the increased affinity of LDL for artery wall proteoglycans at acidic pH⁷⁷. Although apoB100 contains 114 histidine residues, they are not clustered in sufficient numbers to independently form a glycosaminoglycan-binding site. However, histidine residues can contribute positively charged groups to lysine–arginine clusters. One such cluster, site A (residues 3148–3158), contains two histidine residues and five lysine or arginine residues within an 11-amino acid stretch⁷⁷. Consequently, site A might become functional at lower pH levels and, in this state, cooperates with site B to increase LDL binding to proteoglycans⁷⁷.

Before moving on to consider the next stage in the atherosclerotic disease process, it is worth noting that the large body of evidence summarized above supports the concept that lipid accumulation in the initial stages of atherosclerosis occurs independently of the presence of macrophages. Macrophages, either resident or from the circulation, congregate subsequently in lesions with substantial lipid deposition.

Early stages of atherosclerosis: modification of retained LDL and cellular responses

Modification of LDL retained in the artery wall

When retained in the subendothelial space, LDL is susceptible to modification by various proteases and lipases that alter its structure⁷⁸ and to oxidation by non-enzymatic and enzymatic mechanisms, involving lipoxygenases, myeloperoxidase, NADPH oxidases and nitric oxide synthases, among others⁷⁹ (Fig. 2). The expression of these enzymes and the production of reactive oxygen species by vascular cells and, more importantly, by macrophages contribute to LDL modification. Oxidation of LDL generates a myriad of different oxidized lipids that have the capacity to trigger vascular inflammation. Both cholesterol and phospholipids in LDL can undergo oxidative modification, giving rise to oxidized cholesteryl esters and various oxidized phospholipids (oxPLs), as well as their many degradation products. For example, oxidation of the abundant phospholipid 1-palmitoyl-2-arachidonoyl-*sn*-phosphatidylcholine results in the generation of highly reactive breakdown products of the *sn*-2 polyunsaturated fatty acids, including malondialdehyde and 4-hydroxynonenal, which in turn form adducts with the ϵ -amino groups of (generally) lysine residues of apoB or the amino groups of other phospholipids such as phosphatidylethanolamine⁸⁰. Similarly, the remaining core aldehyde carrying the phosphocholine head group can also modify apoB. Different types of oxPL product have been shown to activate endothelial cells and stimulate the expression of chemokines and adhesion molecules, promoting leukocyte recruitment to the artery wall⁸¹. In contrast to native LDL, oxidized LDL (oxLDL) is taken up by macrophages via the scavenger receptors CD36 and SR-A1, leading to foam cell formation⁸¹.

OxPLs associated with LDL particles can also be detected in the circulation by an immunoassay that measures the number of moles of oxPL per apoB100-containing particle⁸². These particles can either be released in an oxidized form from atherosclerotic plaques or be oxidized in the periphery. Increased plasma levels of oxPL–apoB have been found to be a marker of the extent of atherosclerosis and to improve risk prediction for myocardial infarction, ischaemic stroke and peripheral artery disease, independent of apoB or LDL cholesterol levels^{83–85}. Of note, oxPLs on lipoprotein(a) (Lp(a)) particles can also be assessed, as discussed below.

Mechanisms of accelerated subendothelial retention of LDL in atherosclerotic lesions

The presence of modified LDL that has been retained in the arterial wall triggers cellular responses that accelerate further LDL trapping. For example, oxLDL-induced release of cytokines and growth factors by inflammatory cells induces VSMCs to proliferate and produce proteoglycans with extended glycosaminoglycan chains, which have increased affinity for LDL^{86,87}. Furthermore, infiltrating macrophages secrete bridging molecules, in particular lipoprotein lipase, which increases lipoprotein retention because these molecules have binding sites for both proteoglycans and lipoproteins^{88–92}. In addition, macrophages release secretory sphingomyelinase (SMase), which increases lipoprotein retention and atherosclerosis in mouse models by promoting LDL aggregation^{93–97}. Aggregation of LDL in the artery wall increases the affinity of the lipoprotein for proteoglycans^{95,98} and accelerates the progression of atherosclerosis in humans⁹⁹. Aggregated LDL is readily taken up by macrophages and induces, in contrast to oxLDL, foam cell formation and mild mitochondrial dysfunction in human macrophages, without triggering oxidative stress or endoplasmic reticulum stress¹⁰⁰. Interestingly, macrophages engulf aggregated LDL and produce free cholesterol in an acidic, hydrolytic extracellular compartment known as the lysosomal synapse¹⁰¹. Through actin polymerization, macrophages create a tight seal around aggregated LDL, releasing lysosomal contents into this space in a process called digestive exophagy¹⁰¹.

The augmented binding of LDL to artery proteoglycans that occurs in established atherosclerotic lesions (mediated by bridging molecules, aggregation and low pH) might provide a mechanistic explanation for a puzzling clinical observation; namely, a lifetime plasma LDL cholesterol level of ≤ 2 mmol/l is associated with a very low risk of developing atherosclerosis¹⁰², whereas the same plasma level of LDL cholesterol is associated with atherosclerotic plaque progression in individuals with existing ASCVD who are receiving lipid-lowering treatment¹⁰³.

After LDL and other apoB-containing lipoproteins aggregate in the artery wall, their diffusion back into the plasma becomes nearly impossible owing to the substantial size of the aggregates and their greatly increased affinity as multivalent ligands for the arterial extracellular matrix. In addition, conformational changes occur that are likely to expose additional positively charged domains on apoB. Sneek and colleagues elucidated the mechanism behind SMase-induced aggregation of LDL by showing that SMase induces a substantial and widespread conformational change in apoB, exposing hydrophobic motifs that seem to stick digested LDL particles together¹⁰⁴. Interestingly, the susceptibility of plasma LDL to aggregation after exposure to SMase *ex vivo* varies considerably between individuals, predicts future death from ASCVD and can be altered by diet⁹⁹. This work adds to the growing body of evidence implicating SMase in human atherosclerosis^{79,93}.

Resident and recruited macrophages in atherosclerotic lesions

The focal accumulation of modified lipoproteins in vulnerable areas of artery walls leads to the migration of monocytes and macrophages into the area as part of the cellular response (Fig. 2). Single-cell analyses have identified substantial heterogeneity of macrophages in atherosclerotic plaques: ‘resident-like’, ‘lipid-associated’ and ‘inflammatory’ subtypes are all present^{105–107}. Aortic intima-resident macrophages, which are sustained by local proliferation and are found at sites of intimal thickening, are the first macrophage subtype to take up oxLDL and aggregated LDL¹⁰⁸. Indeed, early exposure to hypercholesterolaemia results in the depletion and reprogramming of resident macrophages in the intima

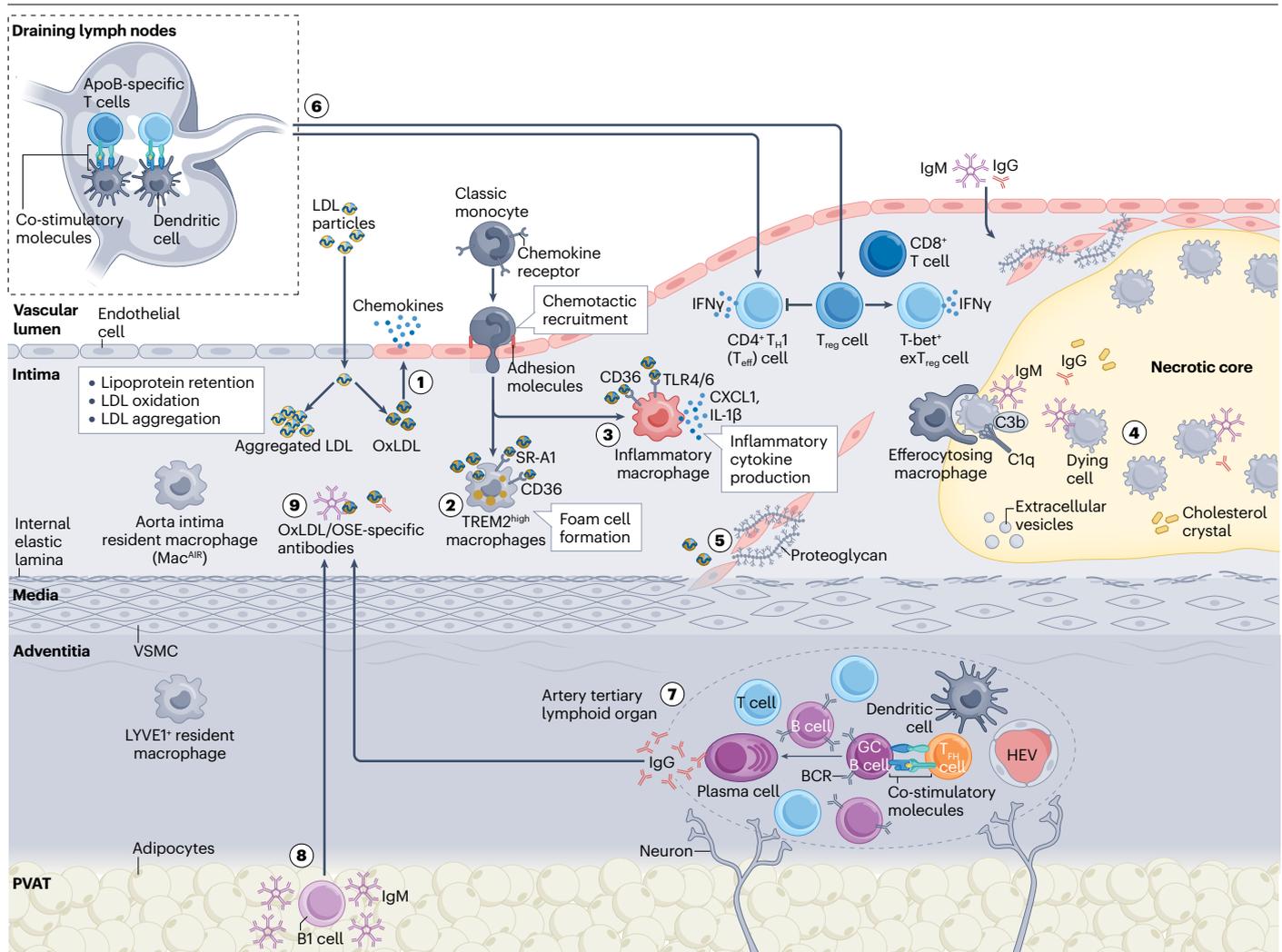


Fig. 2 | Cellular responses to LDL retention. (1) Retention of LDL exposes the lipoprotein to various modifications that alter the biological activity of LDL components, resulting in an array of cellular responses. Generation of oxidized LDL (oxLDL) leads to the formation of various oxidized lipid species that trigger inflammation. OxLDL and oxidized phospholipids in particular activate endothelial cells, inducing the expression of adhesion molecules and chemokines, which results in the recruitment of monocytes into the artery wall. (2) Resident and monocyte-derived macrophages take up oxidized and aggregated LDL, leading to the formation of foam cells. Aorta intima resident macrophages (Mac^{AIR}) and adventitial resident macrophages (LYVE1⁺) are the earliest to respond to hypercholesterolaemia, resulting in their depletion and reprogramming, respectively. During progression of atherosclerosis, TREM2^{high} macrophages make up the main foam cell subset and mediate protective homeostatic functions, including lipid handling and efferocytosis. (3) Inflammatory macrophages secrete pro-inflammatory cytokines and chemokines in response to damage-associated molecular patterns, such as oxidation-specific epitopes (OSEs) present on oxLDL. Cholesterol crystals taken up by or formed in macrophages trigger the activation of the NLRP3 inflammasome and the secretion of IL-1β and IL-18. (4) Impaired cellular cholesterol homeostasis results in cell death and the accumulation of dying cells, particularly during advanced disease stages

when efferocytosis is impaired. (5) OxLDL also stimulates vascular smooth muscle cells (VSMCs) to proliferate and produce proteoglycans but can also trigger VSMC death. (6) Both effector T (T_{eff}) cells and regulatory T (T_{reg}) cells are primed in secondary lymphoid organs and possibly in advanced artery tertiary lymphoid organs during advanced atherosclerosis. T_{eff} cells and T_{reg} cells are recruited to the intima where they mediate pro-atherogenic and anti-atherogenic effects, respectively. Some of these T cells have specificity for apolipoprotein B (apoB) peptides, and apoB-specific T_{reg} cells can convert to pro-inflammatory exT_{reg} cells during atherosclerotic disease progression. (7) Lymphoid organs, including artery tertiary lymphoid organs, also contain B cells that differentiate to plasma cells in germinal centre (GC) reactions to produce class-switched IgG antibodies against oxLDL or OSE. (8) By contrast, IgM against oxLDL or OSE are largely derived from B1 cells that are prominently found in perivascular adipose tissue (PVAT). (9) Antibodies, including those with specificity for oxLDL or OSE, are present in the intima of atherosclerotic lesions. IgM can inhibit the disease process by neutralizing oxLDL or extracellular vesicles and by increasing the clearance of dying cells by macrophages, whereas the function of IgG is less clear. BCR, B cell receptor; CX3CL1, CX3C-motif ligand 1; HEV, high endothelial venule; IFNγ, interferon-γ; SR-A1, scavenger receptor type A1; T_{FH}, T follicular helper; T_{H1}, T helper 1; TLR, Toll-like receptor.

and adventitia, respectively¹⁰⁹. Subsequently, newly recruited inflammatory monocytes arrive and give rise to several macrophage subsets, including those that scavenge modified LDL¹¹⁰. Accordingly, intermittent hyperlipidaemia in mice during youth is associated with subsequent rapid progression of atherosclerosis¹⁰⁹. This finding underscores the crucial need to manage abnormal plasma lipid levels effectively from an early age onwards.

Surprisingly, *in vitro* formation of lipid-loaded macrophages (foam cells) is linked to the suppression of inflammatory gene expression¹¹¹. In agreement, the gene expression profile of TREM2^{high} macrophages, which have been identified as a major foam cell subset, points to a function in lipid metabolism rather than inflammation¹¹². Indeed, our studies in mice demonstrated a protective function for TREM2⁺ macrophages in atherogenesis, by promoting efferocytosis and efficient lipid handling¹¹³. However, Monaco and colleagues identified PLIN2^{high}TREM1^{high} macrophages, which also express IL-1 β , as Toll-like receptor (TLR)-dependent inflammatory lipid-associated macrophages that might derive from anti-inflammatory TREM2^{high} foam cells¹¹⁴. The local inflammatory milieu in plaques could contribute to the transition of TREM2^{high} lipid-associated macrophages to inflammatory PLIN2^{high}TREM1^{high} macrophages.

To what extent other macrophage subsets, such as pro-inflammatory MHCII⁺ cytokine-expressing macrophages, cavity macrophages or interferon-inducible macrophages that have a characteristic type I interferon signature, respond to (oxidized) lipids is unknown¹¹⁵. Cellular lipid accumulation and an insufficient capacity to handle cholesterol lead to an increased rate of foam cell apoptosis, which results in the accumulation of apoptotic and necrotic cells due to defective efferocytosis^{116,117}. Indeed, atherogenic lipids have been shown to trigger CD36–TLR2-dependent apoptosis in endoplasmic reticulum-stressed macrophages *in vitro*¹¹⁸. The higher rate of cell death by several mechanisms leads to the release of intracellularly stored cholesterol and the formation of extracellular lipid deposits and acellular necrotic areas, which are characteristic of unstable, rupture-prone atherosclerotic plaques¹¹⁹.

Progression of atherosclerosis: cholesterol crystal formation, DAMPs and inflammatory response

Formation of extracellular lipid deposits and their role in atherogenesis

The extracellular lipid deposits in atherosclerotic plaques contain oxidized lipids associated with oxLDL, as well as cholesterol crystals, which can also be derived from lipoprotein particles that were not scavenged by leukocytes¹²⁰ (Fig. 2). Atherosclerotic plaque acidification has been shown to increase extracellular lipid accumulation¹²¹. Although the presence of large cholesterol crystals was documented in the earliest microscopic examinations of atherosclerotic plaques⁷⁵, the functional role of microcrystals of cholesterol in the inflammatory response was identified only much later. Indeed, cholesterol crystals are now considered key damage-associated molecular patterns (DAMPs) that drive the inflammatory processes in atherosclerosis.

Cholesterol crystals are potent activators of all three complement pathways, which modulate atherosclerotic lesion formation¹²². When taken up by macrophages or generated in lysosomes, cholesterol crystals also trigger the activation of the NLRP3 inflammasome, an intracellular pattern recognition receptor complex that promotes the generation and release of active IL-1 β and IL-18 (refs. 123,124). Deficiencies in several components of the NLRP3 signalling complex have mostly been shown to limit atherosclerotic lesion formation in mice¹²⁵,

and inhibition of IL-1 β with a monoclonal antibody has been shown to significantly reduce the occurrence of ASCVD events in patients with previous myocardial infarction compared with placebo¹²⁶. The FDA approved the use of a low dose of the broad anti-inflammatory agent colchicine for patients with established ASCVD, because this approach has been shown to reduce cardiovascular events compared with placebo in clinical trials¹²⁷. Moreover, the ongoing ZEUS trial¹²⁸ is testing the efficacy of ziltivekimab, an antibody targeting the pro-inflammatory cytokine IL-6, which is downstream of IL-1 β , for the prevention of cardiovascular events in patients with cardiovascular disease, chronic kidney disease and inflammation.

Adducts formed by phospholipid peroxidation products, which have been termed oxidation-specific epitopes (OSEs), also show robust immunoreactivity and are another class of DAMPs⁷⁹. Phosphocholine-containing oxPLs present in oxLDL are recognized by CD36, which cooperates with TLR4 and TLR6 in transmitting pro-inflammatory signals in macrophages, resulting in nuclear factor- κ B (NF- κ B) activation and the transcription of various chemokines, such as CXCL1, CXCL2, CCL9 and CCL5, as well as components of the NLRP3 inflammasome^{129,130}. Although studies using *in vitro* models of oxLDL need to be interpreted with care, specific active moieties of oxLDL, such as phosphocholine-containing oxPLs, have been clearly documented *in vivo*⁸². Indeed, the importance of phosphocholine-containing oxPLs in driving the inflammatory response during atherogenesis is underscored by experimental data by Witztum and colleagues demonstrating that Western diet-fed *Ldlr*^{-/-} mice overexpressing a single-chain variable fragment of the oxPL-neutralizing phosphocholine-specific antibody E06 have reduced levels of systemic inflammation and decreased atherosclerosis^{82,131}. In addition, malondialdehyde adducts also trigger chemokine secretion by macrophages, which is dependent on scavenger receptor-mediated uptake¹³².

The innate immune activation by lipid DAMPs not only promotes the secretion of downstream effector cytokines, such as IL-6, but also propagates vascular inflammation and results in the activation of adaptive immune responses and the subsequent recruitment of T cells, which have been shown to modulate atherosclerosis by participating in a maladaptive immune response¹³³ (Fig. 2).

Dendritic cells, T cells and B cells in the atherosclerotic plaque

In the adventitia of normal arteries, lymphocytes and dendritic cells (DCs) are an immune–vascular unit that mediates vascular homeostasis. Importantly, conventional DC subsets take up cholesterol-containing lipoproteins in the setting of dyslipidaemia, which results in their activation in an oxysterols receptor LXR β -dependent manner¹³⁴. Plasmacytoid and conventional DC subsets can sense vascular antigens and promote both tolerogenic and inflammatory responses, depending on the type of T cell they instruct^{133,135}. Whereas early homeostatic responses typically promote the expansion of regulatory T (T_{reg}) cells, the sustained pro-inflammatory and dyslipidaemic environment of atherosclerotic plaques alters the DC activation state, favouring the generation of CD4⁺ T helper (T_H) cells and effector T (T_{eff}) cells. Interestingly, oxPLs (as they are found in oxLDL) have been shown to induce a hyperactivated DC state¹³⁶. Both CD4⁺ T_{eff} and T_{reg} cells are likely to be primed in secondary lymphoid organs and recruited to the artery wall by specific chemokine–chemokine receptor pairs¹³⁷. Experimental studies in atherosclerosis-prone mice have documented the contribution of different DC and T cell subsets in atherosclerotic lesion formation, but insights into the antigen specificity of these responses is

still limited^{135,137}. Of note, Ley and colleagues have shown that many CD4⁺ T cells that show specificity for apoB peptides, which are presented by antigen-presenting cells, might initially have a T_{reg} cell identity^{138,139}. Only later, triggered by the local milieu of inflammatory cytokines, continuous antigen exposure and cellular metabolic alterations associated with progressing atherosclerotic lesions, do other T cell subsets with greater cytotoxic and inflammatory functions dominate, including those with an exT_{reg} cell phenotype¹⁴⁰. Indeed, T_{reg} cells protect mice from atherosclerotic lesion formation, whereas pro-inflammatory T_{H1} cells, including T_{H1}-like exT_{reg} cells, promote atherogenesis by propagating inflammation¹³⁷. The pivotal role of T cells in atherogenesis is also underscored by accumulating evidence that patients with cancer who are treated with immune checkpoint inhibitors, which unleash T cell activity, have an increased risk of vascular inflammation, atheroprogession and ASCVD events^{141–144}. In humans, T cells have been reported to be a prominent cellular component of atherosclerotic lesions, showing autoimmune-like reactivity^{145,146}, and atherosclerotic plaques from symptomatic compared with asymptomatic patients have a higher prevalence of chronically activated effector memory T cells with IFN γ signatures^{145,146}. CD8⁺ T cells are also found particularly in advanced atherosclerotic plaques, where they show various phenotypes and exert their function by mediating target cell lysis¹⁴⁷. Some CD8⁺ T cells have specificity for apoB¹⁴⁸.

In contrast to T cells, B cells are usually absent from intimal atherosclerotic lesions, but they are present in the so-called artery tertiary lymphoid organs, which are found in the adventitia surrounding advanced atherosclerotic plaques and are sites for germinal centre (GC) reactions¹⁴⁹ (Fig. 2). GC responses, which involve the help of T follicular helper cells, lead to the generation and clonal expansion of B cells that undergo affinity maturation and class switching and can further differentiate into memory B cells and long-lived plasma cells. Hyperlipidaemia has been found to promote GC responses in mice¹⁵⁰. Atherosclerosis is generally reduced in mice with B cells that are unable to class switch or to generate GC-dependent IgG antibodies, and was increased in antibody-deficient *Ldlr*^{-/-} mice that received IgG preparations from hyperlipidaemic *ApoE*^{-/-} mice compared with IgG from normolipidaemic wild-type donor mice¹⁴⁹. The specificity of these pathogenic IgG antibodies is unknown, but they might include anti-oxLDL antibodies as well as autoantibodies to classic self-antigens such as nucleic acids¹⁵¹. Indeed, in response to excessive lipid accumulation, DCs can promote the formation of autoantibodies by producing key survival factors, such as BAFF and APRIL¹⁵², for antibody-producing cells¹³⁴. Of note, both IgM and IgG are present in atherosclerotic plaques of humans and many of these immunoglobulins bind to epitopes of oxLDL¹⁵³.

In contrast to IgG antibodies, anti-oxLDL IgM antibodies are thought to mediate protective effects. Several epidemiological studies have shown that high plasma levels of IgM against epitopes of oxLDL are associated with a lower risk of cardiovascular events¹⁵⁴. Similarly, mice deficient in or with increased levels of natural IgM antibodies, which mostly bind to OSEs, show increased or reduced atherosclerosis, respectively^{155–157}. The atheroprotective mechanisms include neutralization of the pro-inflammatory effects of oxLDL and inhibition of oxLDL-induced foam cell formation. However, the same OSEs found on oxLDL are also present on dying cells and a subset of extracellular vesicles⁷⁹. Therefore, OSE-specific IgM can mediate additional protective functions by promoting the clearance and inhibiting the pro-inflammatory and pro-coagulatory properties of dying cells and extracellular vesicles, respectively¹⁵⁸. Innate-like B1 cells secreting IgM

are present in the perivascular adipose tissue of healthy arteries, where they might mediate important homeostatic functions in this regard¹⁵⁹.

Several strategies are also being investigated to exploit adaptive immune responses for anti-inflammatory therapies, including the expansion of T_{reg} cells and neutralizing anti-OSE-based antibody therapies targeting crucial moieties of oxLDL¹³³.

Beyond LDL: relative atherogenicity of apoB-containing lipoproteins

Are all apoB-containing lipoproteins equally atherogenic?

Conceptually, in atherogenesis, apoB can be considered to be an ‘anchor’ – the common structural protein that, because of its properties (as described above), facilitates the retention of apoB-containing lipoproteins at susceptible sites in artery walls. ApoB itself is not inherently atherogenic; instead, it is the ‘cargo’ of the lipoprotein constituents that is first anchored and then released that has pro-atherogenic effects. Given the marked compositional variation in apoB-containing lipoproteins, from large VLDL and chylomicron remnants to small LDL and Lp(a), it should not be surprising that particles have different potentials to cause pathogenic changes. Even for relatively uniform LDL particles, variation in the proteome and the lipidome, and possibly in the glycosylation pattern, modulates their affinity for artery wall proteoglycans, supporting the proposition that not all apoB-containing lipoproteins are equally atherogenic^{160,161}. We have introduced the concept of ‘relative per-particle atherogenicity’, defined as the increase in ASCVD risk per unit change in lipoprotein concentration, relative to the benchmark of LDL^{160,161}. We now develop this concept further by categorizing pro-atherogenic properties as being ‘intrinsic’, ‘acquired’ or ‘permissive’, as illustrated in Fig. 3.

The major apoB-containing lipoprotein classes – LDL; triglyceride-rich lipoproteins (TRLs), comprising chylomicrons and VLDL, and their remnants (TRL/remnants); and Lp(a) – vary considerably in size, content of major and minor lipid components, and the ancillary apolipoproteins present. TRLs and their remnants contain apoC-I, apoC-II, apoC-III and apoE¹⁶², whereas Lp(a) has the large, highly polymorphic apo(a) attached to apoB¹⁶³ (Fig. 3). All these features can alter the atherogenic potential of a lipoprotein.

LDL atherogenicity

LDL particles are fairly homogeneous in size and composition, and variation in particle atherogenicity is likely to be dependent mainly on the conformation of apoB, as detailed above³. LDL subclasses exist, and the small, dense subfraction that accumulates in individuals with elevated plasma triglyceride levels has been reported to be more atherogenic than normal-sized LDL¹⁶⁴. This higher atherogenicity despite a lower cholesterol content per particle^{3,165} is potentially explained by the greater ability of small, dense LDL to bind to artery wall proteoglycans^{3,166,167}.

LDL can be classified as having primarily intrinsic pro-atherogenic properties, based on the ability of its apoB to bind to artery wall proteoglycans and on its cholesterol ‘load’ (Fig. 3). The relative homogeneity of LDL in the general population allows the use of LDL metrics as a benchmark when investigating the quantitative association of other lipoproteins with ASCVD^{3,160,161}.

Relative atherogenicity of TRL/remnants

Most studies elucidating how apoB-containing lipoproteins induce atherogenesis have focused on LDL, because it is the predominant carrier of cholesterol in the blood, and the principal means of delivering

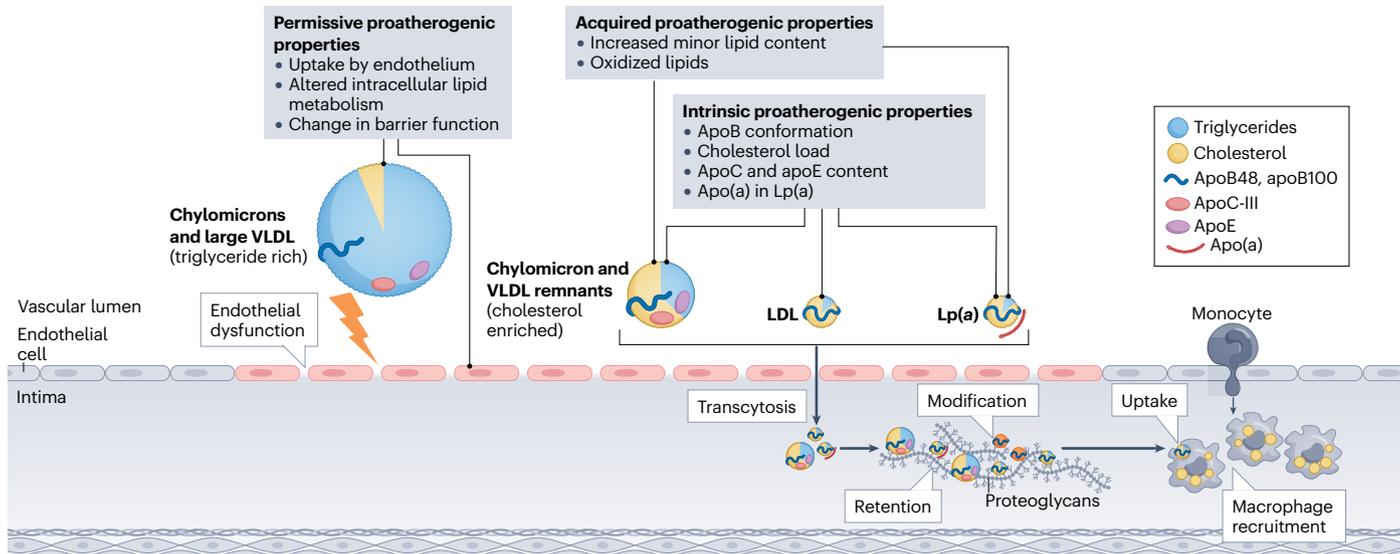


Fig. 3 | Concepts in atherogenicity of apoB-containing lipoproteins.

Apolipoprotein B (apoB)-containing lipoproteins are complex structures that vary substantially in size and composition. In the circulation, they undergo metabolic transformations, either gaining or losing constituents. This figure conceptualizes the notion that the atherogenic properties of a lipoprotein can be usefully categorized as being either intrinsic or acquired. Intrinsic properties are inherent to the core structure of a lipoprotein, such as the presence of apoB and

major lipids. Acquired properties result from the lipoprotein gaining potentially atherogenic components (such as minor apoproteins or oxidized lipids) by transfer or by metabolic modification occurring in the circulation or in the artery wall. In addition, permissive atherogenicity describes circumstances in which a particle itself does not enter the artery wall but induces pro-atherogenic changes, such as by increasing the atherogenic potential of other lipoprotein species. Lp(a), lipoprotein(a).

cholesterol into the artery wall. However, with the rising prevalence of obesity and type 2 diabetes, attention has widened to consider in greater detail the contribution of TRLs and their remnants (TRL/remnants) to atherogenesis, because these particles accumulate in individuals with these conditions. Furthermore, worldwide epidemiological studies have shown that TRL/remnants are major risk factors for cardiovascular disease in the general population, and Mendelian randomization studies have clearly demonstrated that TRL/remnants are causally associated with an increased risk of ASCVD events¹⁶⁸. The availability of large cohorts with genetic, phenotypic and outcome data has made it possible to investigate and quantify the relative atherogenicity of lipoproteins. Theoretically, by selecting allelic variants (single-nucleotide polymorphisms (SNPs)) that affect mainly a single lipoprotein class, it is possible to compare the increase in ASCVD risk associated with a change in the concentration of different types of lipoprotein. Furthermore, by focusing on changes in apoB, the atherogenicity of lipoproteins can be compared on a per-particle basis.

Early studies selected a small number of SNPs that seemed to affect only TRLs (such as variants in *LPL*) and compared them with SNPs that seemed to affect only LDL (such as variants in *LDLR*). Each set of SNPs altered plasma levels of apoB, and when the ASCVD risk per standardized change in apoB (particle number) was calculated, it was found to be similar for TRL and LDL. This result suggested that both lipoprotein classes were equally atherogenic¹⁶⁹. However, in a subsequent investigation¹⁶⁰, we used an arguably more objective approach by including all 1,125 SNPs affecting TRL/remnants and LDL in the Mendelian randomization analysis and found that no variant affected only one lipoprotein class^{11,170}. We divided the SNPs into subsets based on their effect on TRL/remnants relative to LDL. For the set of SNPs that more affected TRL/remnants, the ASCVD risk per standardized change

in apoB was increased by an odds ratio of 1.76 (95% CI 1.58–1.96), compared with an odds ratio of 1.33 (95% CI 1.26–1.40) for the set of SNPs that more affected LDL¹⁶⁰. Further investigation refined this observation and revealed that TRL/remnant particles have an approximately fourfold greater atherogenicity than LDL¹⁷¹, a finding with implications for our understanding of atherogenesis, cardiovascular risk assessment and the interpretation of clinical trials.

Mechanisms of TRL atherogenicity

Various mechanisms might explain the high atherogenicity of TRL/remnants. First, the increased atherogenic potential might be due to the high cholesterol content per particle or to preferential binding of TRL/remnants to artery wall proteoglycans due to an altered apoB conformation or to the presence of apoE or apoC-III^{162,172–175}. These features can all be considered to increase the intrinsic atherogenicity of the particle (Fig. 3). The fact that TRL/remnants contain about two times more cholesterol per apoB than LDL means that the cholesterol load delivered per particle is higher, and evidence also indicates that remnant particles can be ingested by macrophages without the requirement for modification (unlike LDL)¹⁷⁶. Furthermore, whereas LDL contains a single molecule of apoB per particle, TRLs contain several molecules of apoE in addition to apoB, allowing TRL and their remnants to bind more strongly to proteoglycans than LDL. Therefore, an augmented LDL-like mechanism of cholesterol deposition at atherosclerotic lesion sites might be at least partly responsible for the higher atherogenicity of TRL/remnants than of LDL. In support of this contention, we observed in genetic studies that TRL/remnant cholesterol content seemed to explain a substantial portion of the increased per-particle atherogenicity¹⁷¹. However, we and others have also observed that cholesterol from TRL/remnants is associated with a

higher ASCVD risk per standardized increase (per millimole per litre) than cholesterol from LDL, suggesting that factors other than just cholesterol load contribute to the higher atherogenicity of TRL/remnant particles^{160,171,177,178}.

A second plausible explanation lies in the observed relationship between plasma triglyceride levels and inflammation. A strong, positive association exists between plasma triglyceride levels and biomarkers of chronic systemic inflammation^{162,179,180}. The mechanism underlying this link is unclear, but it seems to be causal given that SNPs affecting TRL concentration influence plasma C-reactive protein levels and blood cell indices of inflammation¹⁷¹. This finding might reflect states of metabolic inflammation associated with lipid accumulation in the liver and adipose tissue rather than vascular inflammation. Conversely, TRLs carry high levels of apoC-III, which has been shown to trigger activation of the NLRP3 inflammasome in human monocytes, a process associated with the production of reactive oxygen species and pro-inflammatory cytokines¹⁸¹. An additional potential mechanism linked to inflammation might be that, during intravascular lipolysis, TRL/remnants accumulate minor lipids (such as ceramide) and partial lipolysis products (such as diglycerides and lysophospholipids), which when released from TRL/remnants that have been retained in the subendothelial space can trigger pathogenic pathways^{11,170}. Alternatively, similar bioactive lipids can be generated by in situ lipolysis of proteoglycan-bound TRL/remnants, given that lipoprotein lipase is secreted by cells present in the arterial wall⁸⁸. Free fatty acids released during the lipolysis of TRL triglyceride have been linked to pro-inflammatory effects on endothelial cells and monocyte-derived macrophages^{182,183}. This action is more pronounced when triglycerides are rich in saturated fatty acids rather than polyunsaturated fatty acids^{182,184}. Therefore, although the major lipid in TRL – triglyceride – might not itself contribute to atherogenesis, metabolic changes induced in TRL constituents with the generation of partial lipolysis products and also oxidized lipid species (due to the presence of reactive oxygen species in the artery wall) can be considered to be examples of acquired atherogenicity^{185–189} (Fig. 3).

A third potential mechanism linking TRL to pathogenic changes in artery walls lies in the cellular changes that occur in the endothelium when it is exposed to TRL. The observation that large TRL particles (chylomicrons and large VLDL) are too big to cross the endothelium by transcytosis led to the belief that these particles did not contribute to atherogenesis¹⁹⁰. However, studies now indicate that endothelial cells can internalize circulating large TRLs via the SR-B1 receptor¹⁸⁵. This uptake promotes changes in intracellular lipid metabolism and deleterious effects on the normal quiescent state of endothelial cells, impairing their barrier function^{186,191,192}. Two studies have uncovered a novel mechanistic link between large TRLs and vascular disease, mediated by the induction of lipid droplet formation in the endothelium^{191,193}. The accumulation of these droplets in endothelial cells promoted atherosclerosis and hypertension. More specifically, when mice were exposed to a high-fat diet or subjected to the loss of adipose triglyceride lipase (one of the major intracellular lipases), triglyceride-rich droplets formed in the endothelial layer of artery walls. Moreover, the presence of these droplets was associated with inhibition of nitric oxide production, thereby reducing endothelium-mediated relaxation and promoting vasoconstriction, which in turn contributed to the development of hypertension. In addition, there was activation of the NF- κ B pathway, leading to upregulation of VCAM1, a factor that facilitates leukocyte adhesion to endothelial cells and promotes atherosclerotic plaque inflammation^{191–193}. These changes in the state of the endothelium

can in turn induce increased influx of other apoB-containing lipoproteins (remnants and LDL) into the artery wall, and can be classed as permissive atherogenic effects¹⁸⁶ (Fig. 3).

Finally, when considering the role of TRL in atherosclerosis, the observation that the plasma levels of chylomicrons and large VLDL increase markedly after a meal must be taken into account. An acute elevation in the plasma levels of TRL and their remnants during the postprandial phase in humans has been shown to result in impaired vasodilatation, increased production of pro-inflammatory cytokines, heightened endothelial inflammatory response, increased expression of VCAM1 and monocyte activation^{182,183}. These effects are mediated through both direct and indirect mechanisms^{182,183}. In addition, chylomicrons and VLDL bind to and transport coagulation factor VII and factor X^{194,195}. Consequently, throughout the day as meals are absorbed, the appearance of TRLs and their remnants can create conditions that promote atherosclerotic plaque rupture and thrombus formation.

Relative atherogenicity of Lp(a)

Lp(a) is formed in the liver by the addition of apo(a) to an LDL particle. Apo(a), the product of the *LPA* gene, is a large, highly polymorphic protein of unknown function that is found in a few animal species, including humans. Apo(a) shares substantial homology with plasminogen and, on this basis, was initially thought to be a regulator of thrombosis¹⁶³. The plasma concentration of Lp(a) is highly variable and largely under genetic control. Epidemiological studies have consistently shown that Lp(a) is one of the strongest inherited causal risk factors for ASCVD and, accordingly, Lp(a) is considered to be a prime target for novel drug-based intervention^{163,196,197}. Attempts have been made to quantify the relationship between Lp(a) and ASCVD, and a range of results were obtained, suggesting that large reductions – approximately 50–100 mg/dl in total Lp(a) mass – might be required to see the same benefit as a 1 mmol/l reduction in plasma LDL cholesterol level^{163,198,199}. However, Mendelian randomization studies focusing on the apoB present in Lp(a) and LDL reported that Lp(a) particles were substantially (approximately sixfold) more atherogenic than LDL particles¹⁶¹, a result that was confirmed using measured Lp(a) and apoB levels²⁰⁰. This finding suggests that lowering plasma Lp(a) levels might be a useful intervention in a wider group of people than previously thought.

Role of Lp(a) in atherogenesis

In statistical models, the ASCVD risk associated with LDL cholesterol can be fully accounted for by adjusting for apoB, but the risk associated with Lp(a) cannot be explained in the same way^{161,201}. This finding suggests that the increased atherogenicity of Lp(a) is likely to be due to components that are unique to Lp(a) and which are not present on LDL. Leading candidates include the apo(a) protein and minor lipid constituents, such as oxPLs^{82,202,203}.

Apo(a) on Lp(a) might promote atherogenesis by binding to laminin, a component of the extracellular matrix in artery walls. As noted for TRL/remnants, augmented particle retention in the artery wall can conceivably facilitate cholesterol deposition and consequently the initiation and growth of atherosclerotic lesions²⁰⁴. However, most research on the apo(a) protein has focused on its structural similarity to the fibrinolytic zymogen plasminogen. Apo(a) contains multiple copies of sequences similar to plasminogen kringle IV, followed by sequences similar to kringle V and the protease domain of plasminogen^{205,206}. Nevertheless, the role of Lp(a) in the regulation of thrombosis and in thrombotic disease remains unclear²⁰⁷. Although

apo(a) can inhibit plasmin-mediated fibrinolysis *in vitro*, *ex vivo* studies have shown no change in clot lysis times using plasma taken from humans before and after reducing Lp(a) levels with an apo(a) antisense oligonucleotide²⁰⁸. Earlier studies in humans did suggest a positive association between high plasma Lp(a) levels and venous thromboembolism^{209,210}, but subsequent reports are inconclusive^{211,212}, and genetic evidence does not support this association except at extremely high plasma Lp(a) levels¹⁹⁷.

Another consideration about the role of apo(a) in cardiovascular disease is the marked size polymorphism of the protein. Most people carry *LPA* alleles coding for apo(a) protein of different lengths, and genetic studies suggest that ASCVD risk is influenced not only by the total plasma Lp(a) concentration but also by the size of the apo(a) protein, which is determined by the number of kringle IV repeats present^{163,197,213}. The specific effect of this polymorphism on the overall atherogenic potential of Lp(a) remains unclear.

OxPLs formed by enzymatic and non-enzymatic lipid peroxidation are found in tissues throughout the body. These minor lipid species have a crucial role as endogenous DAMPs, which are recognized by the innate immune system and trigger a sterile inflammatory response⁸¹. For example, oxPLs can activate the NF- κ B pathway and engage the NLRP3 inflammasome, leading to the production of pro-inflammatory cytokines⁷⁹. Although oxPLs are present in all lipoproteins in the circulation, about 85% of lipoprotein-associated oxPLs in humans are carried by Lp(a)²¹⁴, either bound to the apo(a) protein or located on the surface of the particle²¹⁵.

OxPLs can promote atherogenesis by increasing the expression of adhesion molecules and cytokines in endothelial cells, thereby facilitating the infiltration of inflammatory cells into the vessel wall, and by stimulating the proliferation of VSMCs and macrophages²¹⁶. Studies in animal models have shown that reducing the cellular uptake of oxPLs substantially decreases atherosclerosis¹³¹. Likewise, female double transgenic mice expressing human apoB100 and human apo(a) have increased atherosclerosis with a vulnerable plaque phenotype compared with those expressing human apoB100 only²¹⁷. Translating these findings to humans, plasma oxPL levels have been identified as novel risk factors for ASCVD in a number of epidemiological studies⁸².

The results of the genetic association studies described above are encouraging but a note of caution is warranted in using these findings to predict the outcome of Lp(a)-lowering clinical trials, because we do not yet understand the basis of the greater atherogenicity of Lp(a). If the presence of apo(a) alters the conformation of apoB and this change increases particle atherogenicity, or if apo(a) itself has important pro-atherogenic properties, then Lp(a) can be considered to have high intrinsic atherogenicity¹⁶³ (Fig. 3). Conversely, the greater atherogenicity of Lp(a) might be an acquired characteristic, due primarily to the presence of minor lipids (such as oxPLs) and other potentially bioactive molecules (such as diglycerides and lysophosphatidic acid) that provoke pro-inflammatory pathways^{82,214,218}. In an informative investigation in humans, these minor lipid species were shown to be enriched in Lp(a) and could activate monocytes, increasing the capacity of these monocytes to transit across an endothelial layer²¹⁸. However, profound lowering of plasma Lp(a) levels did not lead to a concomitant decrease in total plasma diacylglycerol levels, and the decrease in oxPLs was much less than that of Lp(a)²¹⁸. These intriguing results might be explained by redistribution of these minor lipid species to other lipoproteins that were not reduced by the treatment.

Of note, Lp(a) has been identified as a causal risk factor for aortic valve stenosis through mechanisms related to promotion of calcification of valve tissue¹⁶³. The role of Lp(a) in this disease is beyond the scope of this Review, but this association points to the possibility that Lp(a) might be involved in pathological mechanisms beyond atherogenesis.

An integrated view of apoB-related atherogenesis

Any assessment of the atherogenicity of apoB-containing lipoproteins must consider their relative abundance in the blood circulation. TRL/remnants and Lp(a) might be more atherogenic than LDL on a per-particle basis, but LDL is much more abundant. Typically, the concentration of apoB contained in LDL is about 80 mg/dl and that contained in TRL/remnants is about 10 mg/dl, whereas the concentration of apoB contained in Lp(a) varies from almost zero to about 20 mg/dl¹⁶³. Lowering plasma LDL levels is, therefore, still the first-line intervention strategy. However, in people who are receiving LDL-lowering therapy and who achieve the new guideline-recommended targets of a plasma LDL cholesterol level of <1.4 mmol/l (55 mg/dl)²¹⁹, the concentration of apoB contained in LDL would be <35 mg/dl, and other apoB-containing lipoproteins, if their plasma levels are elevated, would contribute substantially to the residual risk of ASCVD.

Identifying the causes of the increased particle atherogenicity of different apoB-containing lipoproteins might be important because it is conceivable that only the intrinsic atherogenic effects are decreased in direct proportion to reducing the concentration of the apoB-containing lipoprotein in the bloodstream. Indeed, for LDL, which has mainly intrinsic atherogenicity, a meta-analysis of trials has shown a linear relationship between LDL cholesterol reduction and ASCVD risk reduction²²⁰. If the cause of increased TRL/remnant and Lp(a) atherogenicity is partly acquired, then reductions in the plasma levels of these lipoprotein classes by specific lipid-lowering treatments might or might not lead to a proportional reduction in ASCVD event rates. Acquired bioactive components might partly or wholly redistribute to other lipoproteins and continue to exert a pro-atherogenic influence.

When clinical trial experience of novel therapies such as specific Lp(a)-lowering and TRL/remnant-lowering agents is available, the outcome can be compared with quantitative predictions, especially those from Mendelian randomization studies, which have many of the features of a randomized clinical trial. Information on therapies that can reduce plasma Lp(a) levels by >80% will be published in the next few years²²¹. Trials on lowering of plasma TRL/remnant levels have produced mixed results. Early investigations of fibrates, which lower plasma TRL/remnant levels by 25–40%, demonstrated reductions in the risk of ASCVD²²². However, in the PROMINENT trial²²³, pema-fibrate lowered plasma triglyceride and TRL cholesterol levels by 25–30% but showed no reduction in the risk of ASCVD. This outcome might be attributable to the concomitant rise in plasma LDL cholesterol and apoB levels in those receiving pema-fibrate^{224,225}, even allowing for the difference in atherogenicity between TRL/remnants and LDL particles¹⁷¹.

Conclusions

A large body of evidence supports the predominant role of LDL as a causative agent in initiating and progressing the formation of atherosclerotic plaques. We now have a detailed understanding of how LDL penetrates, is retained by and elicits a maladaptive response in the artery wall at sites of susceptibility. The other apoB-containing lipoproteins – TRL and Lp(a) – seem to be even more atherogenic than archetypal LDL but are usually present at lower concentrations in the

plasma. Multiple targets for intervention can be identified, including lowering the plasma levels of apoB-containing lipoproteins and modulating the cellular responses to the presence of lipoproteins in artery walls. These strategies remain to be tested in clinical outcome trials.

Published online: 02 January 2025

References

1. Tsao, C. W. et al. Heart disease and stroke statistics-2022 update: a report from the American Heart Association. *Circulation* **145**, e153–e639 (2022).
2. Ference, B. A. et al. Low-density lipoproteins cause atherosclerotic cardiovascular disease. 1. Evidence from genetic, epidemiologic, and clinical studies. A consensus statement from the European Atherosclerosis Society Consensus Panel. *Eur. Heart J.* **38**, 2459–2472 (2017).
3. Boren, J. et al. Low-density lipoproteins cause atherosclerotic cardiovascular disease: pathophysiological, genetic, and therapeutic insights: a consensus statement from the European Atherosclerosis Society Consensus Panel. *Eur. Heart J.* **41**, 2313–2330 (2020).
4. Skalen, K. et al. Subendothelial retention of atherogenic lipoproteins in early atherosclerosis. *Nature* **417**, 750–754 (2002).
5. Tabas, I., Williams, K. J. & Boren, J. Subendothelial lipoprotein retention as the initiating process in atherosclerosis: update and therapeutic implications. *Circulation* **116**, 1832–1844 (2007).
6. Robinson, J. G. et al. Eradicating the burden of atherosclerotic cardiovascular disease by lowering apolipoprotein B lipoproteins earlier in life. *J. Am. Heart Assoc.* **7**, e009778 (2018).
7. Williams, K. J. & Tabas, I. The response-to-retention hypothesis of early atherogenesis. *Arterioscler. Thromb. Vasc. Biol.* **15**, 551–561 (1995).
8. Camejo, G., Lopez, A., Vegas, H. & Paoli, H. The participation of aortic proteins in the formation of complexes between low density lipoproteins and intima-media extracts. *Atherosclerosis* **21**, 77–91 (1975).
9. Segrest, J. P., Jones, M. K., De Loof, H. & Dashti, N. Structure of apolipoprotein B-100 in low density lipoproteins. *J. Lipid Res.* **42**, 1346–1367 (2001).
10. Segrest, J. P. et al. Apolipoprotein B-100: conservation of lipid-associating amphipathic secondary structural motifs in nine species of vertebrates. *J. Lipid Res.* **39**, 85–102 (1998).
11. Boren, J., Taskinen, M. R., Bjornson, E. & Packard, C. J. Metabolism of triglyceride-rich lipoproteins in health and dyslipidaemia. *Nat. Rev. Cardiol.* **19**, 577–592 (2022).
12. Frank, P. G. & Lisanti, M. P. Caveolin-1 and caveolae in atherosclerosis: differential roles in fatty streak formation and neointimal hyperplasia. *Curr. Opin. Lipidol.* **15**, 523–529 (2004).
13. Fernandez-Hernando, C. et al. Genetic evidence supporting a critical role of endothelial caveolin-1 during the progression of atherosclerosis. *Cell Metab.* **10**, 48–54 (2009).
14. Frank, P. G., Pavlides, S. & Lisanti, M. P. Caveolae and transcytosis in endothelial cells: role in atherosclerosis. *Cell Tissue Res.* **335**, 41–47 (2009).
15. Armstrong, S. M. et al. Novel assay for detection of LDL transcytosis across coronary endothelium reveals an unexpected role for SR-B1 [abstract]. *Circulation* **130** (Suppl. 2), A11607 (2014).
16. Kraehling, J. R. et al. Genome-wide RNAi screen ALK1 mediates LDL uptake and transcytosis in endothelial cells. *Nat. Commun.* **7**, 13516 (2016).
17. Minick, C. R., Stemerman, M. G. & Insull, W. Jr Effect of regenerated endothelium on lipid accumulation in the arterial wall. *Proc. Natl Acad. Sci. USA* **74**, 1724–1728 (1977).
18. Minick, C. R., Stemerman, M. B. & Insull, W. Jr Role of endothelium and hypercholesterolemia in intimal thickening and lipid accumulation. *Am. J. Pathol.* **95**, 131–158 (1979).
19. Armstrong, S. M. et al. A novel assay uncovers an unexpected role for SR-B1 in LDL transcytosis. *Cardiovasc. Res.* **108**, 268–277 (2015).
20. Huang, L. et al. SR-B1 drives endothelial cell LDL transcytosis via DOCK4 to promote atherosclerosis. *Nature* **569**, 565–569 (2019).
21. Sessa, W. C. Estrogen reduces LDL (low-density lipoprotein) transcytosis. *Arterioscler. Thromb. Vasc. Biol.* **38**, 2276–2277 (2018).
22. Ghaffari, S., Naderi Nabi, F., Sugiyama, M. G. & Lee, W. L. Estrogen inhibits LDL (low-density lipoprotein) transcytosis by human coronary artery endothelial cells via GPER (G-protein-coupled estrogen receptor) and SR-B1 (scavenger receptor class B type 1). *Arterioscler. Thromb. Vasc. Biol.* **38**, 2283–2294 (2018).
23. Mathur, P., Ostadal, B., Romeo, F. & Mehta, J. L. Gender-related differences in atherosclerosis. *Cardiovasc. Drugs Ther.* **29**, 319–327 (2015).
24. Bian, F., Yang, X. Y., Xu, G., Zheng, T. & Jin, S. CRP-induced NLRP3 inflammasome activation increases LDL transcytosis across endothelial cells. *Front. Pharmacol.* **10**, 40 (2019).
25. Jia, X. et al. VCAM-1-binding peptide targeted cationic liposomes containing NLRP3 siRNA to modulate LDL transcytosis as a novel therapy for experimental atherosclerosis. *Metabolism* **135**, 155274 (2022).
26. Arsenault, B. J., Carpentier, A. C., Poirier, P. & Despres, J. P. Adiposity, type 2 diabetes and atherosclerotic cardiovascular disease risk: use and abuse of the body mass index. *Atherosclerosis* **394**, 117546 (2024).
27. Bartels, E. D., Christoffersen, C., Lindholm, M. W. & Nielsen, L. B. Altered metabolism of LDL in the arterial wall precedes atherosclerosis regression. *Circ. Res.* **117**, 933–942 (2015).
28. Mundi, S. et al. Endothelial permeability, LDL deposition, and cardiovascular risk factors – a review. *Cardiovasc. Res.* **114**, 35–52 (2018).
29. van den Berg, B. M., Spaan, J. A., Rolf, T. M. & Vink, H. Atherogenic region and diet diminish glycocalyx dimension and increase intima-to-media ratios at murine carotid artery bifurcation. *Am. J. Physiol. Heart Circ. Physiol.* **290**, H915–H920 (2006).
30. Lewis, J. C., Taylor, R. G., Jones, N. D., St Clair, R. W. & Cornhill, J. F. Endothelial surface characteristics in pigeon coronary artery atherosclerosis. I. Cellular alterations during the initial stages of dietary cholesterol challenge. *Lab. Invest.* **46**, 123–138 (1982).
31. Cancel, L. M., Ebong, E. E., Mensah, S., Hirschberg, C. & Tarbell, J. M. Endothelial glycocalyx, apoptosis and inflammation in an atherosclerotic mouse model. *Atherosclerosis* **252**, 136–146 (2016).
32. Banerjee, S., Mwangi, J. G., Stanley, T. K., Mitra, R. & Ebong, E. E. Regeneration and assessment of the endothelial glycocalyx to address cardiovascular disease. *Ind. Eng. Chem. Res.* **60**, 17328–17347 (2021).
33. Faber, M. The human aorta; sulfate-containing polyuronides and the deposition of cholesterol. *Arch. Pathol.* **48**, 342–350 (1949).
34. Camejo, G. et al. Differences in the structure of plasma low-density lipoproteins and their relationship to the extent of interaction with arterial wall-components. *Ann. N. Y. Acad. Sci.* **275**, 153–168 (1976).
35. Camejo, G. The interaction of lipids and lipoproteins with the intercellular matrix of arterial tissue: its possible role in atherogenesis. *Adv. Lipid Res.* **19**, 1–53 (1982).
36. Camejo, G., Olofsson, S. O., Lopez, F., Carlsson, P. & Bondjers, G. Identification of Apo B-100 segments mediating the interaction of low density lipoproteins with arterial proteoglycans. *Arteriosclerosis* **8**, 368–377 (1988).
37. Hirose, N., Blankenship, D. T., Krivanek, M. A., Jackson, R. L. & Cardin, A. D. Isolation and characterization of four heparin-binding cyanogen bromide peptides of human plasma apolipoprotein B. *Biochemistry* **26**, 5505–5512 (1987).
38. Weisgraber, K. H. & Rall, S. C. Jr Human apolipoprotein B-100 heparin-binding sites. *J. Biol. Chem.* **262**, 11097–11103 (1987).
39. Boren, J. et al. Identification of the principal proteoglycan-binding site in LDL. A single-point mutation in apo-B100 severely affects proteoglycan interaction without affecting LDL receptor binding. *J. Clin. Invest.* **101**, 2658–2664 (1998).
40. Boren, J. et al. Identification of the low density lipoprotein receptor-binding site in apolipoprotein B100 and the modulation of its binding activity by the carboxyl terminus in familial defective apo-B100. *J. Clin. Invest.* **101**, 1084–1093 (1998).
41. Chan, L. Apolipoprotein B, the major protein component of triglyceride-rich and low density lipoproteins. *J. Biol. Chem.* **267**, 25621–25624 (1992).
42. Flood, C. et al. Identification of the proteoglycan binding site in apolipoprotein B48. *J. Biol. Chem.* **277**, 32228–32233 (2002).
43. Flood, C. et al. Molecular mechanism for changes in proteoglycan binding on compositional changes of the core and the surface of low-density lipoprotein-containing human apolipoprotein B100. *Arterioscler. Thromb. Vasc. Biol.* **24**, 564–570 (2004).
44. Camejo, G., Olsson, U., Hurt-Camejo, E., Baharamian, N. & Bondjers, G. The extracellular matrix on atherogenesis and diabetes-associated vascular disease. *Atheroscler. Suppl.* **3**, 3–9 (2002).
45. Sartipy, P., Camejo, G., Svensson, L. & Hurt-Camejo, E. Phospholipase A₂ modification of low density lipoproteins forms small high density particles with increased affinity for proteoglycans and glycosaminoglycans. *J. Biol. Chem.* **274**, 25913–25920 (1999).
46. Kugiyama, K. et al. Circulating levels of secretory type II phospholipase A₂ predict coronary events in patients with coronary artery disease. *Circulation* **100**, 1280–1284 (1999).
47. The Lp-PLA2 Studies Collaboration et al. Lipoprotein-associated phospholipase A₂ and risk of coronary disease, stroke, and mortality: collaborative analysis of 32 prospective studies. *Lancet* **375**, 1536–1544 (2010).
48. Griffin, B. A. et al. Role of plasma triglyceride in the regulation of plasma low density lipoprotein (LDL) subfractions: relative contribution of small, dense LDL to coronary heart disease risk. *Atherosclerosis* **106**, 241–253 (1994).
49. Austin, M. A., King, M. C., Vranizan, K. M. & Krauss, R. M. Atherogenic lipoprotein phenotype. A proposed genetic marker for coronary heart disease risk. *Circulation* **82**, 495–506 (1990).
50. Nicholls, S. J. et al. Varespladib and cardiovascular events in patients with an acute coronary syndrome: the VISTA-16 randomized clinical trial. *JAMA* **311**, 252–262 (2014).
51. Gregson, J. M. et al. Genetic invalidation of Lp-PLA₂ as a therapeutic target: large-scale study of five functional Lp-PLA₂-lowering alleles. *Eur. J. Prev. Cardiol.* **24**, 492–504 (2017).
52. Schwenke, D. C. & Carew, T. E. Initiation of atherosclerotic lesions in cholesterol-fed rabbits. II. Selective retention of LDL vs. selective increases in LDL permeability in susceptible sites of arteries. *Arteriosclerosis* **9**, 908–918 (1989).
53. Tran-Lundmark, K. et al. Heparan sulfate in perlecan promotes mouse atherosclerosis: roles in lipid permeability, lipid retention, and smooth muscle cell proliferation. *Circ. Res.* **103**, 43–52 (2008).
54. Mahley, R. W. & Huang, Y. Atherogenic remnant lipoproteins: role for proteoglycans in trapping, transferring, and internalizing. *J. Clin. Invest.* **117**, 94–98 (2007).
55. Hiukka, A. et al. ApoCIII-enriched LDL in type 2 diabetes displays altered lipid composition, increased susceptibility for sphingomyelinase, and increased binding to biglycan. *Diabetes* **58**, 2018–2026 (2009).
56. Olin-Lewis, K. et al. ApoC-III content of apoB-containing lipoproteins is associated with binding to the vascular proteoglycan biglycan. *J. Lipid Res.* **43**, 1969–1977 (2002).

57. Jayaraman, S. et al. Effects of triacylglycerol on the structural remodeling of human plasma very low- and low-density lipoproteins. *Biochim. Biophys. Acta Mol. Cell Biol. Lipids* **1864**, 1061–1071 (2019).
58. Willner, E. L. et al. Deficiency of acyl CoA:cholesterol acyltransferase 2 prevents atherosclerosis in apolipoprotein E-deficient mice. *Proc. Natl Acad. Sci. USA* **100**, 1262–1267 (2003).
59. Aviram, M., Lund-Katz, S., Phillips, M. C. & Chait, A. The influence of the triglyceride content of low density lipoprotein on the interaction of apolipoprotein B-100 with cells. *J. Biol. Chem.* **263**, 16842–16848 (1988).
60. Hagensen, M. K. et al. Increased retention of LDL from type 1 diabetic patients in atherosclerosis-prone areas of the murine arterial wall. *Atherosclerosis* **286**, 156–162 (2019).
61. O'Brien, K. D. et al. Comparison of apolipoprotein and proteoglycan deposits in human coronary atherosclerotic plaques: colocalization of biglycan with apolipoproteins. *Circulation* **98**, 519–527 (1998).
62. Nakashima, Y., Fujii, H., Sumiyoshi, S., Wight, T. N. & Sueishi, K. Early human atherosclerosis: accumulation of lipid and proteoglycans in intimal thickenings followed by macrophage infiltration. *Arterioscler. Thromb. Vasc. Biol.* **27**, 1159–1165 (2007).
63. She, Z. G. et al. NG2 proteoglycan ablation reduces foam cell formation and atherogenesis via decreased low-density lipoprotein retention by synthetic smooth muscle cells. *Arterioscler. Thromb. Vasc. Biol.* **36**, 49–59 (2016).
64. Tsiantoulas, D. et al. APRIL limits atherosclerosis by binding to heparan sulfate proteoglycans. *Nature* **597**, 92–96 (2021).
65. Brito, V. et al. Atheroregressive potential of the treatment with a chimeric monoclonal antibody against sulfated glycosaminoglycans on pre-existing lesions in apolipoprotein E-deficient mice. *Front. Pharmacol.* **8**, 782 (2017).
66. Nakashima, Y., Chen, Y. X., Kinukawa, N. & Sueishi, K. Distributions of diffuse intimal thickening in human arteries: preferential expression in atherosclerosis-prone arteries from an early age. *Virchows Arch.* **441**, 279–288 (2002).
67. Stary, H. C. et al. A definition of the intima of human arteries and of its atherosclerosis-prone regions. A report from the Committee on Vascular Lesions of the Council on Arteriosclerosis, American Heart Association. *Arterioscler. Thromb.* **12**, 120–134 (1992).
68. Kaprio, J., Norio, R., Pesonen, E. & Sarna, S. Intimal thickening of the coronary arteries in infants in relation to family history of coronary artery disease. *Circulation* **87**, 1960–1968 (1993).
69. Allahverdian, S., Ortega, C. & Francis, G. A. Smooth muscle cell-proteoglycan-lipoprotein interactions as drivers of atherosclerosis. *Handb. Exp. Pharmacol.* **270**, 335–358 (2022).
70. Kolodgie, F. D., Burke, A. P., Nakazawa, G. & Virmani, R. Is pathologic intimal thickening the key to understanding early plaque progression in human atherosclerotic disease? *Arterioscler. Thromb. Vasc. Biol.* **27**, 986–989 (2007).
71. Kijani, S., Vazquez, A. M., Levin, M., Boren, J. & Fogelstrand, P. Intimal hyperplasia induced by vascular intervention causes lipoprotein retention and accelerated atherosclerosis. *Physiol. Rep.* **5**, e13334 (2017).
72. Kalan, J. M. & Roberts, W. C. Morphologic findings in saphenous veins used as coronary arterial bypass conduits for longer than 1 year: necropsy analysis of 53 patients, 123 saphenous veins, and 1865 five-millimeter segments of veins. *Am. Heart J.* **119**, 1164–1184 (1990).
73. Williams, K. J. Arterial zones that take a pause in early plaque development. *Arterioscler. Thromb. Vasc. Biol.* **43**, 650–653 (2023).
74. Lewis, E. A. et al. Capacity for LDL (low-density lipoprotein) retention predicts the course of atherogenesis in the murine aortic arch. *Arterioscler. Thromb. Vasc. Biol.* **43**, 637–649 (2023).
75. Oorni, K. et al. Acidification of the intimal fluid: the perfect storm for atherogenesis. *J. Lipid Res.* **56**, 203–214 (2015).
76. Tomas, L. et al. Altered metabolism distinguishes high-risk from stable carotid atherosclerotic plaques. *Eur. Heart J.* **39**, 2301–2310 (2018).
77. Glise, L. et al. pH-dependent protonation of histidine residues is critical for electrostatic binding of low-density lipoproteins to human coronary arteries. *Arterioscler. Thromb. Vasc. Biol.* **42**, 1037–1047 (2022).
78. Lorey, M. B., Oorni, K. & Kovanen, P. T. Modified lipoproteins induce arterial wall inflammation during atherogenesis. *Front. Cardiovasc. Med.* **9**, 841545 (2022).
79. Binder, C. J., Papac-Milicevic, N. & Witztum, J. L. Innate sensing of oxidation-specific epitopes in health and disease. *Nat. Rev. Immunol.* **16**, 485–497 (2016).
80. Esterbauer, H., Schaur, R. J. & Zollner, H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free. Radic. Biol. Med.* **11**, 81–128 (1991).
81. Bochkov, V. N. et al. Generation and biological activities of oxidized phospholipids. *Antioxid. Redox Signal.* **12**, 1009–1059 (2010).
82. Tsimikas, S. & Witztum, J. L. Oxidized phospholipids in cardiovascular disease. *Nat. Rev. Cardiol.* **21**, 170–191 (2024).
83. Byun, Y. S. et al. Relationship of oxidized phospholipids on apolipoprotein B-100 to cardiovascular outcomes in patients treated with intensive versus moderate atorvastatin therapy: the TNT trial. *J. Am. Coll. Cardiol.* **65**, 1286–1295 (2015).
84. Gilliland, T. C. et al. Lipoprotein(a), oxidized phospholipids, and coronary artery disease severity and outcomes. *J. Am. Coll. Cardiol.* **81**, 1780–1792 (2023).
85. Tsimikas, S. et al. Relationship of oxidized phospholipids on apolipoprotein B-100 particles to race/ethnicity, apolipoprotein(a) isoform size, and cardiovascular risk factors: results from the Dallas Heart Study. *Circulation* **119**, 1711–1719 (2009).
86. Camejo, G., Fager, G., Rosengren, B., Hurt-Camejo, E. & Bondjers, G. Binding of low density lipoproteins by proteoglycans synthesized by proliferating and quiescent human arterial smooth muscle cells. *J. Biol. Chem.* **268**, 14131–14137 (1993).
87. Chang, M. Y., Potter-Perigo, S., Tsoi, C., Chait, A. & Wight, T. N. Oxidized low density lipoproteins regulate synthesis of monkey aortic smooth muscle cell proteoglycans that have enhanced native low density lipoprotein binding properties. *J. Biol. Chem.* **275**, 4766–4773 (2000).
88. Gustafsson, M. et al. Retention of low-density lipoprotein in atherosclerotic lesions of the mouse: evidence for a role of lipoprotein lipase. *Circ. Res.* **101**, 777–783 (2007).
89. Pentikainen, M. O., Oksjoki, R., Oorni, K. & Kovanen, P. T. Lipoprotein lipase in the arterial wall: linking LDL to the arterial extracellular matrix and much more. *Arterioscler. Thromb. Vasc. Biol.* **22**, 211–217 (2002).
90. Pentikainen, M. O., Oorni, K. & Kovanen, P. T. Lipoprotein lipase (LPL) strongly links native and oxidized low density lipoprotein particles to decorin-coated collagen. Roles for both dimeric and monomeric forms of LPL. *J. Biol. Chem.* **275**, 5694–5701 (2000).
91. Babaev, V. R. et al. Macrophage lipoprotein lipase promotes foam cell formation and atherosclerosis in vivo. *J. Clin. Invest.* **103**, 1697–1705 (1999).
92. Wilson, K., Fry, G. L., Chappell, D. A., Sigmund, C. D. & Medh, J. D. Macrophage-specific expression of human lipoprotein lipase accelerates atherosclerosis in transgenic apolipoprotein E knockout mice but not in C57BL/6 mice. *Arterioscler. Thromb. Vasc. Biol.* **21**, 1809–1815 (2001).
93. Tabas, I. et al. Lipoprotein lipase and sphingomyelinase synergistically enhance the association of atherogenic lipoproteins with smooth muscle cells and extracellular matrix. A possible mechanism for low density lipoprotein and lipoprotein(a) retention and macrophage foam cell formation. *J. Biol. Chem.* **268**, 20419–20432 (1993).
94. Devlin, C. M. et al. Acid sphingomyelinase promotes lipoprotein retention within early atheromata and accelerates lesion progression. *Arterioscler. Thromb. Vasc. Biol.* **28**, 1723–1730 (2008).
95. Oorni, K., Hakala, J. K., Annala, A., Ala-Korpela, M. & Kovanen, P. T. Sphingomyelinase induces aggregation and fusion, but phospholipase A2 only aggregation, of low density lipoprotein (LDL) particles. Two distinct mechanisms leading to increased binding strength of LDL to human aortic proteoglycans. *J. Biol. Chem.* **273**, 29127–29134 (1998).
96. Wong, M. L. et al. Acute systemic inflammation up-regulates secretory sphingomyelinase in vivo: a possible link between inflammatory cytokines and atherogenesis. *Proc. Natl Acad. Sci. USA* **97**, 8681–8686 (2000).
97. Marathe, S., Kuriakose, G., Williams, K. J. & Tabas, I. Sphingomyelinase, an enzyme implicated in atherogenesis, is present in atherosclerotic lesions and binds to specific components of the subendothelial extracellular matrix. *Arterioscler. Thromb. Vasc. Biol.* **19**, 2648–2658 (1999).
98. Marathe, S., Choi, Y., Leventhal, A. R. & Tabas, I. Sphingomyelinase converts lipoproteins from apolipoprotein E knockout mice into potent inducers of macrophage foam cell formation. *Arterioscler. Thromb. Vasc. Biol.* **20**, 2607–2613 (2000).
99. Ruuth, M. et al. Susceptibility of low-density lipoprotein particles to aggregate depends on particle liposome, is modifiable, and associates with future cardiovascular deaths. *Eur. Heart J.* **39**, 2562–2573 (2018).
100. Sanda, G. M. et al. Aggregated LDL turn human macrophages into foam cells and induce mitochondrial dysfunction without triggering oxidative or endoplasmic reticulum stress. *PLoS ONE* **16**, e0245797 (2021).
101. Steinfeld, N., Ma, C. J. & Maxfield, F. R. Signaling pathways regulating the extracellular digestion of lipoprotein aggregates by macrophages. *Mol. Biol. Cell* **35**, ar5 (2024).
102. Cohen, J. C., Boerwinkle, E., Mosley, T. H. Jr. & Hobbs, H. H. Sequence variations in PCSK9, low LDL, and protection against coronary heart disease. *N. Engl. J. Med.* **354**, 1264–1272 (2006).
103. Nissen, S. E. et al. Effect of torcetrapib on the progression of coronary atherosclerosis. *N. Engl. J. Med.* **356**, 1304–1316 (2007).
104. Sneek, M. et al. Conformational changes of apoB-100 in SMase-modified LDL mediate formation of large aggregates at acidic pH. *J. Lipid Res.* **53**, 1832–1839 (2012).
105. Zerneck, A. et al. Meta-analysis of leukocyte diversity in atherosclerotic mouse aortas. *Circ. Res.* **127**, 402–426 (2020).
106. Depuydt, M. A. C. et al. Microanatomy of the human atherosclerotic plaque by single-cell transcriptomics. *Circ. Res.* **127**, 1437–1455 (2020).
107. de Winther, M. P. J. et al. Translational opportunities of single-cell biology in atherosclerosis. *Eur. Heart J.* **44**, 1216–1230 (2023).
108. Williams, J. W. et al. Limited proliferation capacity of aortic intima resident macrophages requires monocyte recruitment for atherosclerotic plaque progression. *Nat. Immunol.* **21**, 1194–1204 (2020).
109. Takaoka, M. et al. Early intermittent hyperlipidaemia alters tissue macrophages to fuel atherosclerosis. *Nature* **634**, 457–465 (2024).
110. Moore, K. J. et al. Macrophage trafficking, inflammatory resolution, and genomics in atherosclerosis: JACC Macrophage in CVD Series (part 2). *J. Am. Coll. Cardiol.* **72**, 2181–2197 (2018).
111. Spann, N. J. et al. Regulated accumulation of desmosterol integrates macrophage lipid metabolism and inflammatory responses. *Cell* **151**, 138–152 (2012).
112. Cochain, C. et al. Single-cell RNA-seq reveals the transcriptional landscape and heterogeneity of aortic macrophages in murine atherosclerosis. *Circ. Res.* **122**, 1661–1674 (2018).
113. Piolet, M. et al. TREM2 protects from atherosclerosis by limiting necrotic core formation. *Nat. Cardiovasc. Res.* **3**, 269–282 (2024).

114. Dib, L. et al. Lipid-associated macrophages transition to an inflammatory state in human atherosclerosis increasing the risk of cerebrovascular complications. *Nat. Cardiovasc. Res.* **2**, 656–672 (2023).
115. Zernecke, A. et al. Integrated single-cell analysis-based classification of vascular mononuclear phagocytes in mouse and human atherosclerosis. *Cardiovasc. Res.* **119**, 1676–1689 (2023).
116. Adkar, S. S. & Leeper, N. J. Efferocytosis in atherosclerosis. *Nat. Rev. Cardiol.* **21**, 762–779 (2024).
117. Doran, A. C., Yurdagül, A. Jr. & Tabas, I. Efferocytosis in health and disease. *Nat. Rev. Immunol.* **20**, 254–267 (2020).
118. Seimon, T. A. et al. Atherogenic lipids and lipoproteins trigger CD36-TLR2-dependent apoptosis in macrophages undergoing endoplasmic reticulum stress. *Cell Metab.* **12**, 467–482 (2010).
119. De Meyer, G. R. Y., Zurek, M., Puylaert, P. & Martinet, W. Programmed death of macrophages in atherosclerosis: mechanisms and therapeutic targets. *Nat. Rev. Cardiol.* **21**, 312–325 (2024).
120. Lehti, S. et al. Extracellular lipids accumulate in human carotid arteries as distinct three-dimensional structures and have proinflammatory properties. *Am. J. Pathol.* **188**, 525–538 (2018).
121. Oorni, K. & Kovanen, P. T. Enhanced extracellular lipid accumulation in acidic environments. *Curr. Opin. Lipidol.* **17**, 534–540 (2006).
122. Kiss, M. G. & Binder, C. J. The multifaceted impact of complement on atherosclerosis. *Atherosclerosis* **351**, 29–40 (2022).
123. Duester, P. et al. NLRP3 inflammasomes are required for atherogenesis and activated by cholesterol crystals. *Nature* **464**, 1357–1361 (2010).
124. Rajamaki, K. et al. Cholesterol crystals activate the NLRP3 inflammasome in human macrophages: a novel link between cholesterol metabolism and inflammation. *PLoS ONE* **5**, e11765 (2010).
125. Grebe, A., Hoss, F. & Latz, E. NLRP3 inflammasome and the IL-1 pathway in atherosclerosis. *Circ. Res.* **122**, 1722–1740 (2018).
126. Ridker, P. M. et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N. Engl. J. Med.* **377**, 1119–1131 (2017).
127. Fiolet, A. T. L. et al. Efficacy and safety of low-dose colchicine in patients with coronary disease: a systematic review and meta-analysis of randomized trials. *Eur. Heart J.* **42**, 2765–2775 (2021).
128. Ridker, P. M. From RESCUE to ZEUS: will interleukin-6 inhibition with ziltivekimab prove effective for cardiovascular event reduction? *Cardiovasc. Res.* **117**, e138–e140 (2021).
129. Stewart, C. R. et al. CD36 ligands promote sterile inflammation through assembly of a Toll-like receptor 4 and 6 heterodimer. *Nat. Immunol.* **11**, 155–161 (2010).
130. Sheedy, F. J. et al. CD36 coordinates NLRP3 inflammasome activation by facilitating intracellular nucleation of soluble ligands into particulate ligands in sterile inflammation. *Nat. Immunol.* **14**, 812–820 (2013).
131. Que, X. et al. Oxidized phospholipids are proinflammatory and proatherogenic in hypercholesterolaemic mice. *Nature* **558**, 301–306 (2018).
132. Busch, C. J. et al. Malondialdehyde epitopes are sterile mediators of hepatic inflammation in hypercholesterolemic mice. *Hepatology* **65**, 1181–1195 (2017).
133. Mallat, Z. & Binder, C. J. The why and how of adaptive immune responses in ischemic cardiovascular disease. *Nat. Cardiovasc. Res.* **1**, 431–444 (2022).
134. Ito, A. et al. Cholesterol accumulation in CD11c⁺ immune cells is a causal and targetable factor in autoimmune disease. *Immunity* **45**, 1311–1326 (2016).
135. Gil-Pulido, J. & Zernecke, A. Antigen-presenting dendritic cells in atherosclerosis. *Eur. J. Pharmacol.* **816**, 25–31 (2017).
136. Zhivaki, D. & Kagan, J. C. Innate immune detection of lipid oxidation as a threat assessment strategy. *Nat. Rev. Immunol.* **22**, 322–330 (2022).
137. Saigusa, R., Winkels, H. & Ley, K. T cell subsets and functions in atherosclerosis. *Nat. Rev. Cardiol.* **17**, 387–401 (2020).
138. Saigusa, R. et al. Single cell transcriptomics and TCR reconstruction reveal CD4 T cell response to MHC-II-restricted APOB epitope in human cardiovascular disease. *Nat. Cardiovasc. Res.* **1**, 462–475 (2022).
139. Kimura, T. et al. Regulatory CD4⁺ T cells recognize major histocompatibility complex class II molecule-restricted peptide epitopes of apolipoprotein B. *Circulation* **138**, 1130–1143 (2018).
140. Freuchet, A. et al. Identification of human exT_{reg} cells as CD16⁺CD56⁺ cytotoxic CD4⁺ T cells. *Nat. Immunol.* **24**, 1748–1761 (2023).
141. Calabretta, R. et al. Immune checkpoint inhibitor therapy induces inflammatory activity in large arteries. *Circulation* **142**, 2396–2398 (2020).
142. Drobni, Z. D. et al. Association between immune checkpoint inhibitors with cardiovascular events and atherosclerotic plaque. *Circulation* **142**, 2299–2311 (2020).
143. Suero-Abreu, G. A., Zanni, M. V. & Neilan, T. G. Atherosclerosis with immune checkpoint inhibitor therapy: evidence, diagnosis, and management: JACC: Cardiooncology State-of-the-Art Review. *JACC CardioOncol* **4**, 598–615 (2022).
144. Vuong, J. T. et al. Immune checkpoint therapies and atherosclerosis: mechanisms and clinical implications: JACC State-of-the-Art Review. *J. Am. Coll. Cardiol.* **79**, 577–593 (2022).
145. Depuydt, M. A. C. et al. Single-cell T cell receptor sequencing of paired human atherosclerotic plaques and blood reveals autoimmune-like features of expanded effector T cells. *Nat. Cardiovasc. Res.* **2**, 112–125 (2023).
146. Fernandez, D. M. & Giannarelli, C. Immune cell profiling in atherosclerosis: role in research and precision medicine. *Nat. Rev. Cardiol.* **19**, 43–58 (2022).
147. Schafer, S. & Zernecke, A. CD8⁺ T cells in atherosclerosis. *Cells* **10**, 37 (2020).
148. Dimayuga, P. C. et al. Identification of apoB-100 peptide-specific CD8⁺ T cells in atherosclerosis. *J. Am. Heart Assoc.* **6**, e005318 (2017).
149. Porsch, F., Mallat, Z. & Binder, C. J. Humoral immunity in atherosclerosis and myocardial infarction: from B cells to antibodies. *Cardiovasc. Res.* **117**, 2544–2562 (2021).
150. Centa, M. et al. Acute loss of apolipoprotein E triggers an autoimmune response that accelerates atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* **38**, e145–e158 (2018).
151. Porsch, F. & Binder, C. J. Autoimmune diseases and atherosclerotic cardiovascular disease. *Nat. Rev. Cardiol.* **21**, 780–807 (2024).
152. Mackay, F. & Schneider, P. Cracking the BAFF code. *Nat. Rev. Immunol.* **9**, 491–502 (2009).
153. Yla-Herttuala, S. et al. Rabbit and human atherosclerotic lesions contain IgG that recognizes epitopes of oxidized LDL. *Arterioscler. Thromb.* **14**, 32–40 (1994).
154. Taleb, A. et al. High immunoglobulin-M levels to oxidation-specific epitopes are associated with lower risk of acute myocardial infarction. *J. Lipid Res.* **64**, 100391 (2023).
155. Morgan-Hughes, J. A. et al. The molecular pathology of human respiratory chain defects. *Rev. Neurol.* **147**, 450–454 (1991).
156. Gruber, S. et al. Sialic acid-binding immunoglobulin-like lectin G promotes atherosclerosis and liver inflammation by suppressing the protective functions of B-1 cells. *Cell Rep.* **14**, 2348–2361 (2016).
157. Chou, M. Y. et al. Oxidation-specific epitopes are dominant targets of innate natural antibodies in mice and humans. *J. Clin. Invest.* **119**, 1335–1349 (2009).
158. Deroissart, J. & Binder, C. J. Mapping the functions of IgM antibodies in atherosclerotic cardiovascular disease. *Nat. Rev. Cardiol.* **20**, 433–434 (2023).
159. Srikakulapu, P. et al. Perivascular adipose tissue harbors atheroprotective IgM-producing B cells. *Front. Physiol.* **8**, 719 (2017).
160. Björnson, E. et al. Triglyceride-rich lipoprotein remnants, low-density lipoproteins, and risk of coronary heart disease: a UK Biobank study. *Eur. Heart J.* **44**, 4186–4195 (2023).
161. Björnson, E. et al. Lipoprotein(a) is markedly more atherogenic than LDL: an apolipoprotein B-based genetic analysis. *J. Am. Coll. Cardiol.* **83**, 385–395 (2024).
162. Ginsberg, H. N. et al. Triglyceride-rich lipoproteins and their remnants: metabolic insights, role in atherosclerotic cardiovascular disease, and emerging therapeutic strategies – a consensus statement from the European Atherosclerosis Society. *Eur. Heart J.* **42**, 4791–4806 (2021).
163. Kronenberg, F. et al. Lipoprotein(a) in atherosclerotic cardiovascular disease and aortic stenosis: a European Atherosclerosis Society consensus statement. *Eur. Heart J.* **43**, 3925–3946 (2022).
164. Sacks, F. M. & Campos, H. Clinical review 163: cardiovascular endocrinology: low-density lipoprotein size and cardiovascular disease: a reappraisal. *J. Clin. Endocrinol. Metab.* **88**, 4525–4532 (2003).
165. Chapman, M. J. et al. LDL subclass lipidomics in atherogenic dyslipidemia: effect of statin therapy on bioactive lipids and dense LDL. *J. Lipid Res.* **61**, 911–932 (2020).
166. Krauss, R. M. Small dense low-density lipoprotein particles: clinically relevant? *Curr. Opin. Lipidol.* **33**, 160–166 (2022).
167. Anber, V., Millar, J. S., McConnell, M., Shepherd, J. & Packard, C. J. Interaction of very-low-density, intermediate-density, and low-density lipoproteins with human arterial wall proteoglycans. *Arterioscler. Thromb. Vasc. Biol.* **17**, 2507–2514 (1997).
168. Varbo, A. et al. Remnant cholesterol as a causal risk factor for ischemic heart disease. *J. Am. Coll. Cardiol.* **61**, 427–436 (2013).
169. Ference, B. A. et al. Association of triglyceride-lowering LPL variants and LDL-C-lowering LDLR variants with risk of coronary heart disease. *JAMA* **321**, 364–373 (2019).
170. Packard, C. J., Boren, J. & Taskinen, M. R. Causes and consequences of hypertriglyceridemia. *Front. Endocrinol.* **11**, 252 (2020).
171. Björnson, E. et al. Quantifying triglyceride-rich lipoprotein atherogenicity, associations with inflammation, and implications for risk assessment using non-HDL cholesterol. *J. Am. Coll. Cardiol.* **84**, 1328–1338 (2024).
172. Sacks, F. M. The crucial roles of apolipoproteins E and C-III in apoB lipoprotein metabolism in normolipidemia and hypertriglyceridemia. *Curr. Opin. Lipidol.* **26**, 56–63 (2015).
173. Brown, W. V., Sacks, F. M. & Sniderman, A. D. JCL roundtable: apolipoproteins as causative elements in vascular disease. *J. Clin. Lipidol.* **9**, 733–740 (2015).
174. Boren, J., Packard, C. J. & Taskinen, M. R. The roles of ApoC-III on the metabolism of triglyceride-rich lipoproteins in humans. *Front. Endocrinol.* **11**, 474 (2020).
175. Salinas, C. A. A. & Chapman, M. J. Remnant lipoproteins: are they equal to or more atherogenic than LDL? *Curr. Opin. Lipidol.* **31**, 132–139 (2020).
176. Van Lenten, B. J. et al. Receptor-mediated uptake of remnant lipoproteins by cholesterol-loaded human monocyte-macrophages. *J. Biol. Chem.* **260**, 8783–8788 (1985).
177. Varbo, A. & Nordestgaard, B. G. Remnant lipoproteins. *Curr. Opin. Lipidol.* **28**, 300–307 (2017).
178. Navarese, E. P. et al. Independent causal effect of remnant cholesterol on atherosclerotic cardiovascular outcomes: a mendelian randomization study. *Arterioscler. Thromb. Vasc. Biol.* **43**, e373–e380 (2023).
179. Varbo, A., Benn, M., Tybjaerg-Hansen, A. & Nordestgaard, B. G. Elevated remnant cholesterol causes both low-grade inflammation and ischemic heart disease, whereas elevated low-density lipoprotein cholesterol causes ischemic heart disease without inflammation. *Circulation* **128**, 1298–1309 (2013).

180. Wadstrom, B. N., Pedersen, K. M., Wulff, A. B. & Nordestgaard, B. G. Inflammation compared to low-density lipoprotein cholesterol: two different causes of atherosclerotic cardiovascular disease. *Curr. Opin. Lipidol.* **34**, 96–104 (2023).
181. Zewinger, S. et al. Apolipoprotein C3 induces inflammation and organ damage by alternative inflammasome activation. *Nat. Immunol.* **21**, 30–41 (2020).
182. Schwartz, E. A. & Reaven, P. D. Lipolysis of triglyceride-rich lipoproteins, vascular inflammation, and atherosclerosis. *Biochim. Biophys. Acta* **1821**, 858–866 (2012).
183. Higgins, L. J. & Rutledge, J. C. Inflammation associated with the postprandial lipolysis of triglyceride-rich lipoproteins by lipoprotein lipase. *Curr. Atheroscler. Rep.* **11**, 199–205 (2009).
184. De Caterina, R., Liao, J. K. & Libby, P. Fatty acid modulation of endothelial activation. *Am. J. Clin. Nutr.* **71**, 213S–223S (2000).
185. Cabodevilla, A. G. et al. Eruptive xanthoma model reveals endothelial cells internalize and metabolize chylomicrons, leading to extravascular triglyceride accumulation. *J. Clin. Invest.* **131**, e145800 (2021).
186. Goldberg, I. J., Cabodevilla, A. G. & Younis, W. In the beginning, lipoproteins cross the endothelial barrier. *J. Atheroscler. Thromb.* **31**, 854–860 (2024).
187. Kontush, A. & Chapman, M. J. Lipidomics as a tool for the study of lipoprotein metabolism. *Curr. Atheroscler. Rep.* **12**, 194–201 (2010).
188. Mucinski, J. M. et al. Relationships between very low-density lipoproteins-ceramides, -diacylglycerols, and -triacylglycerols in insulin-resistant men. *Lipids* **55**, 387–393 (2020).
189. Nieddu, G. et al. Molecular characterization of plasma HDL, LDL, and VLDL lipids cargos from atherosclerotic patients with advanced carotid lesions: a preliminary report. *Int. J. Mol. Sci.* **23**, 12449 (2022).
190. Nordestgaard, B. G., & Tybjaerg-Hansen, A. IDL, VLDL, chylomicrons and atherosclerosis. *Eur. J. Epidemiol.* **8**, 92–98 (1992).
191. Boutagy, N. E. et al. Dynamic metabolism of endothelial triglycerides protects against atherosclerosis in mice. *J. Clin. Invest.* **134**, e170453 (2024).
192. Jaffe, I. Z. & Karumanchi, S. A. Lipid droplets in the endothelium: the missing link between metabolic syndrome and cardiovascular disease? *J. Clin. Invest.* **134**, e176347 (2024).
193. Kim, B. et al. Endothelial lipid droplets suppress eNOS to link high fat consumption to blood pressure elevation. *J. Clin. Invest.* **133**, e173160 (2023).
194. Doi, H. et al. Remnant lipoproteins induce proatherothrombotic molecules in endothelial cells through a redox-sensitive mechanism. *Circulation* **102**, 670–676 (2000).
195. de Sousa, J. C. et al. Association between coagulation factors VII and X with triglyceride rich lipoproteins. *J. Clin. Pathol.* **41**, 940–944 (1988).
196. Tsimikas, S., Moriarty, P. M. & Stroes, E. S. Emerging RNA therapeutics to lower blood levels of Lp(a): JACC Focus Seminar 2/4. *J. Am. Coll. Cardiol.* **77**, 1576–1589 (2021).
197. Reyes-Soffer, G. et al. Lipoprotein(a): a genetically determined, causal, and prevalent risk factor for atherosclerotic cardiovascular disease: a scientific statement from the American Heart Association. *Arterioscler. Thromb. Vasc. Biol.* **42**, e48–e60 (2022).
198. Burgess, S. et al. Association of LPA variants with risk of coronary disease and the implications for lipoprotein(a)-lowering therapies: a mendelian randomization analysis. *JAMA Cardiol.* **3**, 619–627 (2018).
199. Madsen, C. M., Kamstrup, P. R., Langsted, A., Varbo, A. & Nordestgaard, B. G. Lipoprotein(a)-lowering by 50 mg/dL (105 nmol/L) may be needed to reduce cardiovascular disease 20% in secondary prevention: a population-based study. *Arterioscler. Thromb. Vasc. Biol.* **40**, 255–266 (2020).
200. Marston, N. A. et al. Per-particle cardiovascular risk of lipoprotein(a) vs Non-Lp(a) apolipoprotein B-containing lipoproteins. *J. Am. Coll. Cardiol.* **83**, 470–472 (2024).
201. Bjornson, E., Adiels, M., Boren, J. & Packard, C. J. Lipoprotein(a) is a highly atherogenic lipoprotein: pathophysiological basis and clinical implications. *Curr. Opin. Cardiol.* **39**, 503–510 (2024).
202. Boffa, M. B. & Koschinsky, M. L. Oxidized phospholipids as a unifying theory for lipoprotein(a) and cardiovascular disease. *Nat. Rev. Cardiol.* **16**, 305–318 (2019).
203. Rader, D. J. & Bajaj, A. Lipoprotein(a) and oxidized phospholipids: partners in crime or individual perpetrators in cardiovascular disease? *J. Am. Coll. Cardiol.* **81**, 1793–1796 (2023).
204. D'Angelo, A. et al. The apolipoprotein(a) component of lipoprotein(a) mediates binding to laminin: contribution to selective retention of lipoprotein(a) in atherosclerotic lesions. *Biochim. Biophys. Acta* **1687**, 1–10 (2005).
205. McLean, J. W. et al. cDNA sequence of human apolipoprotein(a) is homologous to plasminogen. *Nature* **330**, 132–137 (1987).
206. Koschinsky, M. L., Stroes, E. S. G. & Kronenberg, F. Daring to dream: targeting lipoprotein(a) as a causal and risk-enhancing factor. *Pharmacol. Res.* **194**, 106843 (2023).
207. Boffa, M. B. & Koschinsky, M. L. Lipoprotein (a): truly a direct prothrombotic factor in cardiovascular disease? *J. Lipid Res.* **57**, 745–757 (2016).
208. Boffa, M. B. et al. Potent reduction of plasma lipoprotein (a) with an antisense oligonucleotide in human subjects does not affect ex vivo fibrinolysis. *J. Lipid Res.* **60**, 2082–2089 (2019).
209. von Depka, M. et al. Increased lipoprotein (a) levels as an independent risk factor for venous thromboembolism. *Blood* **96**, 3364–3368 (2000).
210. Nowak-Gottl, U. et al. Increased lipoprotein(a) is an important risk factor for venous thromboembolism in childhood. *Circulation* **100**, 743–748 (1999).
211. Marcucci, R. et al. Increased plasma levels of lipoprotein(a) and the risk of idiopathic and recurrent venous thromboembolism. *Am. J. Med.* **115**, 601–605 (2003).
212. Vormittag, R. et al. Lipoprotein (a) in patients with spontaneous venous thromboembolism. *Thromb. Res.* **120**, 15–20 (2007).
213. Kronenberg, F. et al. Frequent questions and responses on the 2022 lipoprotein(a) consensus statement of the European Atherosclerosis Society. *Atherosclerosis* **374**, 107–120 (2023).
214. Bergmark, C. et al. A novel function of lipoprotein [a] as a preferential carrier of oxidized phospholipids in human plasma. *J. Lipid Res.* **49**, 2230–2239 (2008).
215. Leibundgut, G. et al. Determinants of binding of oxidized phospholipids on apolipoprotein (a) and lipoprotein (a). *J. Lipid Res.* **54**, 2815–2830 (2013).
216. Nie, J., Yang, J., Wei, Y. & Wei, X. The role of oxidized phospholipids in the development of disease. *Mol. Asp. Med.* **76**, 100909 (2020).
217. Assini, J. M. et al. High levels of lipoprotein(a) in transgenic mice exacerbate atherosclerosis and promote vulnerable plaque features in a sex-specific manner. *Atherosclerosis* **384**, 117150 (2023).
218. Dzobo, K. E. et al. Diacylglycerols and lysophosphatidic acid, enriched on lipoprotein(a), contribute to monocyte inflammation. *Arterioscler. Thromb. Vasc. Biol.* **44**, 720–740 (2024).
219. Mach, F. et al. 2019 ESC/EAS guidelines for the management of dyslipidaemias: lipid modification to reduce cardiovascular risk. *Eur. Heart J.* **41**, 111–188 (2020).
220. Cholesterol Treatment Trialists' (CTT) Collaboration Efficacy and safety of LDL-lowering therapy among men and women: meta-analysis of individual data from 174,000 participants in 27 randomised trials. *Lancet* **385**, 1397–1405 (2015).
221. Tsimikas, S. Lipoprotein(a) in the year 2024: a look back and a look ahead. *Arterioscler. Thromb. Vasc. Biol.* **44**, 1485–1490 (2024).
222. Kim, N. H. & Kim, S. G. Fibrates revisited: potential role in cardiovascular risk reduction. *Diabetes Metab. J.* **44**, 213–221 (2020).
223. Das Pradhan, A. et al. Triglyceride lowering with pemafibrate to reduce cardiovascular risk. *N. Engl. J. Med.* **387**, 1923–1934 (2022).
224. Doi, T., Langsted, A. & Nordestgaard, B. G. Remnant cholesterol, LDL cholesterol, and apoB absolute mass changes explain results of the PROMINENT trial. *Atherosclerosis* **393**, 117556 (2024).
225. Tokgozoglul, L., Pirillo, A. & Catapano, A. L. Disconnect between triglyceride reduction and cardiovascular outcomes: lessons from the PROMINENT and CLEAR Outcomes trials. *Eur. Heart J.* **45**, 2377–2379 (2024).

Author contributions

The authors contributed substantially to all aspects of the article.

Competing interests

The authors declare no competing interests.

Additional information

Peer review information *Nature Reviews Cardiology* thanks Katarina Öörni and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© Springer Nature Limited 2025