

Published in final edited form as:

Science. 2013 January 11; 339(6116): 211–214. doi:10.1126/science.1227166.

Suppression of Oxidative Stress by β -Hydroxybutyrate, an Endogenous Histone Deacetylase Inhibitor

Tadahiro Shimazu^{1,2}, Matthew D. Hirshey^{1,2}, John Newman^{1,2}, Wenjuan He^{1,2}, Kotaro Shirakawa^{1,2}, Natacha Le Moan³, Carrie A. Grueter^{4,5}, Hyungwook Lim^{1,2}, Laura R. Saunders^{1,2}, Robert D. Stevens⁶, Christopher B. Newgard⁶, Robert V. Farese Jr.^{2,4,5}, Rafael de Cabo⁷, Scott Ulrich⁸, Katerina Akassoglou³, and Eric Verdin^{1,2,*}

¹Gladstone Institute of Virology and Immunology, San Francisco, CA 94158, USA

²Department of Medicine, University of California, San Francisco, CA 94143, USA

³Gladstone Institute of Neurological Disease, San Francisco, CA 94158, USA

⁴Gladstone Institute of Cardiovascular Disease, San Francisco, CA 94158, USA

⁵Department of Biochemistry and Biophysics, University of California, San Francisco, CA 94143, USA

⁶Sarah W. Stedman Nutrition and Metabolism Center, and Departments of Pharmacology and Cancer Biology and Medicine, Duke University Medical Center, Durham, NC 27704, USA

⁷Laboratory of Experimental Gerontology, National Institute on Aging, National Institutes of Health, Baltimore, MD 21224, USA

⁸Department of Chemistry, Ithaca College, Ithaca, NY 14850, USA

Abstract

Concentrations of acetyl-coenzyme A and nicotinamide adenine dinucleotide (NAD⁺) affect histone acetylation and thereby couple cellular metabolic status and transcriptional regulation. We report that the ketone body D- β -hydroxybutyrate (β OHB) is an endogenous and specific inhibitor of class I histone deacetylases (HDACs). Administration of exogenous β OHB, or fasting or calorie restriction, two conditions associated with increased β OHB abundance, all increased global histone acetylation in mouse tissues. Inhibition of HDAC by β OHB was correlated with global changes in transcription, including that of the genes encoding oxidative stress resistance factors FOXO3A and MT2. Treatment of cells with β OHB increased histone acetylation at the *Foxo3a* and *Mt2* promoters, and both genes were activated by selective depletion of HDAC1 and HDAC2. Consistent with increased FOXO3A and MT2 activity, treatment of mice with β OHB conferred substantial protection against oxidative stress.

Cellular metabolites such as acetyl-coenzyme A (acetyl-CoA) and nicotinamide adenine dinucleotide (NAD⁺) influence gene expression by serving as cofactors for epigenetic modifiers that mediate posttranslational modification of histones (1). The activity of histone acetyltransferases (HATs) is dependent on nuclear acetyl-CoA concentrations (2, 3) and the

*To whom correspondence should be addressed. everdin@gladstone.ucsf.edu.

Supplementary Materials: <http://www.sciencemag.org/cgi/content/full/science.1227166/DC1>

Materials and Methods

Figs. S1 to S15

Tables S1 to S3

References (23–26)

deacetylase activity of class III HDACs, also called sirtuins, is dependent on NAD⁺ concentrations (4). Class I (HDAC1, 2, 3, 8), class II (HDAC4, 5, 6, 7, 9, 10), and class IV (HDAC11) HDACs are zinc-dependent enzymes, but endogenous regulators are not known.

Small-molecule inhibitors of class I and class II HDACs include butyrate, a product of bacterial anaerobic fermentation (5). Butyrate is closely related to β -hydroxybutyrate (β OHB) (Fig. 1A), the major source of energy for mammals during prolonged exercise or starvation (6). Accumulation of β OHB in blood increases to 1 to 2 mM during fasting when the liver switches to fatty acid oxidation (7, 8), and to even higher concentrations during prolonged fasting (6 to 8 mM) (6) or in diabetic ketoacidosis (>25 mM) (9).

To determine whether β OHB might have HDAC inhibitor activity, we treated human embryonic kidney 293 (HEK293) cells with different amounts of β OHB for 8 hours, and measured histone acetylation levels by Western blot with antibodies to acetylated histone H3 lysine 9 (AcH3_{K9}) and to acetylated histone H3 lysine 14 (AcH3_{K14}) (Fig. 1, B and C, and figs. S1 and S2). β OHB increased histone acetylation in a dose-dependent manner, even at 1 to 2 mM, which can occur in humans after a 2- to 3-day fast or strenuous exercise (6, 8, 10). Like butyrate, β OHB did not increase acetylation of α -tubulin, indicating that it inhibits class I HDACs but not the class IIb tubulin deacetylase, HDAC6.

To test the HDAC inhibitor activity of β OHB and its possible selectivity, we purified recombinant human HDACs after transient transfection of expression vectors for human epitope-tagged (FLAG) HDAC1, HDAC3, HDAC4, and HDAC6 in HEK293T cells. We purified the HDACs, incubated them with ³H-labeled acetylated histone H4 peptides, and measured their deacetylase activity (Fig. 1D) (11). β OHB inhibited HDAC1, HDAC3, and HDAC4 in a dose-dependent manner with a median inhibitory concentration (IC₅₀) of 5.3, 2.4, and 4.5 mM, respectively. The HDAC6 IC₅₀ was much higher (48.5 mM) (Fig. 1E), and β OHB did not inhibit HDAC6 activity on its natural substrate tubulin (fig. S3). To examine the possibility that histone acetylation was enhanced by increased concentration of acetyl-CoA (because β OHB is catabolized into acetyl-CoA in target tissues), we directly measured abundance of acetyl-CoA in β OHB-treated HEK293 cells, but no change was observed (fig. S4). We also tested a possible activating effect of β OHB on histone acetyltransferase activity of p300 and PCAF (P300/CBP-associated factor) but did not detect any change induced by β OHB (fig. S5). Thus, millimolar concentrations of β OHB appear to increase histone acetylation directly through HDAC inhibition. High concentrations of acetoacetate (AcAc) also inhibit class I and class IIa HDACs in vitro (fig. S6A) and in HEK293 cells (fig. S6B). However, the concentration of AcAc in blood is one-third or less than that of β OHB during fasting and therefore less likely to reach concentrations that would inhibit HDACs under physiological conditions (12). To test the relative contribution of β OHB, acetoacetate, and acetyl-CoA to histone acetylation in response to β OHB treatment, we depleted cells of β OHB dehydrogenases (BDH1, 2) with small interfering RNA. Both enzymes catalyze the transformation of β OHB into acetoacetate, and their suppression had no effect on histone acetylation in response to β OHB up to 3mM. At higher concentrations of β OHB (10 and 30 mM), however, further increase in histone acetylation was suppressed by depletion of β OHB dehydrogenases (fig. S7, A and B), indicating that either AcAc or acetyl-CoA might also contribute to histone acetylation in cells exposed to concentrations of β OHB above the IC₅₀ for HDACs.

To determine whether changes in β OHB concentrations in vivo might affect histone acetylation, we measured β OHB concentration in mouse serum after a 24-hour fast or in mice on calorie restriction (CR). We also used implanted osmotic pumps to administer exogenous β OHB or phosphate-buffered saline (PBS). β OHB concentrations increased to 1.5 \pm 0.1 mM after a 24-hour fast, 0.6 \pm 0.1 mM in mice on CR and 1.2 \pm 0.1 mM with

administration of β OHB via an intraperitoneal pump (Fig. 2A). We collected tissues from fed or 24-hour-fasted mice and measured histone acetylation by immunoblotting. Acetylation of H3_{K9} and H3_{K14} reflect the competing activities of HATs and HDACs and influence gene expression in several species, including humans (13). Acetylation of H3_{K9} and H3_{K14} increased significantly in several organs in fasted mice, particularly kidney (Fig. 2B and figs. S8 and S9). In kidney, histone acetylation (H3_{K9} and H3_{K14}) also increased two- to fivefold in mice under CR. Kidney histone acetylation and serum β OHB concentrations were strongly correlated for both histone H3_{K9} ($R^2 = 0.772$) and histone H3_{K14} ($R^2 = 0.863$) (Fig. 2C). We first focused on kidney, the organ with the largest changes in histone acetylation, to investigate the effects of β OHB on gene expression and cellular phenotype.

Histone acetylation induced by HDAC inhibitors is associated with transcriptional activation and repression of a subset of cellular genes (14). To identify genes whose expression changed in response to β OHB, we extracted mRNA for microarray analysis from mouse kidneys treated with β OHB or PBS for 24 hours. As β OHB is more abundant in fasting conditions, gene expression changes induced by β OHB may be a subset of those induced by fasting. Of 35,556 genes tested, 284 increased transcription in response to fasting (false discovery rate <0.2, table S1). Four of the five genes with the largest changes in expression in response to β OHB were also activated in response to fasting as determined both by microarray and quantitative real-time polymerase chain reaction (QPCR) ($P < 0.001$ for such overlap via binomial distribution, tables S2 and S3). Ingenuity pathway analysis identified two of the five β OHB-induced genes (*Mt2* and *Lcn2*) as regulated by FOXO3A. Overall, we found five genes in the FOXO3A network (*Foxo3a*, *Mt2*, *Lcn2*, *Lemd3*, and *Hbp1*) that had increased transcription in response to β OHB via QPCR (fig. S10). *Foxo3a*, a transcription factor, induces cell-cycle arrest and resistance to oxidative stress (15). Metallothionein 2 (*Mt2*), the gene with the greatest transcriptional response to β OHB treatment as determined by QPCR, also protects against oxidative stress (16). *Foxo3a* and *Mt2* mRNA expression is also modestly increased during CR as determined by QPCR (Fig. 3, A and B). Chromatin immunoprecipitation (ChIP) analysis of the *Foxo3a* and *Mt2* promoters with two distinct primer pairs for each promoter revealed increased histone H3_{K9} acetylation at both promoters after treatment of HEK293 cells with a high dose of β OHB (10 mM) (Fig. 3C).

Next, we depleted cells of each class I and II HDAC with selective short hairpin-mediated RNAs (shRNAs). The shRNAs selective for each HDAC suppressed expression of their cognate HDAC by at least 60% (fig. S11). Depletion of HDAC1 or HDAC2 caused up-regulation of *Foxo3a* and *Mt2* mRNA by 1.8-fold and 1.5-fold, respectively (Fig. 3D). Depletion of both HDAC1 and 2 further increased accumulation of *Foxo3a* and *Mt2* mRNA (Fig. 3D). ChIP analysis revealed that HDAC1, but not HDAC6, was recruited to the *Foxo3a* and *Mt2* promoters and not recruited to the *Gapdh* promoter (Fig. 3E). Binding of HDAC1 to the *Foxo3a* promoter was unchanged by β OHB treatment; thus, HDAC catalytic activity appears not to be necessary for promoter binding of HDAC1 (fig. S12). β OHB appears to induce local histone acetylation at the promoter of oxidative stress resistance genes, *Foxo3a* and *Mt2*, by inhibiting activity of HDACs 1 and 2.

Mitochondrial superoxide dismutase (Mn-SOD) and catalase are two other well-defined FOXO3A targets that contribute to its protective activity against oxidative stress (15, 17). Protein immunoblotting of kidney tissue isolated from PBS- or β OHB-treated mice showed increased expression of FOXO3A, Mn-SOD, and catalase (Fig. 4A and figs. S13 and S14).

The effect on expression of MT2, FOXO3A, MnSOD, and catalase indicated that β OHB might have protective activity against oxidative stress. Carbonyl derivatives are formed by a direct metal-catalyzed oxidative attack on the amino acid side chains of proline, arginine,

lysine, and threonine. Carbonylation is irreversible and unrepairable and accumulates as organisms age (18). To test the possible protective role of β OHB against oxidative stress, we implanted mice with a subcutaneous pump delivering either β OHB or PBS for 24 hours. They then received an intravenous injection of paraquat, which induces accumulation of reactive oxygen species. Kidney tissue was isolated after 2 hours, and protein carbonylation was assayed by protein immunoblotting with an antibody to dinitrophenyl (DNP) after derivatization of protein with dinitrophenylhydrazine (DNPH) (Fig. 4B). Paraquat treatment of control mice receiving a PBS infusion led to a twofold increase in carbonylated proteins. This increase in protein carbonylation was significantly suppressed ($54 \pm 9\%$ decrease) in mice receiving β OHB (Fig. 4C).

We also examined another marker of oxidative stress: lipid peroxidation. 4-Hydroxynonenal (4-HNE) is a degradation product of polyunsaturated lipid and accumulates in response to oxidative stress (19). Kidney tissue sections from PBS- or paraquat-treated mice were stained with an antibody to 4-HNE, and the amount of 4-HNE staining was quantified with imaging software (fig. S15). Paraquat treatment increased 4-HNE staining threefold in control mice (PBS) (Fig. 4D). This increase was completely suppressed in mice treated with β OHB. Lipid peroxides were also directly quantified in an enzyme-linked immunosorbent assay that measures conversion of ferrous ions to ferric ions. A twofold increase in lipid peroxide in response to paraquat was suppressed significantly by β OHB treatment (Fig. 4E). Thus, β OHB protects against paraquat-induced oxidative stress in mouse kidney.

Our observation that β OHB is an endogenous HDAC inhibitor present in organisms at millimolar concentrations during prolonged fasting and CR reveals an example of integration between metabolic status and epigenetic changes. We show that changes in histone acetylation and gene expression caused by β OHB promote stress resistance in the kidney. Future studies should investigate the specific gene expression and physiological effects of β OHB in other tissues. For example, low-carbohydrate diets that induce substantial ketogenesis are broadly neuroprotective and enhance resistance of neurons to oxidative damage (20). In addition, reduction in HDAC activity by either genetic manipulation or chemical inhibition extends life span in *Drosophila* (21, 22). Inhibition of HDACs by β OHB might contribute to the beneficial effect of ketogenic diets and may be one mechanism by which calorie restriction confers health benefits.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

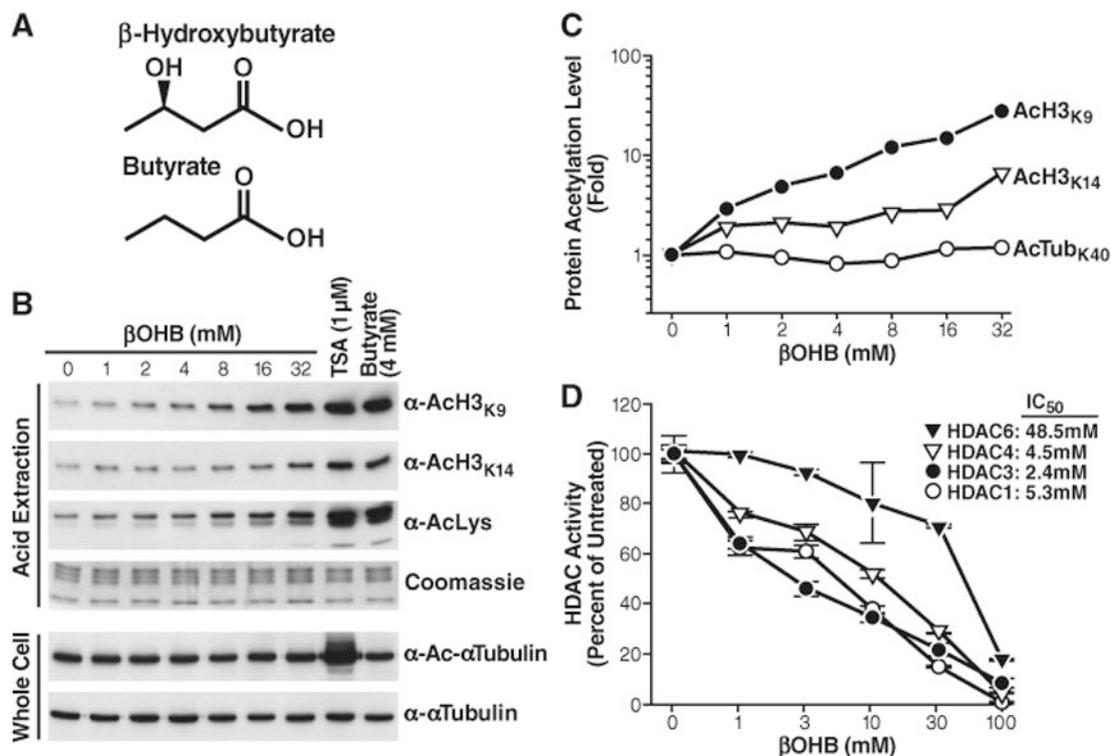
Acknowledgments

We thank J. Carroll for graphics, G. Howard and S. Ordway for editorial assistance, V. Fonseca for administrative assistance, and A. Brunet (Stanford University) for reagents. Supported by funds from the Gladstone Institutes.

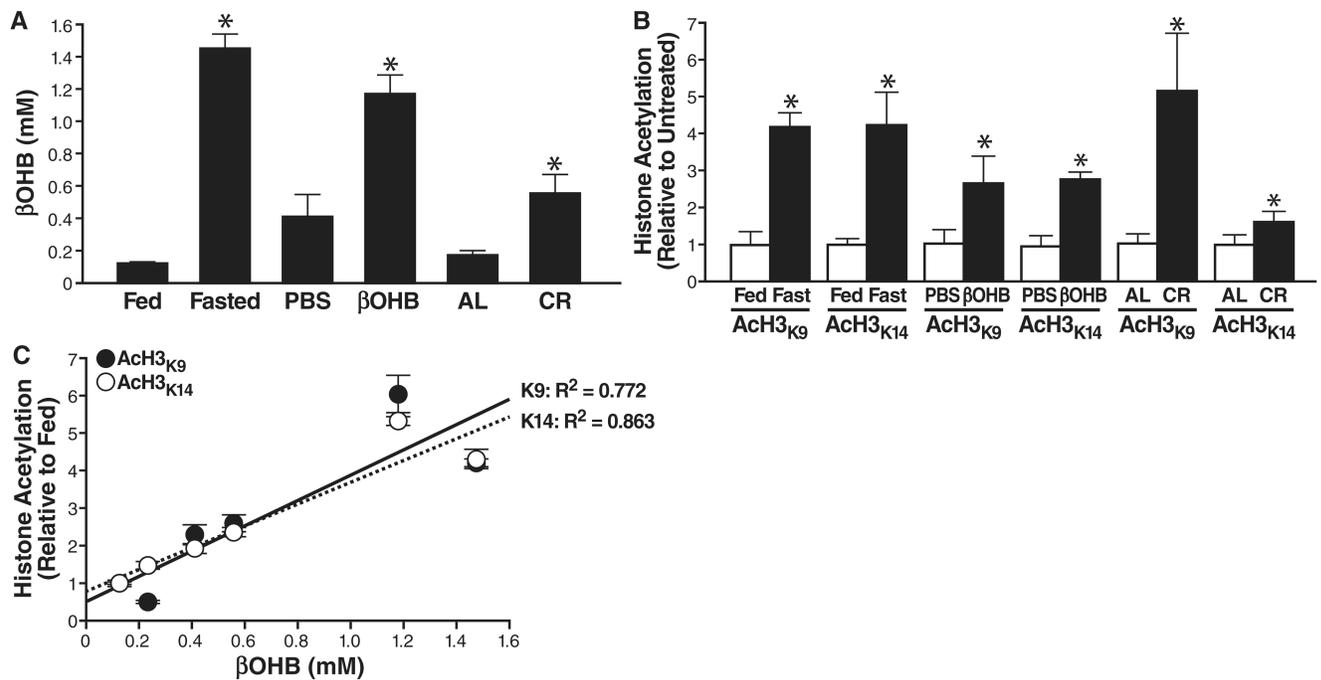
References and Notes

1. Ladurner AG. Mol Cell. 2006; 24:1. [PubMed: 17018288]
2. Wellen KE, et al. Science. 2009; 324:1076. [PubMed: 19461003]
3. Takahashi H, McCaffery JM, Irizarry RA, Boeke JD. Mol Cell. 2006; 23:207. [PubMed: 16857587]
4. Imai S, Armstrong CM, Kaerberlein M, Guarente L. Nature. 2000; 403:795. [PubMed: 10693811]
5. Candido EP, Reeves R, Davie JR. Cell. 1978; 14:105. [PubMed: 667927]
6. Cahill GF Jr. Annu Rev Nutr. 2006; 26:1. [PubMed: 16848698]
7. Cahill GF Jr, et al. J Clin Invest. 1966; 45:1751. [PubMed: 5926444]
8. Robinson AM, Williamson DH. Physiol Rev. 1980; 60:143. [PubMed: 6986618]

9. Laffel L. *Diabetes Metab Res Rev*. 1999; 15:412. [PubMed: 10634967]
10. Koeslag JH, Noakes TD, Sloan AW. *J Physiol*. 1980; 301:79. [PubMed: 6997456]
11. North BJ, Schwer B, Ahuja N, Marshall B, Verdin E. *Methods*. 2005; 36:338. [PubMed: 16091304]
12. Owen OE, et al. *J Clin Invest*. 1973; 52:2606. [PubMed: 4729054]
13. Agalioti T, et al. *Cell*. 2000; 103:667. [PubMed: 11106736]
14. Van Lint C, Emiliani S, Verdin E. *Gene Expr*. 1996; 5:245. [PubMed: 8723390]
15. Kops GJ, et al. *Nature*. 2002; 419:316. [PubMed: 12239572]
16. Kawai K, et al. *Chem Res Toxicol*. 2000; 13:1275. [PubMed: 11123969]
17. Nemoto S, Finkel T. *Science*. 2002; 295:2450. [PubMed: 11884717]
18. Nyström T. *EMBO J*. 2005; 24:1311. [PubMed: 15775985]
19. Esterbauer H, Schaur RJ, Zollner H. *Free Radic Biol Med*. 1991; 11:81. [PubMed: 1937131]
20. Kim Y, et al. *J Neurochem*. 2007; 101:1316. [PubMed: 17403035]
21. Rogina B, Helfand SL, Frankel S. *Science*. 2002; 298:1745. [PubMed: 12459580]
22. Kang HL, Benzer S, Min KT. *Proc Natl Acad Sci USA*. 2002; 99:838. [PubMed: 11792861]

**Fig. 1.**

Inhibition of HDACs by β OHB in vitro and in vivo. **(A)** Structures of β -hydroxybutyrate and butyrate. **(B)** Effect of β OHB, TSA, or butyrate on acetylation of histone H3 and tubulin. HEK293 cells were treated with the indicated concentrations of drugs for 8 hours. Histones were acid-extracted, and their acetylation was assessed by protein immunoblotting with anti-AcH3_{K9}, anti-AcH3_{K14}, or anti-acetyllysine (AcLys). Proteins from whole-cell extracts were analyzed by immunoblotting with antibodies to α -tubulin or Ac- α -Tubulin. **(C)** Quantification of acetylation levels from blots in **(B)**, shown relative to untreated cells (β OHB 0 mM). **(D)** Inhibition of immunopurified HDACs by β OHB in vitro. Flag-tagged HDACs were expressed in HEK293 cells, immunoprecipitated, and incubated in vitro with a ³H-labeled acetylated histone H4 peptide and the indicated concentrations of β OHB. HDAC activity is relative to the activity of each enzyme without β OHB. The IC₅₀ values of β OHB are shown.

**Fig. 2.**

Serum β OHB and global histone acetylation in kidney of mice deprived of food. **(A)** Serum concentration of β OHB was determined in age-matched groups of three C57Bl6 mice fed or fasted (16 weeks old, fasted for 24 hours), implanted with a pump delivering either PBS or β OHB (16 weeks old, 24 hours of pump treatment), fed ad libitum (AL) or on calorie restriction to 60% of AL (CR) (8 months old, 6 months on CR). Mean \pm SE, * P < 0.05 by t test between paired conditions. **(B)** Histones were purified from the kidneys of the same mice as in (A). Acetylation was assessed with anti-Ach3_{K9}, anti-Ach3_{K14}, or anti-Ac- α -tubulin (fig. S8). Acetylation is normalized to total histone H3 and shown relative to the control condition in each pair (e.g., fed versus fasted). Mean \pm SE, * P < 0.05 by t test between paired conditions. **(C)** Serum β OHB concentrations [from (A)] plotted against acetylation of histone H3_{K9} and H3_{K14} [from (B)]; correlation coefficient $R^2 = 0.772$ for H3_{K9} and 0.863 for H3_{K14}.

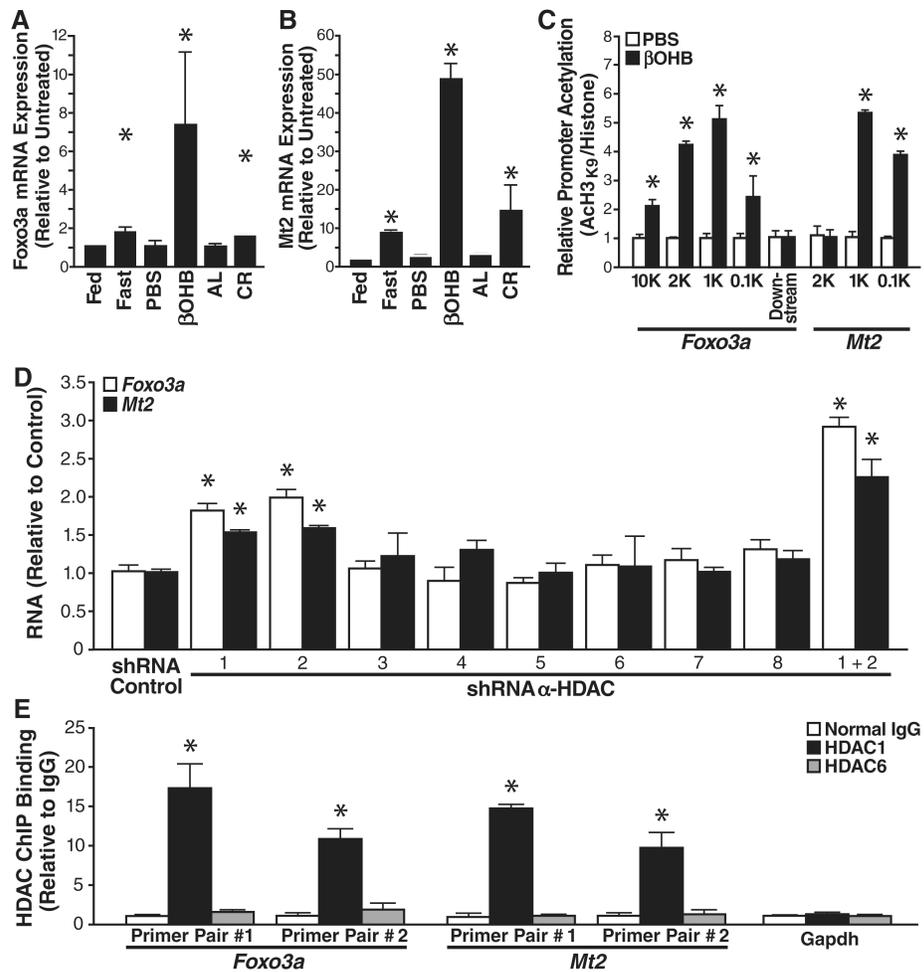
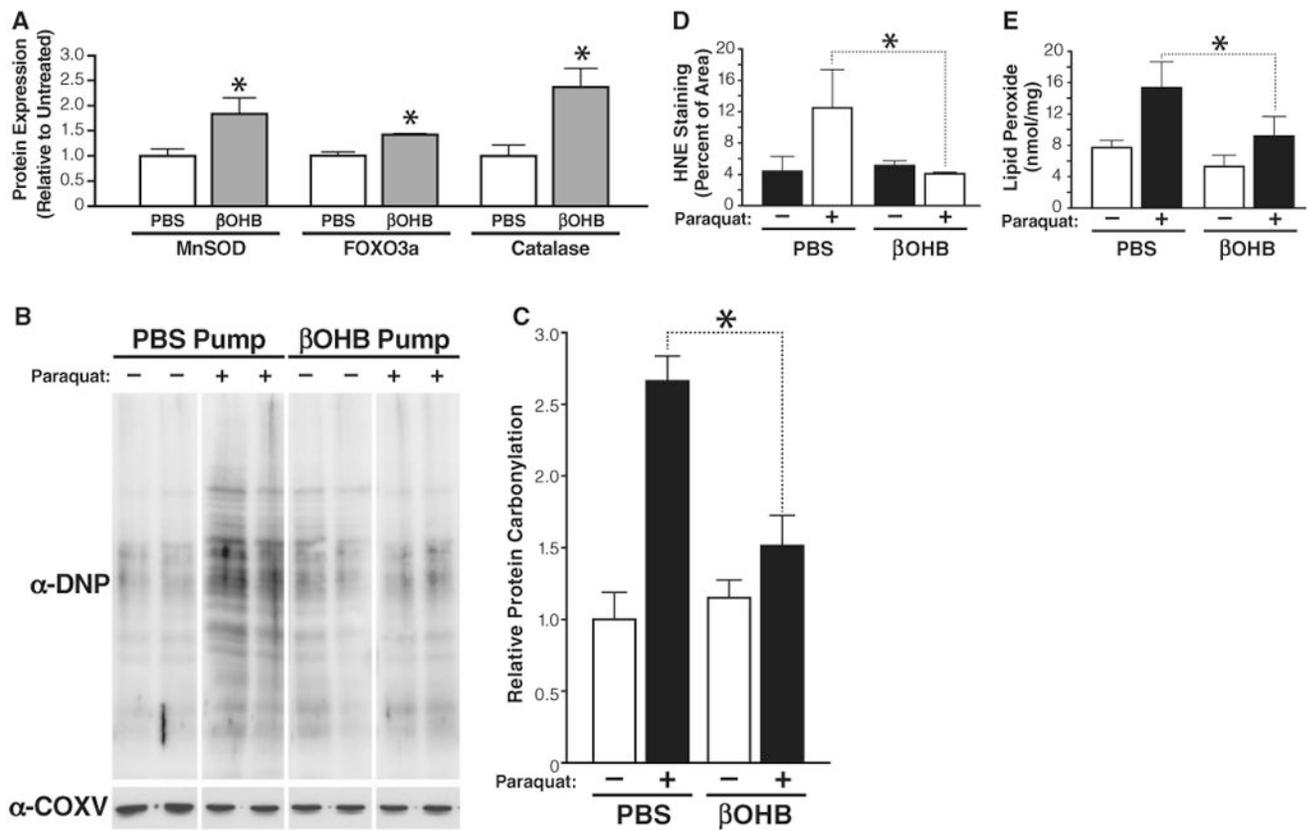


Fig. 3. Increased expression of oxidative stress resistance genes in cells exposed to β OHB. **(A)** Expression of *Foxo3a* under various conditions (see Fig. 2 for details) measured by QPCR. *Foxo3a* expression is normalized to abundance of glyceraldehyde-3-phosphate dehydrogenase (GAPDH). Mean \pm SE, * P < 0.05 by t test between paired conditions. **(B)** Expression of *Mt2*, measured as in (A). **(C)** Promoters of *Mt2* and *Foxo3a* are enriched for acetylated histone H3K9 after β OHB treatment. HEK293 cells were treated with 10 mM β OHB or PBS for 24 hours. Chromatin was immunoprecipitated with anti-H3 or anti-AcH3_{K9}, and the purified DNA was analyzed with primer pairs specific for the *Foxo3a* or *Mt2* promoters. Results are the ratios of AcH3_{K9} to total histone H3. Mean \pm SE, * P < 0.05 by t test between β OHB and PBS conditions. **(D)** HDAC depletion increases *Foxo3a* and *Mt2* mRNAs abundance. HEK293 cells were transfected with shRNAs specific for each class I or class II HDAC, and mRNA abundance was measured by QPCR 72 hours after transfection. Mean \pm SE, * P < 0.05 by t test versus control shRNA. **(E)** HDAC1, but not HDAC6, is enriched at the promoters of *Mt2* and *Foxo3a*. ChIP analysis of the *Foxo3a* and *Mt2* promoters (two primer pairs per promoter) and Gapdh (one primer pair) from HEK293 cells with control immunoglobulin G (IgG), anti-HDAC1, or anti-HDAC6. Relative promoter binding of each HDAC is normalized to input Gapdh. Mean \pm SE, * P < 0.05 by t test versus IgG control.

**Fig. 4.**

Protective effect of β OHB treatment against oxidative stress. **(A)** Amounts of catalase, MnSOD, or FOXO3A measured by protein immunoblotting in kidney tissue from 16-week-old mice implanted with an osmotic pump delivering PBS or β OHB (as in Fig. 2; $n = 3$); mean \pm SE, * $P < 0.05$ by t test between PBS and β OHB conditions. **(B)** Protein carbonylation in kidney samples from mice implanted with an osmotic pump delivering PBS or β OHB (as in Fig. 2; $n = 3$) and treated with paraquat (50 mg/kg) or vehicle for 2 hours. Carbonylation was measured by immunoblotting with anti-DNP. All samples were run on a single gel; after imaging, lanes were rearranged for presentation. **(C)** Quantification of protein carbonylation in (B). Mean \pm SE, * $P < 0.05$ by t test between PBS and β OHB conditions. **(D)** Sections of kidney obtained from the same mice as in (B) were stained with anti-4-HNE and quantified (see fig. S16 for primary picture). Mean \pm SE, * $P < 0.05$ by t test between PBS and β OHB conditions. **(E)** Lipid peroxides were quantified in mice kidneys (LPO assay kit, Cayman, Ann Arbor, MI). Mean \pm SE, * $P < 0.05$ by t test between PBS and β OHB conditions.