



REVIEW

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Deciphering the roles of macrophages in developmental and inflammation stimulated lymphangiogenesis

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Abstract

Lymphatic vessels share an intimate relationship with hematopoietic cells that commences during embryogenesis and continues throughout life. Lymphatic vessels provide a key conduit for immune cell trafficking during immune surveillance and immune responses and in turn, signals produced by immune lineage cells in settings of inflammation regulate lymphatic vessel growth and activity. In the majority of cases, the recruitment and activation of immune cells during inflammation promotes the growth and development of lymphatic vessels (lymphangiogenesis) and enhances lymph flow, effects that amplify cell trafficking to local lymph nodes and facilitate the mounting of effective immune responses. Macrophages comprise a major, heterogeneous lineage of immune cells that, in addition to key roles in innate and adaptive immunity, perform diverse tasks important for tissue development, homeostasis and repair. Here, we highlight the emerging roles of macrophages in lymphangiogenesis, both during development and in settings of pathology. While much attention has focused on the production of pro-lymphangiogenic stimuli including VEGF-C and VEGF-D by macrophages in models of inflammation including cancer, there is ample evidence to suggest that macrophages provide additional signals important for the regulation of lymphatic vascular growth, morphogenesis and function.

Keywords: Lymphangiogenesis, Macrophages, Monocytes, Development, Inflammation, VEGF-C, VEGF-D

The many faces of macrophages

Macrophages encompass a phenotypically heterogeneous population of cells that play a rapidly expanding catalogue of roles during development, homeostasis and disease [1-3]. While perhaps best recognised for the key roles they fulfil in innate and adaptive immunity, macrophages (literally “large eaters”, due to their phagocytic capabilities) also provide apoptosis inducing stimuli important for tissue remodeling and maturation [4-6], cues that instruct organ patterning and morphogenesis [7-12] and signals important for tissue regeneration and repair [13-15].

Macrophage diversity is obvious both in the embryo and the adult. In the embryo, macrophage subtypes can be distinguished on the basis of differential expression of markers including lymphatic vessel endothelial

hyaluronan receptor (LYVE-1) and the angiopoietin receptor Tie2 [16,17]. In the adult, distinct populations of circulating monocytes are categorised as “inflammatory” or “resident” monocytes based on the expression of markers including Gr1/Ly6C and the chemokine receptors CX₃CR1 and CCR2 [18]. Tie2 has been reported to mark a distinct lineage of monocytes, termed Tie2-expressing monocytes (TEMs), that are recruited to the tumor microenvironment where they promote the growth of new blood vessels [19]. In response to stimuli encountered, monocytes may be induced to differentiate into macrophage subtypes including classically activated, “M1” inflammatory macrophages or alternatively activated, “M2” regulatory and wound healing macrophages. There is little doubt, however, that the M1 and M2 classification oversimplifies the extent of macrophage heterogeneity [20,21]. Mature, specialist macrophages found in adult tissues include Kupffer cells of the liver that clear spent erythrocytes from the circulation, microglia in the central

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nervous system that regulate neural development and osteoclasts in the bone marrow important for bone remodeling [1,3].

Recent work has revealed that macrophages play key roles during vascular development; these include directing regression of the hyaloid blood vascular plexus [4,5,22], facilitating the anastomosis of sprouting blood vessels [10] and patterning the retinal vasculature [23,24]. Furthermore, macrophages have been shown to promote neo-vascularization in settings of inflammation and wound repair by producing pro-angiogenic growth factors, chemokines and proteases [19,25-27]. In this review, we focus on the roles that macrophages play in lymphatic vascular growth and development (lymphangiogenesis), both during development and in disease.

Developmental origins of macrophages and lymphatic vessels

The developmental origins of macrophage subsets are being progressively unravelled, with genetic lineage tracing studies providing answers to longstanding questions regarding progenitor cell origin and the differentiation potential of various monocyte/macrophage populations. Current data suggest that maternally derived macrophages are the first to appear in the mouse embryo at approximately embryonic day (E) 7.5 [28]. Subsequently, macrophages originating in the yolk sac (E8) and from definitive hematopoietic progenitor cells arising in the embryonic aorta-gonad-mesonephros (AGM) (E10.0) and foetal liver develop (E10.5-onwards) [28]. After birth, the major site of hematopoiesis is the bone marrow. Recent evidence supports the concept that mature macrophages in the adult are derived from distinct progenitor pools; microglia in the brain appear to be descendents of primitive myeloid progenitors that arise prior to E8 [29], while circulating monocytes that give rise to the majority of tissue macrophages are derived from definitive hematopoietic stem cells.

Construction of the lymphatic vasculature is initiated once the major arteries (dorsal aortae) and veins (cardinal veins) have been established in the embryo, originating from the cardinal veins following the onset of expression of the *Prox1* transcription factor in a polarised population of venous endothelial cells at ~E9.5 [30]. *Prox1*-positive lymphatic endothelial progenitor cells exit the cardinal veins via sprouting and ballooning mechanisms to form lymph sacs and the superficial lymphatic vascular plexus [30,31], a process that is dependent on vascular endothelial growth factor C (VEGF-C) [32]. The initiation of *Prox1* expression in lymphatic endothelial progenitor cells in the cardinal veins signifies lymphatic endothelial cell fate commitment and is dependent on the activity of *Sox18* and *CoupTFII* transcription factors [33,34]. Once established,

the lymphatic vasculature regulates tissue fluid homeostasis, immune cell trafficking and the absorption of dietary fats [35,36].

Work in a variety of vertebrate models has suggested that mesodermal cells, including those of the macrophage lineage, might contribute to genesis of the lymphatic vasculature during development by comprising a pool of lymphatic endothelial progenitor cells [37-40]. These studies have suggested that mesenchymal “lymphangioblasts” expressing both *LEC* (*Prox1/LYVE-1*) and macrophage markers (*LYVE-1*, *CD45*, *F4/80*) integrate to growing lymphatic vessels in the developing embryo [38,40]. In contrast, work from others has concluded that the vast majority of embryonic lymphatic endothelial cells are derived from the venous progenitor pool, with no evidence to support the concept of monocytes or macrophages giving rise to lymphatic endothelial cells [17,41]. These discrepancies may potentially be a result of differences between the vertebrate models employed in these studies, or of the techniques utilized to assess macrophage incorporation into lymphatic vessels (marker expression versus lineage tracing). In fact, embryonic *LYVE-1*-positive macrophages have been found to share an intimate spatial association with embryonic lymphatic vessels (Figure 1) and in some cases appear incorporated into the wall of developing lymphatic vessels, but lineage tracing studies in the mouse embryo have not detected *Prox1* expression in *LYVE-1*-positive, myeloid derived cells [17]. This data suggests that these cells retain a macrophage identity even when resident in lymphatic vessels. Gene expression profiling of embryonic dermal *LYVE-1*-positive macrophages revealed a close resemblance to TEMs [16,17], a population of macrophages that not only play key roles in tumor stimulated angiogenesis, but are important for blood vascular development [10,16,19,42]. These data suggest that *LYVE-1*-positive macrophages may be important for morphogenesis or remodelling of the lymphatic vasculature. Studies utilising real time imaging will be required to discern whether *LYVE-1*-positive macrophages assume localisation in the walls of lymphatic vessels in order to perform an immune surveillance role, whether they transit through the lymphatic endothelium while patrolling the embryo, or whether they fulfil roles important for lymphatic vascular development.

Macrophages as a source of growth and patterning signals in developmental lymphangiogenesis

While it seems unlikely that monocytes/macrophages comprise a lymphatic endothelial progenitor cell pool during embryogenesis, recent work from a number of groups has identified an important role for macrophages in patterning the lymphatic vasculature. Akin to the role

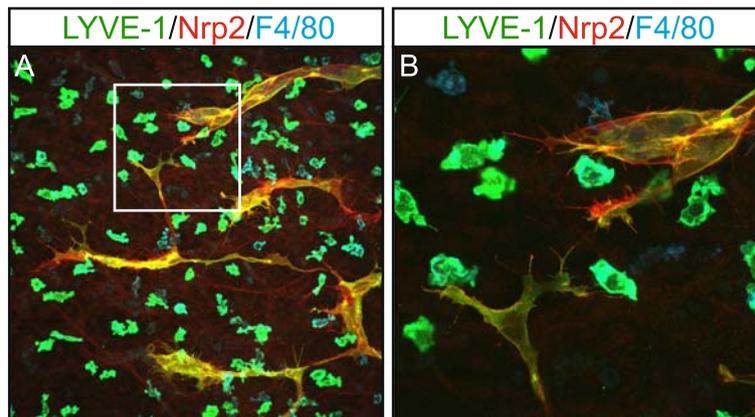


Figure 1 LYVE-1 positive macrophages share an intimate association with growing lymphatic vessels in embryonic mouse skin. Whole mount immunostaining of E14.5 skin illustrating the close association of lymphatic vessel filopodia (Nrp2-positive, LYVE-1-positive) with dermal macrophages (F4/80-positive, LYVE-1-positive). Area boxed in panel **A** is shown at higher magnification in panel **B**.

identified for macrophages in mediating the anastomosis of sprouting blood vessels in the embryonic brain [10] and postnatal retina [23], work from Kubota and colleagues suggested that LYVE-1-positive macrophages regulate density of the lymphatic vasculature in selected postnatal tissues [43]. Analysis of *Csf1^{op/op}* mice that lack a key growth factor for macrophage development, *Csf1*, and are therefore severely depleted of macrophages [44], revealed diminished lymphatic vessel density in the postnatal trachea [43]. Moreover, depletion of macrophages using antibodies to *c-fms* (the receptor for *Csf1*, expressed by macrophages), or the small molecule *c-fms* tyrosine kinase inhibitor Ki20227, between postnatal day (P)8 and P15, resulted in reduced lymphatic vessel branching in the trachea and ears of treated mice. However, these studies found no evidence of perturbed embryonic lymphangiogenesis in *Csf1^{op/op}* mice and lymphatic vascular patterning appeared normal in surviving *Csf1^{op/op}* mice at 3 months of age [43]. These observations suggest a temporal window of macrophage dependence, or potential tissue specific roles, for macrophages in developmental lymphangiogenesis.

Work from Bohmer and colleagues demonstrated that embryonic dermal macrophages expressing the tyrosine kinase *Syk* closely resemble TEMs and express high levels of pro-lymphangiogenic molecules including vascular endothelial growth factor C (VEGF-C), vascular endothelial growth factor D (VEGF-D), fibroblast growth factor 2 (FGF2), matrix metalloprotease 2 (MMP-2) and matrix metalloprotease 9 (MMP-9) [45]. In the absence of *Syk*, an increased number of these pro-lymphangiogenic monocytes/macrophages expressing elevated levels of growth factors and chemokines accumulated in skin and as a result, *Syk^{-/-}* embryos displayed hyperplastic dermal lymphatic vessels. These data suggest that dermal *Syk*-expressing macrophages have the potential to promote

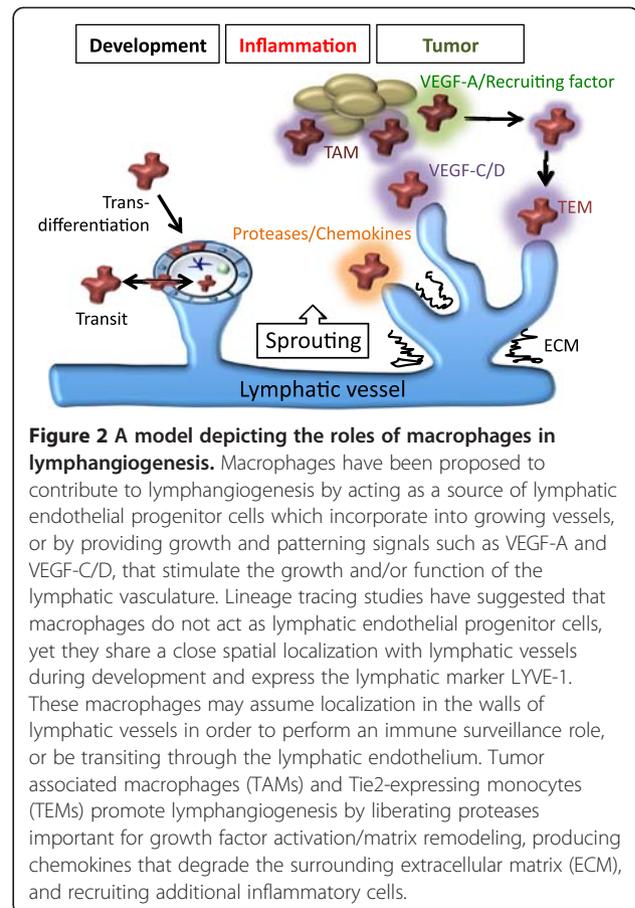
embryonic lymphangiogenesis via the production of a number of pro-lymphangiogenic stimuli. In agreement with these studies, Gordon and colleagues found that primary embryonic dermal macrophages promoted the proliferation of primary embryonic dermal LEC when cultured together *ex vivo* [17]. Intriguingly though, the dermal lymphatic vasculature of embryonic *PLI1^{-/-}* [46] and *Csf1r^{-/-}* [47] macrophage deficient mice was found to be hyperplastic, rather than hypoplastic [17]. These data suggest that while dermal macrophages have the capacity to promote lymphangiogenesis via the production of pro-lymphangiogenic growth factors including VEGF-C and VEGF-D, they may also act to restrain lymphatic endothelial cell proliferation during development. In contrast to the dermal lymphatic vasculature, Gordon and colleagues found that the jugular lymph sacs of embryonic *PLI1^{-/-}* mice were smaller than their wild-type littermates, providing further evidence that macrophages may play distinct, tissue specific roles during developmental lymphangiogenesis [17]. Definitively dissecting the relative contribution of macrophage-derived signals during developmental lymphangiogenesis will rely on the generation of conditional knockout mice in which genes of interest are inactivated in a macrophage selective manner.

Macrophages in inflammation and tumor-stimulated lymphangiogenesis

The roles played by macrophages in pathological lymphangiogenesis are rapidly gaining recognition. The induction of lymphangiogenesis during wound healing and in many pathological settings normally acts to resolve inflammation and edema; in models of infection and acute inflammation, increased lymphangiogenesis has been associated with elevated lymph flow, enhanced immune cell trafficking to draining lymph nodes and the resolution of tissue inflammation and edema [48-50].

While this is advantageous in the majority of settings, lymphangiogenesis induced following organ transplantation has been shown to aid alloimmunization and thereby promote the rejection of kidney, corneal and pancreatic islet transplants [51-53]. Understanding how lymphangiogenesis is regulated during inflammation thereby stands to advance the development of new therapeutics able to promote or ablate lymphangiogenesis dependent on the pathological setting.

Macrophages have been demonstrated to drive lymphangiogenesis in models of inflammation including bacterial infection [48,49,54], wound healing [55,56], organ transplant [57,58], rheumatoid arthritis [59], pancreatic islet inflammation/diabetes [60] and atopic dermatitis [61]. Macrophages also appear to be key players in salt-induced hypertension, where macrophage derived VEGF-C is important for inducing lymphangiogenesis as a buffering mechanism to deal with increased interstitial fluid accumulation [62]. At least two mechanisms by which cells of the monocyte/macrophage lineage promote neo-lymphangiogenesis have been proposed; trans-differentiation to lymphatic endothelial cells [55,57,63-65] and production of pro-lymphangiogenic stimuli including VEGF-C, VEGF-D and VEGF-A [48,49,54,66-68] (Figure 2). The relative contribution that macrophages provide to inflammation-stimulated lymphangiogenesis has been illustrated in studies of macrophage deficient mice [43] and by the depletion of macrophages using clodronate liposomes [49,67], c-fms inhibition [43] or VEGF-A inhibition [49,67,69]. VEGF-C and/or VEGF-D appear to be critical for the pro-lymphangiogenic activity of macrophages in the majority of models studied; blockade of VEGF-C and VEGF-D activity using soluble VEGFR-3 or VEGFR-3 neutralising antibodies has been demonstrated to inhibit macrophage driven lymphangiogenesis [48,49,52,68]. Recent work has suggested that the extracellular matrix protein thrombospondin-1 (TSP-1) acts as an endogenous inhibitor of corneal lymphangiogenesis via acting on macrophages; ligation of CD36 on the macrophage cell surface by Tsp-1 was shown to negatively regulate VEGF-C and VEGF-D production. In the absence of Tsp-1, and in mice deficient in CD36, precocious lymphangiogenesis is induced in the cornea [70]. In addition to VEGF family members, macrophages are a source of proteases including MMP-2 and MMP-9 [1,26,45] that promote growth factor activation and matrix remodelling, as well as cytokines and chemokines that recruit additional cells of the immune system. Immune cells including granulocytes, B and T lymphocytes have also been demonstrated to have pro- and anti lymphangiogenic activities [48,71-73] and their capacity to regulate lymphangiogenesis should be taken into account when looking at the "big picture" of inflammation.



Many studies have shown that the growth of lymphatic vessels in the tumor microenvironment facilitates tumor metastasis [74]. Tumor-stimulated lymphangiogenesis often results in the formation of abnormal, leaky lymphatic vessels, a feature that provides metastatic tumor cells with ready access to the lymphatic vasculature [75,76]. Macrophage recruitment to tumors has been demonstrated to promote lymphangiogenesis in a variety of mouse tumor models [43,66,68,77,78] and tumor associated macrophages (TAMs) have been linked with increased peri-tumoral lymphangiogenesis and metastasis in human cancers including breast cancer [79], cervical cancer [66], squamous cell carcinoma [80] and advanced colorectal cancer [81]. In a mouse model of osteosarcoma, inhibition of Csf1 diminished macrophage recruitment to the tumor environment, suppressed tumor angiogenesis and lymphangiogenesis and reduced tumor metastasis [43]. Similarly, in a model of urinary bladder cancer, depletion of TAMs with clodronate liposomes and suppression of lymphangiogenesis with soluble VEGFR-3 inhibited lymphangiogenesis and tumor metastasis [77]. Studies such as these suggest that therapeutics designed to block macrophage influx or inhibit macrophage activity might provide valuable anti-tumor

and anti-metastatic agents. In addition to promoting the growth of new lymphatic vessels in the vicinity of tumors, macrophage derived pro-lymphangiogenic growth factors including VEGF-C and VEGF-D may act downstream of the initial lymphatics on collecting vessels to promote their dilation and capacity for lymph flow, thereby facilitating metastatic tumor cell transport [35,82]. TAMs may also contribute to lymphangiogenesis associated pathologies in addition to aiding metastasis. When a metastatic human ovarian cancer cell line was transplanted into mice, Jeon and colleagues found that tumor progression was associated with the development of chylous ascites due to a profound dysfunctional lymphangiogenic response [68]. Blockade of VEGF-C/-D with soluble VEGFR-3 and of VEGF-A signaling with VEGF-Trap prevented the formation of chylous ascites, implicating macrophage derived VEGF-C/-D in ascites development due to the failure of aberrant lymphatic vessels to mediate fluid clearance from the peritoneal cavity [68]. This study has implications for the treatment of ascites in ovarian cancer patients.

Perspectives and future directions

A growing body of data now cements “regulation of lymphangiogenesis during development and disease” together with the plethora of important roles that macrophages play in tissue morphogenesis, homeostasis, repair and immunity. Many questions remain to be answered before we will completely understand how macrophages regulate lymphangiogenesis. Current topical questions include: What is the relative contribution of macrophage derived VEGF-C and -D to embryonic and inflammation stimulated lymphangiogenesis? What signals in addition to VEGF family members do macrophages provide that control lymphatic vascular growth and morphogenesis? Do different macrophage subtypes differ with respect to their pro- or anti- lymphangiogenic activity? The generation of new experimental tools including genetically modified mice and agents able to specifically track and manipulate macrophage sub-types will be important for advancing these studies. Though the blockade of macrophages and/or macrophage derived factors poses an attractive strategy for anti-inflammatory and anti-tumor therapies, it will first be important to determine the precise roles of these intriguing cells in developmental and pathological lymphangiogenesis.

Competing interests

The authors declare that they have no competing interests.

Author contributions

NH and EG wrote the manuscript and prepared the figures. All authors read and approved the final manuscript.

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