Possible adverse effects of frying with vegetable oils ☆

Carmen Dobarganes[1]* and Gloria Márquez-Ruiz[2]

[1]*Instituto de la Grasa (IG-CSIC), Avenida Padre García Tejero 4, 41012 Sevilla, Spain*
[2]*Instituto de Ciencia y Tecnología de Alimentos y Nutrición (ICTAN-CSIC), José Antonio Novais, 10, 28040 Madrid, Spain*

Abstract

The question of whether heated fats in the diet may be detrimental to health is nowadays of the upmost concern, but finding an answer is not easy and requires careful consideration of different aspects of lipid oxidation. This review is divided into two sections. The first part deals with the nature of the new compounds formed at high temperature in the frying process as well as their occurrence in the diet while the second part focuses on their possible nutritional and physiological effects. Oxidation products present in abused frying fats and oils are the compounds most suspected of impairing the nutritional properties of the oils or involving adverse physiological effects. The recent studies on their health implications include those related to their fate and those focused on their effects in metabolic pathways and the most prevalent diseases.

Key words: Frying: Vegetable oils: Oxidation products: Oxidised triacylglycerols: Triacylglycerol polymers

During frying, fats and oils are heated to high temperatures while exposed to air and moisture, resulting in a complex series of reactions that generates a wide array of new compounds. Identification and quantification of the new compounds, however, continue to be investigated given the high number of compounds formed from each oxidisable substrate[1–3]. There is general agreement that undesirable or potentially harmful compounds can result from frying, although their biological significance and the levels actually formed are far from clear[4].

Among the different methods of cooking, i.e. frying, baking, boiling, grilling, etc., the greatest concern on the possible adverse nutritional effects has been expressed over intermittent or discontinuous frying because the highest degradation levels have been found under these conditions. Conversely, there is less opportunity for significant alteration either in commercial frying operations using continuous frying because of the high turnover with fresh oil and constant protection of the oil surface by steam water from the food, or in pan-frying, in which oils are heated only for short periods of time and rarely reused[5]. In other cooking methods, the oil is not reused and consequently its degradation is limited.

A survey of the literature shows discrepancies among nutritional studies on used frying oils and fats as experimental conditions vary widely. Besides, information on the heated oils tested is generally based exclusively on the duration of heating, temperature selected and oil used[4]. Therefore, insufficient analytical criteria are provided to establish valid relationships between the degradation compounds formed and the effects observed.

The first part of this review summarises the new compounds formed when heating oils at frying temperature with special attention to those with possible adverse effects while the second part concerns the most significant nutritional studies on used frying oils and oxidation compounds.

Formation and nature of new compounds during frying

During heating at high temperature, the fats and oils undergo the most important lipid reactions, i.e. hydrolysis, oxidation

Table 1. Main groups of new compounds formed during frying

| Alteration	Causative agent	Resulting compounds
Hydrolysis	Moisture	NEFA
		Diacylglycerols
		Monoacylglycerols
Oxidation	Air	Oxidised TAG monomers
		Oxidised TAG polymers (dimers and oligomers)
		Volatile compounds (aldehydes, ketones, alcohols, hydrocarbons, etc.)
Thermal alteration	Temperature	Cyclic TAG monomers
		Trans TAG momomers
		Non-oxidised TAG dimers and oligomers

and thermal alteration, resulting in the loss of quality of the frying oil and thereby of the fried food. Hydrolysis occurs due to the moisture from the foodstuff, which involves the breakage of the ester bond, with subsequent release of fatty acids, monoacylglycerols and diacylglycerols. Because of the presence of air and exposure to high temperature, oxidation and thermal alterations take place in the unsaturated fatty acid, through chain reactions of free radicals, leading mainly to modified TAG with at least one of the three fatty acyl chains altered[6]. Table 1 lists the main groups of alteration compounds formed. Fig. 1 shows examples of simplified, representative structures of each group of compounds. All alteration compounds formed are more polar than their parent non-modified TAG, with the exception of cyclic and *trans* TAG, formed in minimal amounts. For this reason, the new compounds formed

are known as polar compounds, and their total amount can be easily determined by adsorption chromatography[7].

Although general changes in the main oil constituents are known, it is not easy to foresee the rate of degradation due to the high number of variables involved in the frying process. Some of them are linked to the process itself, such as temperature, length of heating, continuous or discontinuous heating and turnover period, and others to the food subjected to frying, i.e. lipid composition, main and minor constituents, etc.; or else to the oil used such as unsaturation degree, initial quality, antioxidant content and additives. Consequently, regulations or guidelines have been established in many countries to guarantee high-quality foods. The most extended limitation establishes that the used frying oil has to be replaced when the total content of polar compounds is higher than 25 % expressed on oil weight[8].

Results obtained in many samples have shown that, at the level of used frying oil rejection for human consumption (25 % polar compounds), TAG polymers (the sum of TAG dimers and TAG oligomers) are by far the major compounds, accounting for 12–15 wt% on oil. As for oxidised TAG monomers, levels of 7–10 wt% on oil are normally found. Regarding hydrolytic compounds, variable contents are reported, but these compounds are less important quantitatively[6,9].

From the nutritional point of view, the compounds formed through hydrolysis of TAG are likewise originated in the stage previous to the fat absorption due to the action of pancreatic lipase and hence have no relevance in this context. Similarly, the volatile oxidation compounds have no nutritional interest because, at the high oil temperature of the frying process, they
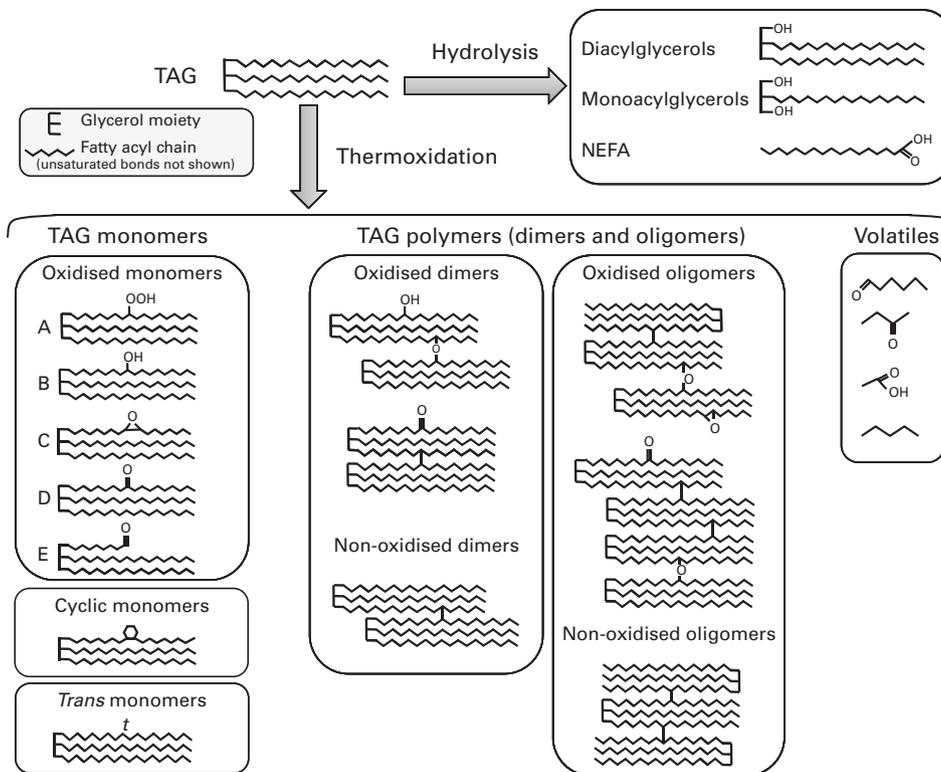
Fig. 1. Schematic representation of the main groups of alteration compounds formed during frying. A, B, C, D and E are simplified structures of hydroperoxy, hydroxy, epoxy, keto and short-chain *n*-oxo fatty acyl groups, respectively, in oxidised TAG.

are released from the oil and are not significant in the fried food. In consequence, to approach the evaluation of the possible adverse effects of heated fats and oils, attention has to be paid to non-volatile compounds coming from oxidation and thermal alterations because these remain in the oil, are absorbed by the food and subsequently ingested. Such compounds can be divided into two groups differing in molecular weight and nutritional significance.

1. Oxidised TAG monomers, characterised by the presence of extra oxygen in at least one of the fatty acyl groups of the TAG molecule (Fig. 1). This group contains different oxygenated groups, i.e. hydroperoxy (A in Fig. 1), hydroxy (B in Fig. 1), epoxy (C in Fig. 1), keto (D in Fig. 1), etc., as well as short-chain *n*-oxo fatty acyl groups (E in Fig. 1) as the main products[10–13]. Their molecular weight is similar to that of the initial or parent non-modified TAG.

2. TAG polymers (dimers and oligomers), the most specific compounds in used frying fats and oils (Fig. 1), formed initially through interaction among TAG monomers. This fraction is of great complexity because the high number of combinations of molecules increases exponentially, from dimers to trimers and higher oligomers[6,14]. Their molecular weights are higher than those of their parent non-modified TAG, and it is very difficult to identify and quantify specific individual compounds.

It is important to remark the quantitative importance of these groups of compounds in the fried foods considering that, at low temperature during storage, foods consumed would contain rarely more than 4–5 % of oxidised TAG because of the clear detection of rancid odour at this oxidation level[15]. Otherwise, in the case of frying fats and oils, the upper limit of 25 % is often considerably surpassed in a significant number of oils and fats from fast food outlets reaching even values about 60 % polar compounds[16,17]. Hence, most of the oxidised fats in foods are expected to come from fats and oils heated at high temperature and more specifically from used frying fats and oils from discontinuous frying.

Health implications of dietary oxidised oils

Physiological and nutritional effects of frying oils have been the subject of intensive investigations since the 1950s. A detailed review of the distinct aspects studied and of the difficulties to reach thorough conclusions has been published recently[4]. The main reason for the difference in the results obtained is the composition of the oils used. On the one hand, those researchers who found high levels of toxicity used abusive heating conditions in an attempt to generate sufficient amounts of degradation products, but the level and structures of the compounds thus formed are not representative of those encountered in oils subjected to normal culinary practices. On the other hand, some researchers applied very soft conditions when heating oils disregarding that the use of good practices in the frying process is

obviously safe. In this short review, only studies applying realistic experimental conditions for the preparation of used frying oils or fried foods, as well as those studies evaluating specific oxidation compounds, are discussed.

Used frying fats and oils

With a few exceptions, classical relevant papers describing works in experimental animals with used frying oils are not alarming and only slight effects on growth rate, liver enlargement and induction of detoxification enzymes involved in the defence mechanisms against *in vivo* lipid peroxidation have been shown[4].

Digestibility has been generally found to decrease in used frying oils or oils heated at frying temperatures[18–23]. Specifically, it has been demonstrated that polymers are poorly hydrolysed by pancreatic lipase. These results were obtained in used frying oils from restaurants and fried food outlets collected by Food Inspection Services that contained from 7·5 to 61·4 % polar compounds. Oxidised TAG monomers were extensively hydrolysed but TAG dimers and higher oligomers gave low hydrolysis values (11–42 %)[20]. Also, it was shown that the hydrolysis of non-oxidised TAG was negatively affected by the presence of large amounts of polar compounds. These results were confirmed by true digestibility measurement through oesophageal probes[21].

With regard to epidemiological studies, researchers have so far not found any direct link between used frying oils and health problems. In fact, fried foods are an important component of the Mediterranean diet, which is strongly associated with a reduced risk of cardiovascular events[24–26]. Nevertheless, a number of studies have been conducted in human subjects to unravel the possible associations between the consumption of fried foods and the incidence of prevalent diseases, mainly cancer[27–30], metabolic syndrome[31–35] and CHD[36–38]. Few studies have found an increased risk of cancer in association with consumption of deep-fried foods, specifically in prostate[27], breast[28], oral/pharyngeal[29], oesophageal[29] and laryngeal[30] cancers. Nevertheless, in all these studies, the intake of frying oil was not defined and the associations reported were not attributed to degradation compounds in the frying oil but to the heterocyclic amines or polycyclic aromatic hydrocarbons formed from meat, or acrylamide formed in carbohydrate-rich foods.

Of special interest are the recent studies regarding the incidence of metabolic syndrome[31–35] and the risk of CHD[36–38]. Variable results were obtained including association with a higher prevalence of arterial hypertension[31], obesity[32,33], or lower HDL-cholesterol levels and a larger waist circumference[34]. Also, a null association with the incidence of metabolic syndrome has been reported in the case of a moderate consumption of fried foods[35]. With regard to the studies evaluating the effect of fried foods on the risk of CHD, either positive[36] or null association[37,38] has been found. Concerning these studies, it is important to remark two aspects. On the one hand, most of the studies have been conducted in Mediterranean countries where olive oil, less prone to degradation than other edible oils, is commonly

used for domestic frying. Therefore, research needs to be extended to other communities consuming preferentially polyunsaturated oils or solid fats. On the other hand, information provided on the oxidation of frying oils or fried foods tested was too scarce. Hence, one important variable contributing to the differences between the results obtained in different studies could be the level of oxidation compounds present in the diet. In fact, a significant association between consumption of fried foods and some of the components of metabolic syndrome was found when the amounts of oxidation compounds in the diet increased. In this respect, two examples are worthy to comment. First, in Soriguer et al.'s[31] study, oil samples were taken from the kitchens of a random subset of 538 participants and 10% of the oils collected contained over 20% polar compounds. A strong association was found between the consumption of such oils and the risk of hypertension, even after inclusion in the models of variables influencing hypertension, such as age, sex and obesity. Also, Sayon-Orea et al.[35], in the SUN cohort study, classified 8289 participants in three groups according to their frequency of fried food consumption and found that those participants who consumed fried foods >4 times/week had a higher risk to develop two out of five components of metabolic syndrome: central adiposity and high blood pressure, compared with those who consumed ≤2 times/week.

Paradoxically, feeding experiments in animals have consistently demonstrated that thermally oxidised oils improves the blood lipid profile, i.e. a reduction in TAG and cholesterol levels in plasma and VLDL, attributed to the activation of hepatic PPARα[39–42]. Even more, it has also been postulated that PPARα activation in the vasculature would inhibit pro-atherogenic events such as monocyte recruitment and smooth muscle cells proliferation and migration[43]. The authors suggest that some of the multiple components found in thermally oxidised oils may exhibit potent regulatory activity on lipid metabolism[44]. Nevertheless, thermally oxidised oils also cause oxidative stress in animals[45–49] probably due to the depletion of antioxidants such as tocopherols in serum and tissues. Hence, the possible atheroprotective effect due to activation of PPARα is probably compromised by the simultaneous induction of intense oxidative stress[42].

Model systems and specific oxidation compounds

A major handicap for researchers focused on individual compounds formed during frying is that few specific compounds have been quantified so far. Alternatively, some investigators make use of model compounds, usually methyl linoleate or linoleic acid, subjected to thermal oxidation under controlled conditions. In this section, the research undergone in relation to the most important groups of compounds, i.e. oxidised TAG monomers and TAG polymers (Fig. 1), is considered separately. Following enzymatic hydrolysis by pancreatic lipase, TAG yield NEFA that are ultimately absorbed. As examples, Fig. 2 represents the hydroxyl fatty acid released from an oxidised TAG monomer and the epoxy fatty acid and dimeric fatty acid released from an oxidised TAG dimer.

Oxidised TAG monomers. Given that the synthesis and analysis of hydroperoxides, the primary oxidation compounds, are well resolved, many studies on their physiological effects have been carried out. Although their occurrence in used frying oils is limited due to their instability at high temperatures, the main effects attributed to hydroperoxides are worth commenting. TAG monohydroperoxides appear to be hydrolysed by pancreatic lipase at similar degree as its parent TAG[50]. Nonetheless, previous reactions in the stomach seem to play an important role[51]. Thus, linoleic acid hydroperoxides administered intragastrically was converted into hydroxyls, epoxyketones, hexanal and 9-oxononanoic acid, time-dependently[52,53]. Other authors have reported that gastrointestinal glutathione peroxidase plays an important role in the mucosal transport and in the conversion of hydroperoxides to more
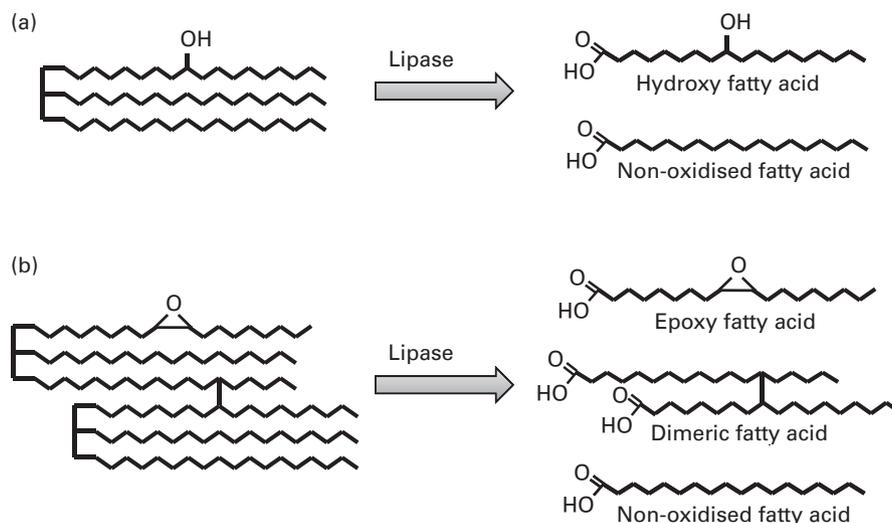
Fig. 2. Schematic representation of fatty acids released from simplified structures of oxidised TAG monomer (a) and oxidised TAG dimer (b) by pancreatic lipase hydrolysis.

Table 2. Summary of the main studies discussed in this review on used frying/thermally oxidised oils, fried products and on model compounds tested in feeding experiments

Substrate	Experimental model	Effect studied	Main results	References
Used frying/thermally oxidised oils	Rats	Digestibility	Decreased digestibility of used frying oil as compared to unused oil	Márquez-Ruiz et al.[18–20], González-Muñoz et al.[21] and Olivero-David et al.[22,23]
			Low lipolysis of TAG polymers	Márquez-Ruiz et al.[20] and González-Muñoz et al.[21]
			TAG polymers decrease lipolysis of non-modified TAG	Márquez-Ruiz et al.[20] and González-Muñoz et al.[21]
		Absorption	Low lymph recovery of non-polar dimeric fatty acids	Perkins & Taubold[78], Combe et al.[79] and Strauss et al.[80]
			Higher absorption of polar dimeric fatty acids as compared to non-polar dimeric fatty acids	Combe et al.[79] and Márquez-Ruiz & Dobarganes[81]
	Rats, mice	Blood lipid profile	Reduction in TAG and cholesterol in plasma and VLDL	Chao et al.[39], Sülzle et al.[40], Luci et al.[41], Ringseis et al.[42], Kammerer et al.[43] and Ringseis & Eder[44]
	Rats, Guinea pigs	Oxidative stress	Induction of antioxidant enzyme systems and reduction of tissue α-tocopherol	Izaki et al.[45], Liu & Huang[46], Liu et al.[47], Ringseis et al.[48] and Olivero-David et al.[49]
Fried foods	Human subjects	Metabolic syndrome	Null association with frequent consumption	Sayon-Orea et al.[35]
			Higher consumption associated with a higher risk of central adiposity and blood pressure	Sayon-Orea et al.[35]
			Higher hypertension for the consumption of oils with >20% polar compounds	Soriguer et al.[31]
			Higher prevalence of obesity	Guallar-Castillón et al.[32] and Sayon-Orea et al.[33]
			Lower HDL-cholesterol and larger waist circumference	Donfrancesco et al.[34]
		CHD	Positive association	Iqbal et al.[36]
			Negative association	Kabagambe et al.[37] and Guallar-Castillon et al.[38]
		Cancer	Not conclusive	Stott-Miller et al.[27], Dai et al.[28], Galeone et al.[29] and Bosetti et al.[30]
TAG hydroperoxides	Rats	Pancreatic lipolysis	High hydrolysis rate of monohydroperoxides	Miyashita et al.[50]
		Gastric digestion	Reduction of hydroperoxides to alcohols and aldehydes	Kanazawa & Ashida[52,53]
Labelled fatty acyls in TAG	Human subjects	Absorption	High absorption of hydroxyl and epoxy fatty acids	Wilson et al.[71,72]
Thermoxidised linoleic acid	Rats	Absorption	Depolymerisation reactions under gastric conditions	Márquez-Ruiz & Dobarganes[81]

Possible adverse effects of frying

S53

stable hydroxy or aldehydic compounds[54–57]. Therefore, dietary hydroperoxides will be largely lost before absorption. In fact, hydroperoxides found in the atherosclerotic process[58–61] are most likely formed endogenously under particular dietary circumstances involving impaired antioxidant status[62]. With regard to studies focused on colorectal cancer, tumour-promoting effects[63,64] and complex metabolic effects of chronic exposure to subtoxic levels have been reported in rats[65,66] and human cells[67,68]. However, other authors have stressed the fact that, with regard to the effect of dietary lipids, dietary components other than oxidised lipids may be responsible for such adverse effects, such as the consumption of red meat and the metabolic activity of the microflora in the colon[69]. Thus, a Hb–Fe-rich diet was found to lead to an increased incidence of colon cancer in rats, attributable to the generation of peroxyl radicals from dietary or membrane lipids of intestinal epithelial cells[70].

With respect to major oxidised compounds in frying oils, high absorption of dietary hydroxy and epoxy fatty acids incorporated in TAG (B and C, respectively, in Fig. 1) was also reported in human subjects, through an excellent approach based on labelled fatty acids included in TAG[71,72]. Later, 13-hydroxylinoleic acid was found to reduce cholesterol content in a macrophage cell line, probably by stimulating apoA-I-dependent cholesterol efflux in a PPAR-dependent manner[73]. Quite in contrast, cytotoxic effects of monoepoxy linoleate or leukotoxin and its corresponding diol, leuko-toxindiol, have been reported[74–77]. However, physiological levels of epoxides in human subjects and significance of dietary epoxides are unknown.

TAG polymers. Polymerisation reactions are accelerated by the high temperatures used in frying but the identification of specific polymeric structures formed is very difficult. Consequently, there are no relevant studies on their effects on metabolic pathways with the exception of those concerning their absorption and digestibility.

Dimeric fatty acids are normally described as non-polar or non-oxidised and polar or oxidised according to the absence or presence, respectively, of one or more oxygenated function either in the dimeric linkage or in the fatty acyl chain. Studies on non-polar dimeric fatty acids (simplified structure in Fig. 2(B)) indicated very low lymph recoveries as values about 1% were found[78,79]. Furthermore, experiments with rats fed labelled non-polar dimers showed recoveries of about 3% radioactivity in urine and CO_2 while approximately 80% of the radioactivity was recovered in the gastrointestinal tract and faeces after 48 h[80]. In contrast, polar dimeric and polymeric fatty acids were comparatively better absorbed[79,81]. This could be in part due to depolymerisation reactions occurring under the strongly acidic conditions in the stomach, as suggested by the presence of non-altered labelled fatty acids in faeces, which were absent in diets[81].

Even though complexity of dimers and polymers is a major handicap for nutritional studies, two key points support further analytical and nutritional research. First, TAG polymers constitute the major fraction in used frying oils and fats, and second, the low absorption of the dimeric and polymeric

fatty acids released does not necessarily means lack of health risk. In fact, it involves increased levels of non-digested, non-absorbed lipids throughout the gastrointestinal tract that might potentially affect epithelial cells and microflora metabolism. In connection with this subject, it has been reported that both unabsorbed lipids and bile acids secreted in response to a high fat intake might injure the intestinal mucose by their detergent activity, and metabolites of bile acids formed by intestinal bacteria (secondary bile acids) act as tumour promoters[82]. Also, an interesting aspect is the potential contribution of intestinal flora to the production of mutagens from the oxidation of faecal lipids and the effect of vitamin E as a chemopreventive agent[83].

Table 2 includes the main studies discussed in this short review focused on used frying/thermally oxidised oils, fried products and on model compounds used in feeding studies.

Conclusions

From the literature reviewed, there is general agreement that a moderate consumption of used frying oils under normal culinary practices is safe, but it is also evident that some compounds formed can impair their nutritional value or be potentially harmful. Therefore, of primary importance is the improvement of the quality control of used frying oils, particularly in the case of fried foods prepared through discontinuous frying process in which used frying oils can reach degradation levels much higher than that established for human consumption.

Overall, the nutritional studies reported on the effects of used frying oils and fats lack the analytical data necessary to establish valid relationships between the alteration compounds present and the effects evaluated on molecular targets, metabolic pathways and chronic diseases. In order to avoid confounding and alarming results, a crucial research assignment is the development of methodologies directed to define the chemical structure and actual levels of the multitude of non-volatile compounds generated from frying and present in the diet. Nowadays, there is no information on the intake and fate of oxidised compounds, which would be essential to guarantee that dietary oxidised lipids are not responsible for any of the adverse physiological effects claimed.

Finally, a specific aspect that remains largely to be explored is the fate of poorly absorbed compounds present in used frying oils, in terms of their interactions with gastrointestinal mucosa and fluids, and with microflora metabolism.

References

1. Choe E & Min DB (2008) Chemistry of deep-fat frying oils. *J Food Sci* **72**, R77–R86.
2. Sánchez-Muniz FJ, Bastida S, Márquez-Ruiz G, *et al.* (2007) Effect of heating and frying on oil and food fatty acids.

In *Fatty Acids in Foods and Their Health Implications*, 3rd ed., pp. 511–542 [CK Chow, editor]. Philadelphia, PA: Taylor & Francis Group.

3. Velasco J, Marmesat S & Dobarganes MC (2009) Chemistry of frying. In *Deep Fat Frying of Foods*, pp. 33–56 [S Sahin and G Sumnu, editors]. Philadelphia, PA: Taylor & Francis Group.

4. Márquez-Ruiz G & Dobarganes MC (2007) Nutritional and physiological effects of used frying fats. In *Deep Frying: Chemistry, Nutrition and Practical Applications*, 2nd ed., pp. 173–203 [MD Erickson, editor]. Champaign, IL: AOCS Press.

5. Márquez-Ruiz G, Ruiz-Méndez MV & Velasco J (2010) Preventing oxidation during frying of foods. In *Oxidation in Foods and Beverages and Antioxidant Applications*, pp. 239–273 [EA Decker, RJ Elias and DJ McClements, editors]. Cambridge, UK: Woodhead Publishing.

6. Dobarganes MC & Márquez-Ruiz G (2007) Formation and analysis of oxidized monomeric, dimeric and higher oligomeric triglycerides. In *Deep Frying: Chemistry, Nutrition and Practical Applications*, 2nd ed., pp. 87–110 [MD Erickson, editor]. Champaign, IL: AOCS Press.

7. International Union of Pure and Applied Chemistry (1992) Standard method 2.507: determination of polar compounds in frying fats. In *Standard Methods for the Analysis of Oils, Fats and Derivatives*, 7th ed. Oxford, UK: Blackwell.

8. Firestone D (2007) Regulation of frying fats and oils. In *Deep Frying: Chemistry, Nutrition and Practical Applications*, 2nd ed., pp. 373–385 [MD Erickson, editor]. Champaign, IL: AOCS Press.

9. Marmesat S, Morales A, Velasco J, *et al.* (2012) Influence of fatty acid composition on formation of new compounds during thermoxidation and frying. *Food Chem* **135**, 2333–2339.

10. Frankel EN (2005) Hydroperoxide decomposition. In *Lipid Oxidation*, 2nd ed., pp. 67–98. Bridgwater: The Oily Press.

11. Marmesat S, Velasco J & Dobarganes MC (2008) Quantitative determination of epoxyacids, ketoacids and hydroxyacids formed in fats and oils at frying temperatures. *J Chromatogr A* **1211**, 129–134.

12. Velasco J, Marmesat M, Bordeaux O, *et al.* (2005) Quantitation of short-chain glycerol-bound compounds in thermoxidized and used frying oils. A monitoring study during thermoxidation of olive and sunflower oils. *J Agric Food Chem* **53**, 4006–4011.

13. Berdeaux O, Fontagné S, Sémon E, *et al.* (2012) A detailed identification study on high-temperature degradation products of oleic and linoleic acid methyl esters by GC–MS and GC–FTIR. *Chem Phys Lipids* **165**, 338–347.

14. Dobarganes MC (1998) Formation and analysis of high molecular-weight compounds in frying fats and oils. *OCL* **5**, 41–47.

15. Dobarganes MC & Márquez Ruiz G (2003) Oxidised fats in foods. *Curr Opin Clin Nutr Metab Care* **6**, 157–163.

16. Saguy S & Dana D (2003) Integrated approach to deep fat frying: engineering, nutrition, health and consumer aspects. *J Food Eng* **56**, 143–152.

17. Marmesat S, Rodrigues E, Velasco J, *et al.* (2007) Used frying fats and oils: comparison of rapid tests based on chemical and physical oil properties. *Int J Food Sci Technol* **42**, 601–608.

18. Márquez-Ruiz G, Pérez-Camino MC & Dobarganes MC (1992) Digestibility of fatty acid monomers, dimers and polymers in the rat. *J Am Oil Chem Soc* **69**, 930–934.

19. Márquez-Ruiz G, Pérez-Camino MC & Dobarganes MC (1993) Evaluation of hydrolysis and absorption of thermally oxidized olive oil in non-absorbed lipids in the rat. *Ann Nutr Metab* **37**, 121–128.

20. Márquez-Ruiz G, Guevel G & Dobarganes MC (1998) Application of chromatographic techniques to evaluate enzymatic hydrolysis of oxidized and polymeric triglycerides by pancreatic lipase 'in vitro'. *J Am Oil Chem Soc* **75**, 119–126.

21. González-Muñoz MJ, Bastida S & Sánchez-Muniz FJ (1998) Short-term *in vivo* digestibility of triglyceride polymers, dimers, and monomers of thermoxidized palm olein used in deep-frying. *J Agric Food Chem* **46**, 5188–5193.

22. Olivero-David R, Sánchez-Muniz FJ, Bastida S, *et al.* (2010) Gastric emptying and short-term digestibility of thermally oxidized sunflower oil used for frying in fasted and non-fasted rats. *J Agric Food Chem* **58**, 9242–9248.

23. Olivero-David R, Paduano A, Fogliano V, *et al.* (2011) Effect of thermally oxidized oil and fasting status on the short-term digestibility of ketolinoleic acids and total oxidized fatty acids in rats. *J Agric Food Chem* **59**, 4684–4691.

24. Estruch R, Ros E, Salas-Salvadó J, *et al.* (2013) Primary prevention of cardiovascular disease with a Mediterranean diet. *N Engl J Med* **368**, 1279–1290.

25. Vallverdú-Queralt A, Rinaldi de Alvarenga JF, Estruch R, *et al.* (2013) Bioactive compounds present in the Mediterranean sofrito. *Food Chem* **141**, 3365–3372.

26. Martínez-González MA & Bes-Rastrollo M (2014) Dietary patterns, Mediterranean diet, and cardiovascular disease. *Curr Opin Lipid* **25**, 20–26.

27. Stott-Miller M, Neuhouser ML & Stanford JL (2013) Consumption of deep-fried foods and risk of prostate cancer. *Prostate* **73**, 960–969.

28. Dai Q, Shu XO, Jin F, *et al.* (2002) Consumption of animal foods, cooking methods, and risk of breast cancer. *Cancer Epidemiol Biomarkers Prev* **11**, 801–808.

29. Galeone C, Pelucchi C, Talamini R, *et al.* (2005) Role of fried foods on oral/pharyngeal and oesophageal cancers. *Br J Cancer* **92**, 2065–2069.

30. Bosetti C, Talamini R, Levi F, *et al.* (2002) Fried foods: a risk factor for laryngeal cancer? *Br J Cancer* **87**, 1230–1233.

31. Soriguer F, Rojo-Martínez G, Dobarganes MC, *et al.* (2003) Hypertension is related to the degradation of dietary frying oils. *Am J Clin Nutr* **78**, 1092–1097.

32. Guallar-Castillón P, Rodríguez-Artalejo F, Lopez-García E, *et al.* (2007) Intake of fried foods is associated with obesity in the cohort of Spanish adults from the European prospective investigation into cancer and nutrition. *Am J Clin Nutr* **86**, 198–205.

33. Sayon-Orea C, Bes-Rastrollo M, Basterra-Gortari FJ, *et al.* (2013) Consumption of fried foods and weight gain in a Mediterranean cohort: the SUN project. *Nutr Metab Cardiovasc Dis* **23**, 144–150.

34. Donfrancesco C, Lo NC, Brignoli O, *et al.* (2008) Italian network for obesity and cardiovascular disease surveillance: a pilot project. *BMC Fam Pract* **9**, 53.

35. Sayon-Orea C, Martínez-González MA, Gea A, *et al.* (2014) Consumption of fried foods and risk of metabolic syndrome: the SUN cohort study. *Clin Nutr* **33**, 545–549.

36. Iqbal R, Anand S, Ounpuu S, *et al.* (2008) Dietary patterns and the risk of acute myocardial infarction in 52 countries: results of the INTERHEART study. *Circulation* **118**, 1929–1937.

37. Kabagambe EK, Baylin A, Siles X, *et al.* (2003) Individual saturated fatty acids and nonfatal acute myocardial infarction in Costa Rica. *Eur J Clin Nutr* **57**, 1447–1457.

38. Guallar-Castillon P, Rodríguez-Artalejo F, Lopez-García E, *et al.* (2012) Consumption of fried foods and risk of coronary heart disease: Spanish cohort of the European prospective investigation into cancer and nutrition study. *BMJ* **344**, e363.

39. Chao PM, Chao CY, Lin FJ, *et al.* (2001) Oxidized frying oil up-regulates hepatic acyl-CoA oxidase and cytochrome 450

4A1 genes in rats and activates PPARα. *J Nutr* **131**, 3166–3174.

40. Sülzle A, Hirche F & Eder K (2004) Thermally oxidized dietary fat upregulates the expression of target genes of PPARα in rat liver. *J Nutr* **134**, 1375–1383.

41. Luci S, Koènig B, Giemsa B, *et al.* (2007) Feeding of a deep-fried fat causes PPAR activation in the liver of pigs as a non-proliferating species. *Br J Nutr* **97**, 872–882.

42. Ringseis E, Gutgesell A, Dathe C, *et al.* (2007) Feeding oxidized fat during pregnancy up-regulates expression of PPARα-responsive genes in the liver of rat foetuses. *Lipids Health Dis* **6**, 6.

43. Kammerer I, Ringseis R & Eder K (2011) Feeding a thermally oxidised fat inhibits atherosclerotic plaque formation in the aortic root of LDL receptor-deficient mice. *Br J Nutr* **105**, 190–199.

44. Ringseis R & Eder K (2011) Regulation of genes involved in lipid metabolism by dietary oxidized fat. *Mol Nutr Food Res* **55**, 109–121.

45. Izaki Y, Yoshikawa S & Uchiyama M (1984) Effect of ingestion of thermally oxidized frying oil on peroxidative criteria in rats. *Lipids* **19**, 324–331.

46. Liu JF & Huang CJ (1995) Tissue α-tocopherol retention in male rats is compromised by feeding diets containing oxidized frying oil. *J Nutr* **125**, 3071–3080.

47. Liu JF, Lee WL & Chan FC (2000) Effects of oxidized frying oil and vitamin C on the hepatic xenobiotic-metabolizing enzyme system of guinea pigs. *J Nutr Sci Vitaminol* **46**, 137–140.

48. Ringseis R, Picwek R & Eder K (2007) Oxidized fat induces oxidative stress but has no effect on NF-κB-mediated proinflammatory gene transcription in porcine intestinal epithelial cells. *Inflamm Res* **56**, 118–125.

49. Olivero-David R, Bastida S, Schultz A, *et al.* (2010) Fasting status and thermally oxidized sunflower oil ingestion affect the intestinal antioxidant enzyme activity and gene expression of male Wistar rats. *J Agric Food Chem* **58**, 2498–2504.

50. Miyashita K, Takagi T & Frankel EN (1990) Preferential hydrolysis of monohydroperoxides of linoleoyl and linolenoyl triacylglycerol by pancreatic lipase. *Biochim Biophys Acta* **1045**, 233–238.

51. Kanner J & Lapidot T (2001) The stomach as a bioreactor: dietary lipid peroxidation in the gastric fluid and the effects of plant-derived antioxidants. *Free Radic Biol Med* **31**, 1388–1395.

52. Kanazawa K & Ashida H (1998) Catabolic fate of dietary trilinoleoylglycerol hydroperoxides in rat gastrointestines. *Biochim Biophys Acta* **1393**, 336–348.

53. Kanazawa K & Ashida H (1998) Dietary hydroperoxides of linoleic acid decompose to aldehydes in stomach before being absorbed into the body. *Biochim Biophys Acta* **1393**, 349–361.

54. Aw TY (1998) Determinants of intestinal detoxication of lipid hydroperoxides. *Free Radic Res* **28**, 637–646.

55. Wingler K, Muller C, Schmehl K, *et al.* (2000) Gastrointestinal glutathione peroxidase prevents transport of lipid hydroperoxides in CaCo-2 cells. *Gastroenterology* **119**, 420–430.

56. Muller C, Friedrich R, Wingler K, *et al.* (2002) Perturbation of lipid metabolism by linoleic acid hydroperoxide in CaCo-2 cells. *Biol Chem* **383**, 637–648.

57. Aw TY (2005) Intestinal glutathione: determinant of mucosal peroxide transport, metabolism, and oxidative susceptibility. *Toxicol Appl Pharmacol* **204**, 320–328.

58. Chisolm GM & Steinberg D (2000) The oxidative modification hypothesis of atherogenesis: an overview. *Free Radic Biol Med* **28**, 1815–1826.

59. Cohn J (2002) Oxidized fat in the diet, postprandial lipaemia and cardiovascular disease. *Curr Opin Lipidol* **13**, 19–24.

60. Jessup W, Krithairides L & Stocker R (2004) Lipid oxidation in atherogenesis: an overview. *Biochem Soc Trans* **32**, 134–138.

61. Stocker R & Keany JF (2004) Role of oxidative modifications in atherosclerosis. *Physiol Rev* **84**, 1381–1478.

62. Duthie GG, Wahle KWJ & James WPT (1989) Oxidants, antioxidants and cardiovascular disease. *Nutr Res Rev* **2**, 51–62.

63. Bull AW, Nigro ND & Marnett LJ (1988) Structural requirements for stimulation of colonic cell proliferation by oxidized fatty acids. *Cancer Res* **48**, 1771–1776.

64. Kanazawa A, Sawa T, Akaike T, *et al.* (2002) Dietary lipid peroxidation products and DNA damage in colon carcinogenesis. *Eur J Lipid Sci Technol* **104**, 439–447.

65. Tsunada S, Iwakiri R, Noda T, *et al.* (2003) Chronic exposure to subtoxic levels of peroxidized lipids suppresses mucosal cell turnover in rat small intestine and reversal by glutathione. *Digest Dis Sci* **48**, 210–222.

66. Tsunada S, Iwakiri R, Fujimoto K, *et al.* (2003) Chronic lipid hydroperoxide stress suppresses mucosal proliferation in rat intestine: potentiation of ornithine decarboxylase activity by epidermal growth factor. *Digest Dis Sci* **48**, 2333–2341.

67. Wang TG, Gotoh Y, Jennings MH, *et al.* (2000) Lipid hydroperoxide-induced apoptosis in human colonic CaCo-2 cells is associated with an early loss of cellular redox balance. *FASEB J* **14**, 1567–1576.

68. Jurek D, Udilova N, Jozkowicz A, *et al.* (2005) Dietary lipid hydroperoxides induce expression of vascular endothelial growth factor (VEGF) in human colorectal tumor cells. *FASEB J* **19**, 97–99.

69. Yang MH & Schaich KM (1996) Factors affecting DNA damage caused by lipid hydroperoxides and aldehydes. *Free Radic Biol Med* **20**, 225–236.

70. Sawa T, Akaike T, Kida K, *et al.* (1998) Lipid peroxyl radicals from oxidized oils and heme−iron: implication of a high fat diet in colon carcinogenesis. *Cancer Epidemiol Biomarkers Prev* **7**, 1007–1012.

71. Wilson R, Fernie CE, Scrimgeour CM, *et al.* (2002) Dietary epoxy fatty acids are absorbed in healthy women. *Eur J Clin Invest* **32**, 79–83.

72. Wilson R, Lyall K, Smyth L, *et al.* (2002) Dietary hydroxy fatty acids are absorbed in humans: implications for the measurement of oxidative stress *in vivo*. *Free Radic Biol Med* **32**, 162–168.

73. Kammerer I, Ringseis R, Biemann R, *et al.* (2011) 13-Hydroxy linoleic acid increases expression of the cholesterol transporters ABCA1, ABCG1 and SR-BI and stimulates apoA-I-dependent cholesterol efflux in RAW264.7 macrophages. *Lipids Health Dis* **10**, 222–231.

74. Mitchell LA, Moran JH & Grant DF (2002) Linoleic acid, *cis*-epoxyoctadecanoic acids, and dihydroxyoctadecadienoic acids are toxic to Sf-21 cells in the absence of albumin. *Toxicol Lett* **126**, 187–196.

75. Moran JH, Mont T, Hendrickson TL, *et al.* (2001) Defining mechanisms of toxicity for linoleic acid monoepoxides and diols in Sf-21 cells. *Chem Res Toxicol* **14**, 431–437.

76. Moran JH, Nowak G & Grant DF (2001) Analysis of the toxic effects of linoleic acid, 12,13-*cis*-epoxyoctadecenoic acid, and 12,13-dihydroxyoctadecenoic acid in rabbit renal cortical mitochondria. *Toxicol Appl Pharmacol* **172**, 150–161.

77. Slim R, Hammock BD, Toborek M, *et al.* (2001) The role of methyl-linoleic acid epoxide and diol metabolites in the amplified toxicity of linoleic acid and polychlorinated biphenyls to vascular endothelial cells. *Toxicol Appl Pharmacol* **171**, 184–193.

78. Perkins EG & Taubold R (1978) Nutritional and metabolic studies of noncyclic dimeric fatty acid methyl esters in the rat. *J Am Oil Chem Soc* **55**, 632–634.

79. Combe N, Constantin MJ & Entressangles B (1981) Lymphatic absorption of nonvolatile oxidation products of heated oils in the rat. *Lipids* **16**, 8–14.

80. Strauss HJ, Piater H & Sterner W (1982) Fütterungsversuche mit dimeren Triglyceriden aus Sojaölraffinat (Feeding Experiments with Dimeric Triglycerides Isolated from Refined Soybean Oil). *Fette Seifen Anstrichm* **84**, 199–203.

81. Márquez-Ruiz G & Dobarganes MC (1995) Assessments on the digestibility of oxidized compounds from [1-^{14}C]-linoleic acid using a combination of chromatographic techniques. *J Chromatogr B* **675**, 1–8.

82. Vonk RJ, Kalivianakis M, Minich DM, et al. (1997) The metabolic importance of unabsorbed dietary lipids in the colon. *Scand J Gastroenterol* **32**, 65–67.

83. Campbell S, Stone WL & Whaley S (2003) Development of gamma (γ)-tocopherol as a colorectal cancer chemopreventive agent. *Crit Rev Oncol Hematol* **47**, 249–259.